

## Circular RNA in Cardiovascular Diseases

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**Abstract:** Circular RNA (circRNA) is a type of RNA molecule mainly formed by a covalent bond. There is a large number of circRNA in vivo, which has tissue and cell specificity. Research related to this shows that circRNA plays an essential role in the pathogenesis, diagnosis and treatment of cardiovascular diseases. This review summarizes the research of circRNA in biosynthesis and function, studies the role of circRNA in cardiovascular diseases, and proposes a new method for diagnosing and treating cardiovascular diseases.

**Keywords:** circular RNA; cardiovascular disease; cardiovascular system; mechanism.

### Introduction

In 1976, it was found that there were viroids in some plants, which were made up of circular RNA (circRNA) and caused infectious diseases to plants<sup>[1]</sup>. Subsequently, several studies<sup>[2-4]</sup> carrying out in-depth research on this observed the presence of circRNA in the human cytoplasm using electron microscopy. Although circRNA was observed by electron microscope, no attention was paid to it because of its low expression. Only when circRNA was ubiquitous in different plants and animals was recognized that circRNA

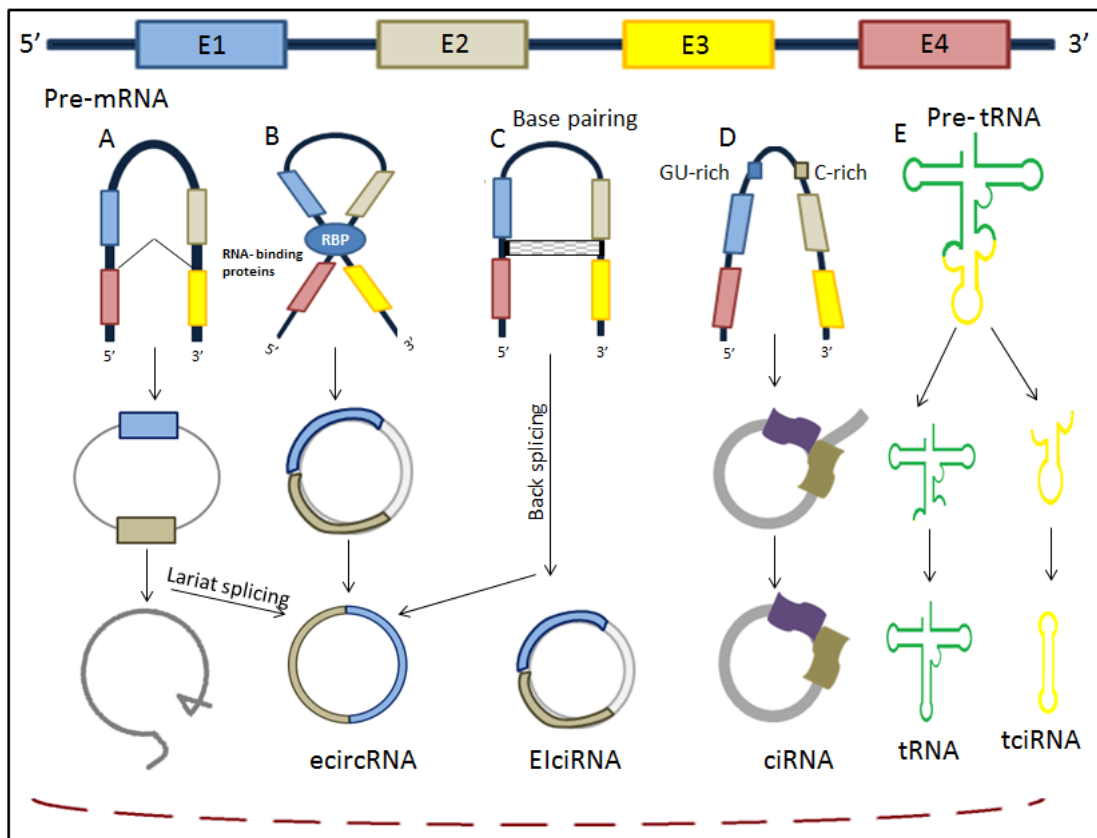
RNA was of great significance to organisms and plants. It has been found that circRNA exists in many tissues and organs and has a wide distribution and strong stability<sup>[4-5]</sup>. Unlike traditional linear RNA, circRNA cannot easily be degraded by exonuclease. Thus, it has a more stable performance. CircRNAs can play a regulatory role in genes and participate in various pathological processes. At present, circRNA plays an important role in a series of cardiovascular diseases such as atherosclerosis and myocardial infarction. Therefore, this paper reviews the relationship between circRNA and cardiovascular diseases, aiming to provide new ideas and targets for diagnosis and treatment of cardiovascular diseases [6-8]. Zhao, 2019, also highlighted the significance of circRNAs in the context of its biological formation, mechanisms performed by it, and its functional attributes 2020<sup>[102]</sup>.

