

Prevalence and Antibiogram of MERSA Isolated from Sample in Tertiary Care Center in Central India

Taruna Singh¹, Shailja Tiwari², Vijay Tiwari³, Balvir Singh⁴, Pramod Khushwaha⁵ & Ashutosh Garg⁶

¹Assistant Professor Department of Microbiology, SSMC, Rewa, M.P.

²Assistant Professor Department of Physiology, SSMC, Rewa, M.P.

³Assistant Professor Department of Microbiology, SSMC, Rewa, M.P.

⁴Assistant Professor Department of Pharmacology, SSMC, Rewa, M.P.

⁵Assistant Professor Department of Microbiology SSMC, Rewa, M.P.

⁶Demonstrator Department of Microbiology SSMC, Rewa, M.P.

First Author: Taruna Singh

Corresponding Author: Balvir Singh

Abstract

Staphylococcus aureus is an important human pathogen causing various kinds of infections ranging from minor skin diseases to life-threatening endocarditis. It has acquired resistance to previously effective antimicrobials including the methicillin. Methicillin resistant *Staphylococcus aureus* (MRSA) is a prototype of resistant bacteria associated with greater lengths of hospital stay, higher mortality, increased costs and more troublesome to patient when compared with methicillin sensitive *Staphylococcus aureus* (MSSA). The present study was a prospective study conducted for a period 1 year 8 month (from Nov 2011- June 2013) all staphylococcal isolate in Microbiology Laboratory, Shri Aurbindo Medical college & P.G. institute indore india. Gram staining of each specimen (except blood) was performed and findings noted. Each specimen was cultured on Blood agar and MacConkey's agar aerobically incubated overnight at 37°C. Staphylococcal isolate were identified by phenotyping methods like Gram stain, catalase test, slide and tube coagulase test growth on manitol salt agar, VP test Phosphates test and bacitracin susceptibility test. The antimicrobial susceptibility testing was performed by Kirby-Bauer disc diffusion method as per CLSI guidelines. In the present study out of 649 *Staphylococcus* 245 (37.8%) strains were methicillin resistant *Staphylococcus aureus* (MRSA) and 404 (62.2%) were methicillin sensitive *Staphylococcus aureus* (MSSA). Out of Total 649 *S. aureus*, 245 were MRSA. Highest isolation of MRSA was found in Tracheal aspirate (73.3%), blood culture (70.97%) followed by suction tip (69.2%), Sputum (63.2%), body fluids/CSF (52.9%), tissue (50%), broncho alveolar lavage (46.2%), pus & wound swab (32.6%), vaginal swab (26.7%) and urine (21.7%). MRSA strain were 100% resistant to penicillin followed by erythromycin (96.7%), clindamycin (88.6%), ciprofloxacin (85.3%), Cotrimoxazole (70.6%), rifampicin (42.9%) and Nitrofurantoin (66.7%).

Key words: MERSA, Antibiogram, prevalence, India

Study Design: Prospective Observational Study.

1. INTRODUCTION

Staphylococcus aureus is an important human pathogen causing various kinds of infections ranging from minor skin diseases to life-threatening endocarditis. It has acquired resistance to previously effective antimicrobials including the methicillin.¹ Methicillin resistant *Staphylococcus aureus* (MRSA) is a prototype of resistant bacteria associated with greater lengths of hospital stay, higher mortality, increased costs and more troublesome to patient when compared with methicillin sensitive *Staphylococcus aureus* (MSSA)²⁻⁶

β -Lactams are considered as the first-choice antibiotics to treat *Staphylococcal* infections. Currently, the increasing resistance against antibacterial drugs is a major public health concern and one of the biggest challenges faced by physicians. In *S. aureus*, resistance to methicillin occurs because of variations in the alteration of constitutive penicillin-binding proteins (PBPs) or expression of the *mecA*.^{7,8} This has triggered alarm of medical community as *S. aureus* causing life-threatening infections in hospitals and community. Now MRSA is one of the most common causes of nosocomial infections accounting for 40% to 70% of *S. aureus* infections in intensive care units (ICUs).⁹

The Clinical and Laboratory Standards Institute (CLSI)¹¹ has recommended the cefoxitin disk test for prediction of *mecA*-mediated resistance. Detection of *mecA* gene by Polymerase chain reaction (PCR) is considered to be gold standards but it is not yet available in all clinical laboratories. Therefore phenotypic methods still remain a method of choice in resource limited settings.¹⁰ The present study was done to characterize and to determine the prevalence of MRSA isolates obtained from clinical specimens in a health care setup.

