

CLASSICAL, ANTIBIOTIC-RESISTANT, EMERGING, AND HYPERVIRULENT STRAINS

Submitted by

ShaimaaAbdElbadi Ali Hassane¹, Dr. Hazem Abdel Wahab Ahmed², Dr. Wafaa Mohammed Khairi³,
Dr.Mohammed Sayed Mohammed⁴, Dr. Noha Anwar Hussein⁵

¹lecturer Assiatant of microbiology ,faculty of medicine,Nahda university

² Professor of Microbiology and Immunology, Faculty of Medicine, Minia University.

³ professor of Microbiology and Immunology, Faculty of Medicine,Minia University.

⁴professor of Microbiology and Immunology,Faculty of Medicine, Minia University.

⁵ professor of Microbiology and Immunology
Faculty of Medicine, Minia University

Klebsiellapneumoniae (*K. pneumoniae*) is a Gram negative, facultative anaerobic bacilli that has great potential for significant morbidity and mortality in acute care settings, particularly in immunocompromised patients (1) .

Moreover, *K. pneumoniae* is one of the most common organisms showing multiple antibiotic resistance worldwide (2) .

These bacteria easily acquire and transfer drug resistance genes through plasmids and transposons . Acquisition of these genes leads to production of β -lactamases of which extended spectrum β -lactamases (ESBLs) are the most common (3) .

ESBLs are capable to hydrolyze extended spectrum penicillins, cephalosporins and monobactams, leaving the carbapenem group of β -lactam antibiotics as the only choice for therapy, hence carbapenem antibiotics are used as a last resort to treat infections caused by these multidrug resistant organisms (4) .

However, there has been emergence of carbapenem resistant *Enterobacteriaceae*, most commonly carbapenem resistant *K. pneumoniae* (CRKP), which have a worldwide prevalence (5), due to high antibiotic use, self-medication by patients and lack of implementation of antibiotic policies in hospitals (6).

Mechanisms described for carbapenem resistance include, production of different classes of carbapenemase, hyperproduction of AmpC β -lactamase with an outer membrane porin mutation, and production of ESBL with a porin mutation or drug efflux. Production of carbapenemases is the most commonly reported mechanism of carbapenem resistance in *K. pneumoniae* (7).

Carbapenemases are β -lactamase enzymes that are capable of hydrolyzing all beta-lactam antibiotics, including monobactams, extended spectrum cephalosporins and carbapenem [8].

The most common carbapenemases include Verona integronmetallo-beta-lactamases types (VIM), imipenemase (IMP) types, *Klebsiellapneumoniae* carbapenemase (KPC), oxacillinase-48 (OXA-48), and New Delhi metallo-beta-lactamase-1 (NDM-1), encoded by carbapenem resistance determining genes *bla*VIM, *bla*IMP, *bla*KPC, *bla*OXA-48-like, and *bla*NDM, respectively (9).

Phenotypic assays are used to identify activity of carbapenemase while molecular assays have developed to identify carbapenemase encoding genes (8) .

Analysis of hospital surveillance data by the Center for Disease Control and Prevention (CDC) suggested that 8% of all *Klebsiella* isolates are carbapenem resistant (10).

Other studies showed that it accounted for 5–24% of *Klebsiella* isolates identified in hospitalized patients (11).

In Egypt, carbapenem resistance is emerging and alarming, one study reported that carbapenem resistance was detected in 44.3% of *K. pneumoniae* isolates in Suez Canal university hospitals (12).

The detection of carbapenem resistance is essential for the proper choice of antibiotic therapy as well as infection control measures to prevent dissemination of resistant strains in hospital settings (13).

Therefore we set out this study to determine the prevalence of carbapenem resistance and carbapenemase encoding genes among clinical *K. pneumoniae* isolates obtained from patients at intensive care units (ICUs) of Minia university hospitals (MUHs), taking in consideration that carbapenems are frequently used as an empiric therapy in ICUs at our institution(14).

Klebsiellapneumoniae has recently gained notoriety as an infectious agent due to a rise in the number of severe infections and the increasing scarcity of effective treatments (15).

These concerning circumstances have arisen due to the emergence of *K. pneumoniae* strains that have acquired additional genetic traits and become either hypervirulent (HV) or antibiotic resistant. *K. pneumoniae* was first isolated in the late 19th century and was initially known as Friedlander's bacterium (16).