## 1. Classification of circRNAs

CircRNAs were previously thought to be non-coding molecules produced by splicing errors. Recently, evidence shows that circRNAs are widespread and diverse in eukaryotic cells. Because of the lack of touchable end, circRNAs can avoid degradation by exonuclease; therefore, they are more stable than linear RNA. However, the formation mechanism and cellular function of circRNAs are not clear. As a member of non-coding RNAs, circRNAs have a unique covalent closed-loop structure, making them different from other non-coding RNAs, such as long non-coding RNAs (lncRNAs) and micro RNAs(miRNAs). High throughput sequencing technology has detected a large number of circRNAs with different lengths and types. Sequencing data analysis shows that they are expressed explicitly in developmental stages and are conservative between mice and humans. The latest research shows that circRNAs have many biological functions and play an important role in many diseases. VO *et al.* found that in more than 2000 tumour samples, the total amount of circRNA was generally reduced compared with adjacent normal tissues<sup>[9-12]</sup>.

With the development of sequencing technology, several types of circRNAs have been found and identified, mainly including four subtypes: exon type circRNAs (ecircRNAs), which mainly come from one or more exons; ring intron type circRNAs (lincRNAs), which only contain introns; exon-intron type circRNAs (eicircRNAs), which are composed of exons and introns; tRNA intron type circRNAs (tricroRNAs) which is formed by splicing the introns of pre tRNA. So far, most of the identified circRNAs are ecircRNAs. As shown in Figure 1A below is a lasso-driven cyclization. When pre-mRNA is spliced, the splicing receptor of the upstream exon and the downstream donor is close to producing a lasso structure containing the exon and the intron. After the intron in the lasso structure is removed, the exon is connected by the phosphodiester bond to form ecircRNAs.

Furthermore, 1B shows the cyclization driven by RNA binding protein (RBP). RBPs can promote the interaction between upstream intron and downstream intron and finally produce ecircRNA. Figure 1C shows the cyclization driven by base pairing. The upstream intron and downstream intron are based on the sequence complementary pairing of reverse repeat and complementary, and the intron is removed or retained to form eiciRNA or eicirRNA respectively. Fig. 1D shows the formation of circRNA<sup>[13]</sup>. The formation of circRNA mainly depends on a 7 NT Gu enrichment element and an 11 NT C enrichment element to avoid its debranching and degradation by exonuclease.



**Figure 1: Formation of tricRNA**

TRNA cleavage enzyme divides pre tRNA into two parts, one of which forms tricRNAs through 3' - 5' phosphate diester bond, the other produces tRNAs

## 2. The mechanisms of circRNAs

The mechanisms of circRNAs in human cancer are diverse, including miRNA sponges, epigenetic regulation, regulating gene splicing or transcription, translating into proteins or peptides and interacting with pro

teins. According to the target genes of circRNAs, there are two types: one is to regulate its host gene; the other is to target other genes<sup>[14]</sup>.

## **2.1 As miRNA Sponges:**

Currently, the main research direction of circRNAs is to use circRNAs as miRNA sponges or to translate them into short peptides to play their functions<sup>[15]</sup>.

As shown in Figure 2D, circRNAs can affect the expression of genes by adsorbing miRNAs. For example, CIRC HIPK3 can adsorb miR-7 and reverse the proto-oncogenes (EGFR, YY1, FAK, IGF1R) targeted by miR-7, thus promoting the progress of colorectal cancer<sup>[16-17]</sup>.