2. MATERIAL AND METHODS

The present study was a prospective study conducted for a period 1 year 8 month (from Nov 2011- June 2013) all staphylococcal isolate in Microbiology Laboratory, Shri Aurbindo Medical College & P.G. Institute Indore India. The specimens like pus and wound swab, blood, bronchoalveolar lavage, sputum, tracheal aspirate, suction tip, urine, vaginal swab, tissue, body fluids/ CSF submitted to microbiology laboratory were processed as per standard procedures.¹² Gram staining of each specimen (except blood) was performed and findings noted. Each specimen was cultured on Blood agar and MacConkey's agar aerobically incubated overnight at 37°C. *Staphylococcal* isolate were identified by phenotyping methods like Gram stain, catalase test, slide and tube coagulase test growth on manitol salt agar, VP test phosphatase test and bacitracin susceptibility test.

All *Staphylococcus aureus* isolates were tested for methicillin resistance by Cefoxitin (30 μ g) disc diffusion test (CDD), Chromogenic agar method. E test to know minimum inhibitory concentration (MIC) of Oxacillin and Vancomycin, Latex agglutination test based on detection of PBP2a in 100 strains of *S. aureus*. Polymerase chain reaction (PCR) for detection of *mecA* gene in 100 cefoxitin resistant strains of MRSA. The antimicrobial susceptibility testing was performed by Kirby-Bauer disc diffusion method as per CLSI

guidelines.¹³ A lawn culture of the test strain was prepared on Mueller- Hinton agar (MHA), plate. With all aseptic precautions the antibiotics discs were placed on Mueller Hinton agar plate and incubated at 37°c overnight. Following antibiotics discs (Hi Media Pvt. Ltd, Mumbai) were used according to CLSI guidelines 2010 and ATCC 25923 (MSSA), 43300(MRSA)strain was used as a control strain.¹³

3. RESULT

Table 01: Prevalence of methicillin resistant Staphylococcus aureus (MRSA) and methicillin sensitive Staphylococcus aureus (MSSA) strains.

S. aureus	MRSA		MSSA	
	Number	Percentage	Number	Percentage
649	245	37.8%	404	62.2%

In the present study of 649 Staphylococcus 245 (37.8%) strains were methicillin resistant Staphylococcus aureus (MRSA) and 404 (62.2%) were, methicillin sensitive Staphylococcus aureus (MSSA).

Figure 01.:Prevalence of methicillin resistant Staphylococcus aureus (MRSA) and methicillin sensitive Staphylococcus aureus (MSSA) strains

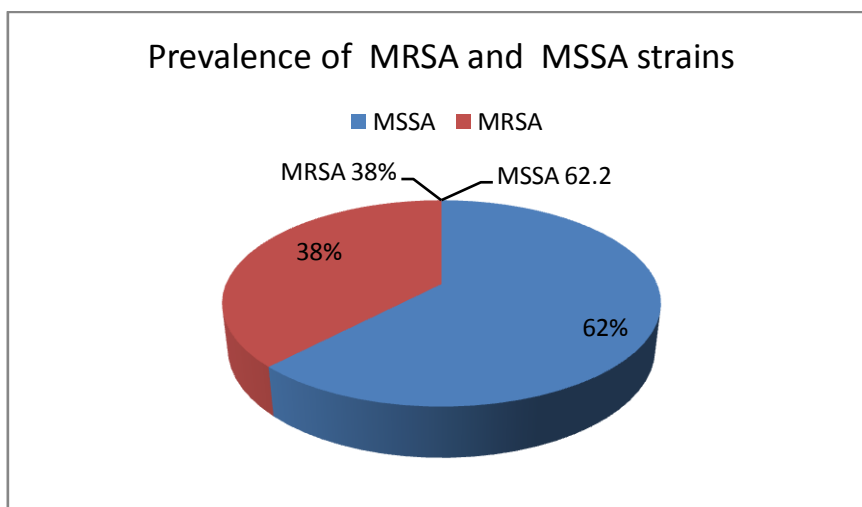


Table 02: Isolation of methicillin resistant Staphylococcus aureus (MRSA) from different clinical specimens (n= 649)

Specimens	S. aureus N=649	MRSA	
		N=245	Percentage
Pus & wound swab	484	158	32.6
Blood	31	22	70.97
BAL	26	12	46.2
Urine	23	5	21.7
Sputum	19	12	63.2

Vaginal swab	15	4	26.7
Tracheal asp.	15	11	73.3
Suction tip	13	9	69.2
Tissue	6	3	50
Body Fluid/CSF	17	9	52.9
TOTAL	649	245	37.8

Out of Total 649 S.aureus, 245 were MRSA. Highest isolation of MRSA was found in Tracheal aspirate (73.3%) , blood culture(70.97%) followed by suction tip (69.2%) , Sputum (63.2%) , body fluids/CSF(52.9%) , tissue(50%) , broncho alveolar lavage(46.2%) , pus & wound swab(32.6%) , vaginal swab (26.7%)and urine(21.7%).