It is a Gram-negative, encapsulated, nonmotile bacterium that resides in the environment, including in soil and surface waters and on medical devices (17).

studied in depth and are important in one or more types of infections as well as on additional *K. pneumoniae* virulence factors that have been identified in recent work. To understand the roles of these factors in the context of *K. pneumoniae* infections, we first review the different types of *K. pneumoniae* strains that are now causing significant disease, the types of diseases caused by these *K. pneumoniae* strains, and the host factors that *K. pneumoniae* encounters when establishing an infection (18).

Over the last few decades, there has been a concerning rise in the acquisition of resistance to a wide range of antibiotics by strains derived from "classical" *K. pneumoniae*. As a consequence of this antibiotic resistance, simple infections such as urinary tract infections (UTIs) have become resistant to treatment, and more serious infections such as pneumonias and bacteremias have become increasingly life-threatening (19).

Two major types of antibiotic resistance have been commonly observed in *K. pneumoniae*. One mechanism involves the expression of extended-spectrum lactamases (ESBLs), which render bacteria resistant to cephalosporins and monobactams. The other mechanism of resistance, which is even more troubling, is the expression of carbapenemases by *K. pneumoniae*, which renders bacteria resistant to almost all available lactams, including the carbapenems (20).

The first case of *K. pneumoniae* expressing a carbapenemase was identified in North Carolina in 1996, and thus, this type of carbapenemase is called KPC (21).

Additional carbapenemases, such as MBL, NDM-1, IMP, and VIM, have since been found in *K. pneumoniae* strains (22).

Notably, all of these carbapenemases, including KPC, have been found in other bacteria, and collectively, they contribute to the worldwide occurrence of carbapenem-resistant bacteria (23).

Regardless of the type of carbapenemase that they carry, carbapenem-resistant *K. pneumoniae* isolates are termed CRE, for carbapenem-resistant Enterobacteriaceae. Due to a lack of available effective treatments, *K. pneumoniae* infections caused by ESBL-producing and carbapenem-resistant bacteria have significantly higher rates of morbidity and mortality than infections with nonresistant bacteria (24).

Work reported by the CDC in 2013 demonstrates the frequency and severity of infections with these strains based on a 2011 survey of 183 hospitals in the United States. ESBL-producing strains caused 23% of nosocomial *K. pneumoniae* infections, equaling 17,000 infections, and resulted in 1,100 deaths (25).

Meanwhile, carbapenem-resistant *K. pneumoniae*. Typically, classical *K. pneumoniae* strains cause serious infections, such as pneumonia, bacteremia, or meningitis, when infecting immunocompromised individuals, including people suffering from diabetes or malignancies (26).

The carriage and expression of drug resistance do not enhance the virulence of *K. pneumoniae* strains despite making them more difficult to treat. However, since the 1980s, strains of *K. pneumoniae* that can cause serious infections in otherwise healthy individuals have also gained traction in the human population (27).

Typically, classical *K. pneumoniae* strains cause serious infections, such as pneumonia, bacteremia, or meningitis, when infecting immunocompromised individuals, including people suffering from diabetes or malignancies (28).

The carriage and expression of drug resistance do not enhance the virulence of *K. pneumoniae* strains despite making them more difficult to treat. However, since the 1980s, strains of *K. pneumoniae* that can cause serious infections in otherwise healthy individuals have also gained traction in the human population (29).

These strains are considered HV compared to classical *K. pneumoniae* strains due to their ability to infect both healthy and immunocompromised populations and because of the increased tendency of these infections to be invasive; i.e., they can establish infection in the liver (30).

This additional virulence correlates with the acquisition of a 200- to 220-kb plasmid containing genes that enhance capsule production as well as encode siderophores (31).

These and other bacterial factors that contribute to the hypervirulence of these strains are discussed in detail below (32).