With the development of deep ribosome sequencing and mass spectrometry, many studies have recently identified and confirmed that some non-coding RNAs (including circRNAs) can encode proteins or small peptides. As shown in Figure 2F, for example, circRNAs derived from linc-pint can encode short peptides with 87 AA, which can directly interact with polymerase related factor complex (paf1c), thus, inhibiting the transcription of extension of a variety of cancers genes. CircRNA FBXW7 can also encode tumour related protein FBXW7-185aa, which can resist the stability of c-myc protein induced by USP28, thus reducing the half-life of c-myc protein; short peptide SHPRH-146aa encoded by circ-SHPRH can mediate the ubiquitination and degradation of PCNA<sup>[18]</sup>.

## **2.2 Epigenetic and post-translational regulation:**

CircRNAs can also play an important role in epigenetic regulation and post-translational regulation. For example, in Figure 2c, circna fecr1 originated from FLI1 gene can interact with FLI1's promoter, and generate extensive demethylation in the CpG island region of FLI1 gene by recruiting demethylase Tet1. At the same time, circRNA fecr1 can be combined with the promoter region enriched in h3k27ac of DNMT1 gene, thus inhibiting the expression of methyltransferase DNMT1 gene. As shown in Fig. 2E, circRNAs can regulate their activity or affect their expression by binding proteins. For example, circ-foxo3 can bind MDM2 and p53. When circ-foxo3 binds MDM2, it can enhance its polyubiquitin effect on p53, promote p53 degrade through the proteasome pathway<sup>[19]</sup>.

## **2.3 Regulation of expression of the host genes:**

Furthermore, circRNAs can also regulate the expression of their host genes.

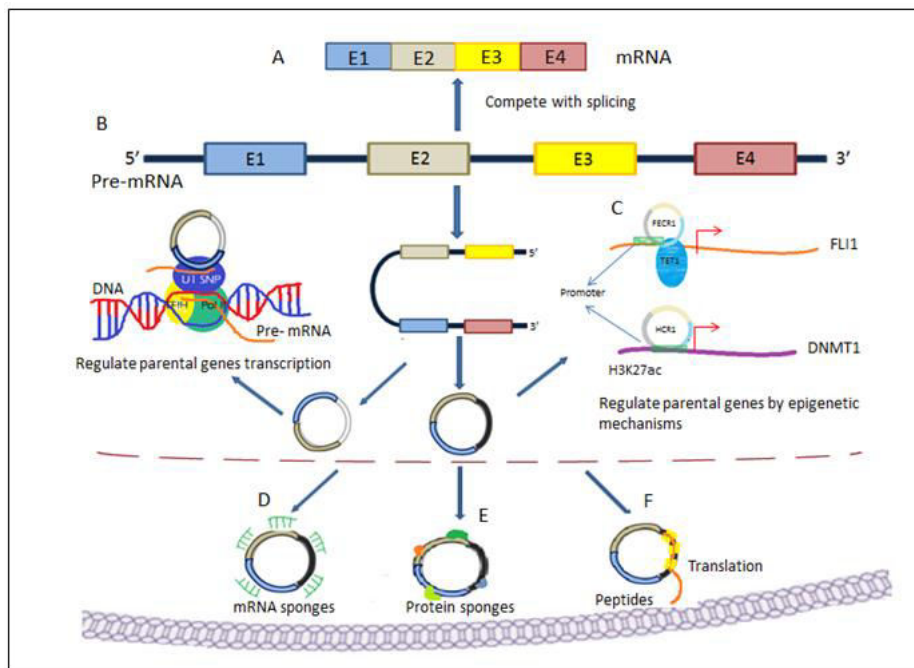
As shown in Figure 2 below, the formation of circRNAs can compete with linear RNAs for their common pre mRNA, thus affecting the abundance of linear RNAs. The circRNAs containing introns, introns can interact with U1 snRNP through RNA-RNA interaction and then combine with pol II transcription complexes, interacting to enhance the expression of their maternal genes<sup>[20]</sup>.