Table 03: Antibiotic susceptibility profile of methicillin resistant Staphylococcus aureus (MRSA) (n=245)

Antibiotics	Methicillin Resistant Staphylococcus aureus (MRSA)			
	Sensitive		Resistant	
	Numbers	Percentage	Numbers	Percentage
Penicillin	0	0	245	100
Erythromycin	8	3.3	237	96.7
Clindamycin	28	11.4	217	88.6
Ciprofloxacin	36	14.7	209	85.3
Cotrimoxazole	72	29.4	173	70.6
Gentamycin	135	55.1	110	44.9
Rifampicin	140	57.1	105	42.9
Tetracycline	171	69.8	74	30.2
Chloramphenicol	204	83.3	41	16.7
Linezolid	245	100	0	0
Vancomycin	245	100	0	0
Nitrofurantoin (Urine samples =5) MRSA= 5 MSSA= 18	3	60	2	40

MRSA strain were 100 % resistant to penicillin followed by erythromycin(96.7%) , clindamycin (88.6%) , ciprofloxacin (85.3%) , Cotrimoxazole (70.6%) rifampicin (42.9%) and Nitrofurantoin (66.7%).

4. DISCUSSION

The distribution of MRSA varies according to factors such as population, areas studied , use of different culture techniques and different interpretation of guidelines.^{14,15}In the present study a total of 649 Staphylococcus aureus strains were isolated from different clinical samples like pus and wound swab, blood, sputum, bronchoalveolar lavage, tracheal aspirate, body fluids , CSF etc. We also evaluated the antibiotic susceptibility

pattern of these isolates and various phenotypic methods for detection of MRSA. The study was also conducted to detect *mecA* gene by PCR in 100 cefoxitin resistant MRSA strains.

Methicillin resistance in *S. aureus* restricts therapeutic options for clinical isolates and the incidence of MRSA is escalating in India. Antibiogram analysis has been found to be a good epidemiological marker for MRSA phenotyping. Currently, majority of *S. aureus* strains are beta-lactamase producer, hence resistant to penicillin. In our study all 245 MRSA and out of 404, 99.3% MSSA isolates were resistant to penicillin while all MRSA and MSSA were sensitive to linezolid and vancomycin similar to **Loveena Oberoi et al 2013**¹⁶ and **Sharma M et al 2013**¹⁷ reported 100% of their MRSA isolates were resistance to penicillin and sensitive to Vancomycin and Linezolid.

Sashirekha B et al 2012¹⁴ reported that 7.14% of isolates were resistant to vancomycin.

In our study MRSA isolates showed higher resistance to erythromycin (96.7%), Clindamycin (88.6%), ciprofloxacin (85.3%), and cotrimoxazole (70.6%). Similar resistance has also been reported by Sharma M 2013¹⁷

Sharma JB et al 2010¹⁸ found in their study that 100% of MRSA isolates were resistant to erythromycin, trimethoprim, ciprofloxacin, gentamycin and tobramycin; 85% to clindamycin and 96% were also resistant to tetracycline, co-trimoxazole and amikacin. **Tiwari et al (2008)**¹⁹ reported 76.1% tetracycline resistance in MRSA.

In our study 83.3% MRSA were sensitive to Chloramphenicol, similar results (80%) have been reported by **Baddour MM et al. (2007)**²⁰ and 73.80% by **Sashirekha B et al (2012)**¹⁴. Thus, our study reported that antibiotics such as Chloramphenicol and Tetracycline can be promising if susceptibility testing is done, reserving vancomycin for life threatening infection. Considerable variations were found in the resistance profiles among MRSA isolated from different countries. The high level resistance of the isolates in the present study to

penicillin, cotrimoxazole, ciprofloxacin, and rifampicin can be attributed to the fact that these antibiotics are frequently used in treatment of common infections. Monotherapy is associated with increased resistance as compared to combination therapy. Therefore, combination treatment is advisable and proven to be beneficial in treatment and eradication of MRSA strains.¹⁴

An urgent need exists for more appropriate selection and use of antimicrobial drugs in the developed as well as in developing countries. The focus in developing countries should be on the availability of safe and effective drugs and on the enforcement of more responsible national drug policies. These issues must be addressed by the collective action of Governments, the pharmaceutical industry, health care providers, and consumers. The developed countries have an important stake in the ways in which antibiotics are used in developing countries because resistant microorganisms do not recognize national boundaries.