References

- (1) Hsieh PF, Lin TL, Yang FL, Wu MC, Pan YJ, et al. (2012) Lipopolysaccharide O1 antigen contributes to the virulence in *Klebsiella pneumoniae* causing pyogenic liver abscess. PLoS One 7: e33155.**
- (2) Mansury D, Motamedifar M, Sarvari J, Shirazi B, Khaledi A. Antibiotic susceptibility pattern and identification of extended spectrum betalactamases (ESBLs) in clinical isolates of *Klebsiella pneumoniae* from Shiraz, Iran. Iran J Microbiol 2016;8:55e61.**
- (3) Giani T, Pini B, Arena F, Conte V, Bracco S, Migliavacca R, et al. Epidemic diffusion of KPC carbapenemase-producing *Klebsiella pneumoniae* in Italy: results of the first countrywide survey, 15 May-30 June 2011. Euro Surveill 2013;18.**
- (4) Nobari S, Shahcheraghi F, Rahmati Ghezeli F, Valizadeh B. Molecular characterization of carbapenem-resistant strains of *Klebsiella pneumoniae* isolated from Iranian patients: first identification of blaKPC gene in Iran. Microb Drug Resist 2014;20:285e93.**

- (5) **Li B, Sun JY, Liu QZ, Han LZ, Huang XH, Ni YX.** First report of *Klebsiella oxytoca* strain coproducing KPC-2 and IMP-8 carbapenemases. *Antimicrob Agents Chemother* **2011;55:2937e41.**
- (6) **Peleg AY, Franklin C, Bell JM, Spelman DW.** Dissemination of the metallo-beta-lactamase gene blaIMP-4 among gram-negative pathogens in a clinical setting in Australia. *Clin Infect Dis* **2005;41:1549e56.**
- (7) **Potron A, Poirel L, Rondinaud E, Nordmann P.** Intercontinental spread of OXA-48 beta-lactamase-producing Enterobacteriaceae over a 11-year period, 2001e**2011.** *Euro Surveill* 2013;1.
- (8) **Wayne P. Performance standards** for antimicrobial susceptibility testing. Clinical and Laboratory Standards Institute (CLSI); 2015. 25th Informational Supplement.
- (9) **Doyle D, Peirano G, Lascols C, Lloyd T, Church DL, Pitout JD.** Laboratory detection of Enterobacteriaceae that produce carbapenemases. *J Clin Microbiol* 2012;50:3877e80.
- (10) **Rasheed JK, Kitchel B, Zhu W, Anderson KF, Clark NC, Ferraro MJ, et al.** New Delhi metallo-beta-lactamase-producing Enterobacteriaceae, United States. *Emerg Infect Dis* **2013;19:870e8.**
- (11) **Rasheed JK, Kitchel B, Zhu W, Anderson KF, Clark NC, Ferraro MJ, et al.** New Delhi metallo-beta-lactamase-producing Enterobacteriaceae, United States. *Emerg Infect Dis* **2013;19:870e8.**
- (12) **Shahcheraghi F, Nobari S, Rahmati Ghezeli F, Nasiri S, Owlia P, Nikbin VS, et al.** First report of New Delhi metallo-beta-lactamase-1-producing *Klebsiella pneumoniae* in Iran. *Microb Drug Resist* **2013;19:30e6.**
- (13) **Fazeli H, Norouzi-Barough M, Ahadi AM, Shokri D, Solgi H.** Detection of New Delhi metallo-beta-lactamase-1 (NDM-1) in carbapenem-resistant *Klebsiella pneumoniae* isolated from a university hospital in Iran. *Hippokratia* **2015;19:205e9.**
- (14) **Zowawi HM, Sartor AL, Balkhy HH, Walsh TR, Al Johani SM, AlJindan RY, et al.** Molecular characterization of carbapenemase-producing *Escherichia coli* and *Klebsiella pneumoniae* in the countries of the Gulf cooperation council: dominance of OXA-48 and NDM producers. *Antimicrob Agents Chemother* **2014;58:3085e90.**
- (15) **Chen Y, Tenover FC, Koehler TM.** Beta-lactamase gene expression in a penicillin-resistant *Bacillus anthracis* strain. *Antimicrob Agents Chemother* 2004;48:4873e7.
- (16) **Abdel-Wahab, F., Ghoneim, M., Khashaba, M., El-Gilany, A. H. & Abdel-Hady, D (2013).** Nosocomial infection surveillance in an Egyptian neonatal intensive care unit. *J Hosp Infect* 83, 196–199 .
- (17) **Cao, X, Yean CY, Rahman RA (2014)** .Molecular characterization of clinical multidrug-resistant *Klebsiella pneumoniae* isolates. *Annals of Clinical Microbiology and Antimicrobials* 13, 16–16, doi: 10.1186/1476-0711-13-16 .