**2.4 Translation of circRNA into proteins:**

In addition to its function at the RNA level, circular RNA has been reported to translate into proteins like linear mRNA molecules. For example, hepatitis  $\delta$ , a circular RNA molecule in the hepatitis B virus, can encode a virus-related protein and play a role in the occurrence of diseases. However, at present, this phenomenon is only observed in viruses. The circular RNA molecules containing ATG initiation codon in eukaryotes can be recognized by ribosomes like linear mRNA for protein translation.

**2.5 Translation of proteins:**

CircRNA has been considered as non-coding RNA since it was found. The vast majority of circRNAs are derived from different protein-coding sequences and open reading frames, ubiquitous in the cytoplasm. Therefore, researchers have long believed that circRNA has the potential to translate proteins. As early as 1995, there was a literature report on circRNA. It is believed that some circRNA can effectively translate proteins. Circ-fbxw7 is a circRNA formed by cyclizing exon 3 and exon 4 of the tumour suppressor gene Fbxw7. It can translate a 185 amino acid protein fbxw7-185aa. Fbxw7 regulates the stability of proto-oncogene c-myc through ubiquitination. At the same time, fbxw7-185aa can cooperate with Fbxw7, reduce the expression of c-myc, and promote the ubiquitination and degradation of c-myc, thus inhibiting the occurrence of glioma. These studies break the understanding that circRNA is non-coding RNA, and have a new understanding of circRNA. However, through the translation of exogenous circRNA, there is direct evidence that endogenous circRNA can also be translated.



## Figure 2: The formation of circRNAs process

### 3. Functional significance of CircRNAs

CircRNA mainly exists in the cytoplasm and has been found in exosomes of the culture medium. CircRNA is related to the development of tissues and organs and may play a role in various disease processes. Its function is still largely unknown:

- (1) A few circRNAs act as "sponges" of miRNA;
- (2) Some circRNAs can "sponge" other factors, such as RNA binding proteins
- (3) A large number of circRNA may have other unknown regulatory functions;
- (4) Increased evidence suggests that circRNA may not be a real non-coding RNA type, at least some of which are translatable<sup>[20-21]</sup>.

Although many circRNAs may have biological functions, it is still likely that a large number of circRNAs may be insignificant by-products of the splicing of precursor mRNA<sup>[22]</sup>.

#### 3.1 CircRNA protein interaction

Some of the endogenous cytosolic circRNA can be used as scaffolds to regulate protein-protein interaction. Some RNA binding proteins (RBPs) can combine with circRNA to form competition between circRNA and its homologous mRNA, in which RBP affects translation. Moreover, our review of the available studies noted that some circRNA could regulate the expression of its binding protein by regulating protein-protein interaction<sup>[23]</sup>.

#### 3.2 Detection of circRNA protein interaction

The global bioinformatics analysis of circRNA sequence showed that there was no enrichment in the binding sites of RBP compared with linear mRNA. However, the third-order structure of circRNA may have more influence on protein binding than linear RNA sequence. So far, the interaction between circRNA and protein has been analyzed mainly by RNA pull-down assay or RNA immunoprecipitation (RIP)<sup>[24]</sup>.

The probe for reverse splicing point is the unique sequence element of circRNA. (1) In RNA pull-down assays, RNA is pulled down by probes to allow an analysis of related proteins. The use of circRNA overexpression and silencing techniques will help identify RNA protein interactions by quantitatively comparing pull-down circRNA with specific pull-down proteins<sup>[25]</sup>.

(2) RNA binding protein immunoprecipitation assay (RIP) followed by circRNA sequencing is another feasible strategy for analyzing circRNA protein interaction.

(3) RNase protection assay (RPA) is an effective method to detect RNA and RNA fragments in cell extracts. RPA can also be used to map protein RNA interactions.

(4) Microscopically analyzing the co-location of circRNA and protein is another strategy to identify the interaction between circRNA and protein<sup>[26]</sup>.

### **3.3 Dynamic circRNA protein interaction**

There is sufficient evidence showing that circRNA expression is dynamic and allows for different spatial and temporal expression profiles. CircRNA may form different third-order models, which prefer to bind specific proteins. Some third-order models may exist dynamically in some tissues and cells, resulting in different affinities with binding proteins. Because of the strong influence of solvent and metal ions on the dynamic third-order structure of circRNA, the main third-order structure of circRNA may be different in different cell lines, tissues and development stages<sup>[27-30]</sup>.