5. REFERENCES

- [1] Chambers HF (2001) The changing epidemiology of *Staphylococcus aureus*? *Emerg Infect Dis* 7: 178–1822.
- [2] Akcam F Z, Tinaz GB, Kaya o, Tigali A, Turu E, and Hosoglu S. Evaluation of methicillin resistance by cefoxitin disk diffusion and PBP2a latex agglutination test in *mecA* – positive *Staphylococcus aureus*, and comparison of *mecA* with *femA*, *femB*, *femX* positivities. *Microbiological Research*, 2009; 164: 400-403
- [3] Loomba P.S, Taneja J, Mishra B. Methicillin and Vancomycin resistant *S. aureus* in hospitalized patients. *J Global Infect Dis* 2010; 2(3): 275-283

- [4] Francis, S.T., Rawal, S., Roberts, H., Riley, P., Planche, T., and Kennea, N.L., Detection of methicillin-resistant staphylococcus aureus (MRSA) colonization in newborn infants using real-time polymerase chain reaction (PCR). *Acta Pædiatrica* 2010; 99 :1691–1694
- [5] Gardam MA. Is methicillin-resistant Staphylococcus aureus an emerging community pathogen? A review of the literature. *Can J Infect Dis* 2000;11(4):202-211.
- [6] Lawes T, Edwards B, López-Lozano J-M, Gould, I., Trends in Staphylococcus aureus bacteraemia and impacts of infection control practices including universal MRSA admission screening in a hospital in Scotland, 2006-2010: retrospective cohort study and time-series intervention analysis. *BMJ Open* 2012;2: 1-16
- [7] Chambers HF. The changing epidemiology of Staphylococcus aureus. *Emerg Infect Dis* 2001;7:178-82.
- [8] Khorvash F, Abdi F, Kashani HH, Naeini FF, Narimani T. Staphylococcus aureus in Acne Pathogenesis: A Case-Control Study. *N Am J Med Sci*. 2012;4:573-6.
- [9] Mahmood K, Tahir T, Jameel T, Ziauddin A, Aslam H.F, Incidence of Methicillin-resistant Staphylococcus Aureus (MRSA) Causing Nosocomial
- [10] Infection in a Tertiary Care Hospital. *ANNALS* 2010 ;16:91-96
- [11] Bhutia OK, Singh TS, Biewas S, and Adhikari L. Evaluation of phenotypic with genotypic methods for species identification and detection of methicillin resistant in Staphylococcus aureus *International Journal of Applied and Basic medical Research*, 2012(2) : 84-91
- [12] Clinical and Laboratory Standards Institute (CLSI). Performance & Standards for antimicrobial susceptibility testing Twenty –third informational supplement. Clinical and Laboratory Standards Institute. 2013; 33(1). Table 2 C, Staphylococcus spp M02 & M07. 72-89
- [13] Baireddi D. Staphylococcus Micrococcus: Cluster forming Gram positive cocci In: Colle JG, Francer AG, Marmison BP, Simmons A, editors. Mackie and McCartney Practical Medical Microbiology, 14th edition. Edinburg; Churchill Livingstone; 2008; 245-262
- [14] Performance & standards for Antimicrobial susceptibility testing Twentieth informational supplement. Clinical and laboratory standards institute. 2010; 30 (1). Table 2C, Staphylococcus spp M02 & M07. 60-69
- [15] Sasirekha B, Usha M.S, Amruta A. J, Ankit S, Brinda N, Divya. Evaluation and Comparison of Different Phenotypic Tests to Detect Methicillin Resistant Staphylococcus aureus and their
- [16] Biofilm Production. *Int.J. PharmTech Res* 2012; 4(2) : 532-541
- [17] Mir BA, Srikanth. Prevalence and antimicrobial susceptibility pattern of Methicillin resistant Staphylococcus aureus and coagulase negative staphylococci in a tertiary care hospital in India. *Asian J Pharm Clin Res* 2013; 6 (3): 231-234
- [18] Oberoi L, Kaur R and Aggarwal A. Prevalence and antimicrobial susceptibility pattern of methicillin-resistant Staphylococcus aureus (MRSA) in a rural tertiary care hospital in north India. *IntJ of Applied Biol and Pharma Technology*. 2012; 3(1): 200-5.
- [19] Sharma M, Pathak S, Srivastava P, Prevalence and antibiogram of methicillin resistance Staphylococcus aureus at a tertiary care hospital in Jaipur Rajasthan. *IJPRBS* 2013; 2(1): 139-147
- [20] Sharma JB, Ahmed GU. Characterization of methicillin resistant S. aureus strains and risk factors for acquisition in a teaching hospital in northeast India. *India J Med Microbiol* 2010; 28(2): 127-9

- [21] Boucher H W, Corey G R. Epidemiology of Methicillin-Resistant *Staphylococcus aureus*. *Clinical Infectious Diseases* 2008; 46:344–9
- [22] Baddour .M.M., AbuElKheir MM., Fatani. A.J., Comparison of *mecA* Polymerase Chain Reaction With Phenotypic Methods for the Detection of Methicillin-Resistant *Staphylococcus aureus*. *CurrMicrobiol* .2007;55:473–479