(18) Clinical Laboratory Standards Institute (2017): Performance standards for antimicrobial susceptibility testing, 16th informational supplements, CLSI document M100-S16. Wayne, PA: Clinical Laboratory Standards Institute.

(19)Daef, E. A. &Elsherbiny, N. M (2012).Clinical and Microbiological Profile of Nosocomial Infections in Adult Intensive Care Units at Assiut University Hospitals, Egypt. *Journal of American Science* 8, 1239–1250 .

(20) Hamzan NI, Yean CY, Rahman RA, Hasan H, Rahman ZA (2015).Detection of blaIMP4 and blaNDM1 harboring Klebsiellapneumoniae isolates in a university hospital in Malaysia.*Emerg Health Threats J* 2015;8:26011.

(21)Rasheed JK, Kitchel B, ZhuW, Anderson KF, Clark NC, Ferraro MJ(2013) .New Delhi metallo-beta-lactamase-producing Enterobacteriaceae, UnitedStates.*Emerg Infect Dis* **2013**;19:870e8.

(22)Jamal WY, Albert MJ, Rotimi VO (2016). High prevalence of New Delhi metallobeta- lactamase-1 (NDM-1) producers among carbapenem-resistant Enterobacteriaceae in Kuwait.*PLoS One* **2016**;11, e0152638.

(23)Mansury D, Motamedifar M, Sarvari J, Shirazi B, Khaledi A(2016). Antibioticsusceptibility pattern and identification of extended spectrum betalactamases (ESBLs) in clinical isolates of Klebsiellapneumoniae fromShiraz, Iran. *Iran J Microbiol*.

(24)Nobari S, Shahcheraghi F, RahmatiGhezelgeh F, Valizadeh B (2014). Molecular characterization of carbapenem-resistant strains of Klebsiellapneumoniae isolated from Iranian patients: first identification of blaKPC gene in Iran. *Microb Drug Resist*.

(25) Wayne P (2015).Performance standards for antimicrobial susceptibility testing.Clinical and Laboratory Standards Institute (CLSI); InformationalSupplement.

(26)Rock C, Thom KA, Masnick M, Johnson JK, Harris AD, Morgan DJ. 2014. Frequency of Klebsiellapneumoniaecarbapenemase (KPC)- producing and non-KPC-producing Klebsiella species contamination ofhealthcare workers and the environment. *Infect Control HospEpidemiol*35:426–429. <http://dx.doi.org/10.1086/675598>.

(27)Kuehn BM. 2013. “Nightmare” bacteria on the rise in US hospitals, long-term care facilities. *JAMA* 309:1573–1574. <http://dx.doi.org/10.1001/jama.2013.2922>.

(28) CDC. 2015. CDC works 24/7 to protect US from health, safety and security threats. CDC, Atlanta, GA .

(29)Yigit H, Queenan AM, Anderson GJ, Domenech-Sanchez A, BiddleJW, Steward CD, Alberti S, Bush K, Tenover FC. 2001. Novel carbapenem-hydrolyzing beta-lactamase, KPC-1, from a carbapenem-resistantstrain of Klebsiellapneumoniae. *Antimicrob Agents Chemother* 45:1151–1161. <http://dx.doi.org/10.1128/AAC.45.4.1151-1161.2001>.

(30) Pitout JD, Nordmann P, Poirel L. 2015. Carbapenemase-producingKlebsiellapneumoniae, a key pathogen set for global nosocomial dominance. *Antimicrob Agents Chemother* 59:5873–5884. <http://dx.doi.org/10.1128/AAC.01019-15>.

(31) Iredell J, Brown J, Tagg K. 2016. Antibiotic resistance in Enterobacteriaceae: mechanisms and clinical implications. *BMJ* 352:h6420. <http://dx.doi.org/10.1136/bmj.h6420>.

(32)CDC. 2013. Vital signs: carbapenem-resistant Enterobacteriaceae. *MMWR Morb Mortal Wkly Rep* 62:165–170.