### **3.4 RNA protein interaction affects the dynamic expression of circRNA and protein**

CircRNA binding proteins play a key role in the regulation of circRNA synthesis and degradation. It has been shown that RNA circulation is promoted by complementary sequences and regulated by specific RNA binding proteins. RNA protein interaction can also promote the formation of circRNA by stabilizing complementary sequences or by inhibiting canonical splicing. RBP can be used as an activator or inhibitor of circRNA formation and regulate the expression level of circRNA. Because of the high stability of circRNA and its assumed resistance to exonuclease degradation, there may be cellular mechanisms controlling the level of circRNA<sup>[31]</sup>. At least some degradation pathways of circRNA can be mediated by endonuclease cleavage. RNA protein interaction affects protein expression, function and biogenesis.

### **3.5 CircRNA for targeted therapy**

Our recent studies have shown that the conjugation of circular RNA expression plasmids with nanoparticles is a useful method for the delivery of circular RNA. Because nanoparticles cannot enter the nucleus, the treatment can only focus on exon cyclic RNA, which is mainly detected in the cytoplasm<sup>[32]</sup>. Because of its high delivery efficiency, siRNA or AON delivery will be a valuable method in the future.

## **4. CircRNA and cardiovascular diseases**

CircRNA is a kind of RNA with a ring structure composed of covalent bonds. The main method for RNA detection is to isolate linear RNA molecules with polyadenylate tail (Polya) structure. In recent years, to identify circRNA by genes, after extracting the total RNA, the researchers remove the ribosome RNA and linear RNA to extract and sequence the circRNA, from which a variety of circRNA can be detected<sup>[33]</sup>. The regulatory role of circRNA runs through the whole process of gene regulation, from mRNA transcriptio

n splicing to RNA degradation and translation. There are two screening methods widely used; one is sequencing by high throughput, the other is sequencing analysis by chips<sup>[34-36]</sup>.

The changes of circRNA in cardiovascular diseases are shown in Table 1<sup>[37]</sup>.

**Table 1: circRNA changes in cardiovascular disease**

Disease	Detection mode	Detection objects	Sample	CircRNA Expression change
<b>Acute myocardial infarction</b>	Chip detection	Myocardial infarction patients	Plasma	Upregulation of 73 circRNAs Downregulation of 87 circRNAs
<b>Myocardial fibrosis</b>	Chip detection	Diabetic mice	Myocardial tissue	Upregulation of 45 circRNAs Downregulation of 31 circRNAs
	Chip detection	Diabetic mice	Myocardial tissue	Upregulation of 24 circRNAs Downregulation of 19 circRNAs
<b>Coronary artery disease</b>	Chip detection	Patients with coronary artery disease	Peripheral blood	Upregulation of 12 circRNAs Downregulation of 10 circRNAs
<b>Diabetes</b>	Chip detection	Diabetic retinopathy	Serum	Upregulation of 30 circRNAs
<b>Heart failure</b>	Chip detection		Myocardial tissue	Upregulation of 29 circRNAs Downregulation of 34 circRNAs
<b>Chronic total occlusion pulmonary hypertension</b>	Chip detection	Chronic total occlusion pulmonary hypertension	Peripheral blood	Upregulation of 122 circRNAs Downregulation of 229 circRNAs

At present, the role of circRNA in cardiovascular disease is still incomplete, and there are few studies in this area. We need to know the role of circRNA in cardiovascular disease to find the relevant treatment measures.<sup>[38]</sup> Wilson *et al.* sequenced circRNA in human heart, mouse heart, and human embryonic stem cell differentiated cardiac tissue and found 15318 and 3017 circRNA in human and mouse hearts. The expression level of these circRNAs is consistent with that of their homologous linear RNAs. The genes correspond



ending to the highest content of circRNAs are also cardiac tissue-specific genes, such as titin, RyR2 and DMD genes<sup>[39]</sup>.

#### 4.1 CircRNA and heart failure

Mir-223 is a positive regulator of cardiac hypertrophy. Apoptosis suppressor with card (ARC) is the downstream target of mir-223, and the cardiac hypertrophy response of arc transgenic mice is decreased. Heart-related circular RNA (HRCR) is a recently reported cardiac circRNA, which acts as a "sponge" of miR-223, directly combines with miR-223, inhibits the activity of miR-223, and thus increases the expression of ARC. The overexpression of Hrcr in cardiomyocytes and mice showed a decreased cardiac hypertrophy<sup>[39-42]</sup>. These findings reveal a new regulatory pathway and therapeutic target of cardiac hypertrophy/heart failure composed of circular, mir-223 and arc. Werfel *et al.* constructed a library for RNA sequencing by removing ribosomal RNA, and analyzed the expression of circRNA in the heart of humans, mice and rats at different stages of development or physiological and pathological states. The results showed that the total expression of circRNA in human cardiac tissue was higher than that of rats and mice<sup>[43]</sup>. The rats were divided into the newborn rat's group and the adult rat's group. There were significant differences between the two groups in the circRNA expression of slc8a1, TTN, eya3 and scmh1 genes. Humans and mice were divided into heart failure group and non-heart failure group<sup>[44]</sup>. It was found that the number and types of circRNA in the heart failure group were higher than those in the non-heart failure group. For example, the expression of circRNA corresponding to slc8a1 and TTN genes in the heart failure group increased significantly. In particular, the ryanodine receptor 2 (RyR2) genes exist in human heart tissue, corresponding to more than 100 subtypes of circRNA<sup>[45]</sup>. In conclusion, circRNA is closely related to the physiological and pathological process of heart, especially the genes corresponding to several differentially expressed circRNA molecules, such as slc8a1, TTN, RyR2, eya3, etc., which can be used as ideal candidate genes for further study of heart failure<sup>[46-49]</sup>.

#### 4.2 CircRNA and CAD

It is reported that compared with the healthy control group, the plasma endothelial cells of CAD patients are rich in endothelial cells. MiRNAs (miR-126, mir-92a and miR-17) were collected from smooth muscle cells (SMCs). MiR-145 and inflammation-related miR-155 were significantly decreased. Another study examined 157 different miRNA microarrays which were detected in peripheral blood mononuclear cells of CAD patients<sup>[50-51]</sup>. The results showed that mir-135a increased and mir-147 decreased significantly in CAD patients, and the ratio can be used in CAD diagnosis. This study further confirmed that increased the l

level of mir-134, 198 and mir-370 may help to distinguish unstable angina patients from stable angina patients. It is suggested that circulating miRNAs can be used to predict acute coronary syndrome in patients with angina pectoris<sup>[52]</sup>. Sun et al. Used QRT PCR to detect 31 cases of CAD, and plasma miR-126 was detected in 36 non-CAD patients. However, it was found that the expression of miR-126 was increased in patients with elevated low-density lipoprotein cholesterol (LDL-C) and decreased in patients with low-density lipoprotein cholesterol (LDL-C), suggesting a specific correlation between miR-126 and lipid metabolism. In addition, mir-149 was found to be associated with an increased risk of CAD in the Chinese Han population<sup>[53-56]</sup>.

Serum miR-31 was abnormally elevated in patients with CAD restenosis compared with patients without CAD restenosis. Compared with the healthy control group, circulating blood mir-133a and mir-208a levels were up-regulated in patients with stable coronary artery disease, while miR-126, miR-17 and mir-208a were up-regulated. The levels of miR-92a and miR-155 decreased significantly, and miR-214 was found to be beneficial to CAD-Patient. This may be a promising biomarker for severe CAD; the loss of its protection may increase placental growth factors and the deterioration of atherosclerosis. Recently, the result of several studies showed that the miR-122 and mir-370 levels were circulated. It may be related to the severity of CAD. The expression level was positively correlated with TC, TG and LDL-C levels. All in all, follow miRNAs can improve the diagnosis of CAD<sup>[57-58]</sup>.

### **4.3 CircRNA and myocardial fibrosis**

Tang *et al.* found that circRNA\_000203 upregulated in the myocardium of diabetic mice and Ang-II induced mice cardiac fibroblasts. CircRNA\_000203 has two potential binding areas for microRNA-26b-5p (miR-26b-5p), which shows the anti-fibrotic effect targeting I collagen and connective tissue. In availability of a higher number of circRNA\_000203 could obliterate the anti-fibrotic effect of miR-26b-5p in cardiac fibroblasts. Thus circRNA\_000203 promotes the proliferation of cardiac fibroblasts by blocking the function of miR-26b-5p. It has also been found that circacta2, as a circRNA, inhibits the miR-548f-5p pair.

### **4.4 CircRNA and diabetic cardiomyopathy**

It has been proved that diabetes mellitus can cause changes of myocardial structure and function in varying degrees, which will lead to cardiac hypertrophy and myocardial fibrosis. Tang *et al.* the circRNA in mouse cardiac tissue found 76 different circRNA expressions, 45 of which were up-regulated and 31 of which were down-regulated<sup>[58-62]</sup>. At the same time, the researchers selected the up-regulated circRNA000203 and found that there are two binding sites between circRNA000203 and miR-26b-5p<sup>[63-65]</sup>. Overexpression of circRNA\_000203 in cardiac fibroblasts will lead to up-regulation of the downstream target fibrosis-related g

enes COL1A2 and CTGF of mir-26b-5p. Therefore, cir-crn000203 may be a new target for the treatment of myocardial fibrosis in diabetic cardiomyopathy [65-67].

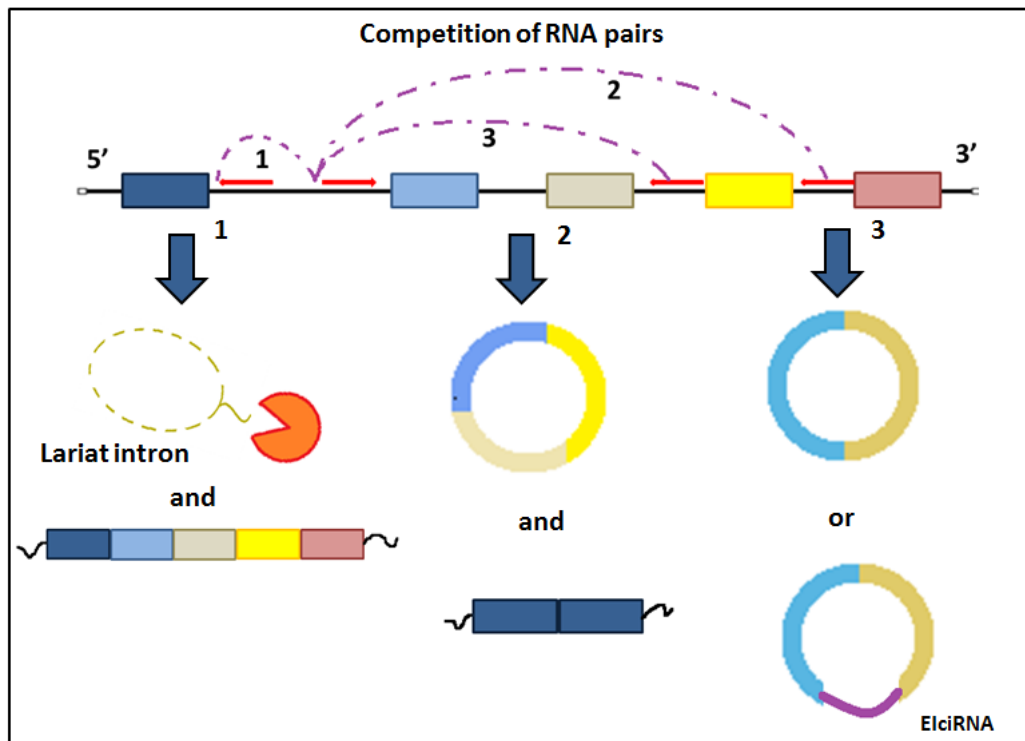


Figure 3: The categories exon derived circular RNAs

**Conclusion**

The mortality of cardiovascular disease is high, and treatment and nursing are complex. In addition to conventional treatment, gene therapy technology can also be used to treat cardiovascular disease. The use of disease-specific antisense miRNA molecules to inhibit disease has entered the clinical trial stage. At the molecular level, the antisense nucleotide chain of miRNA can improve the affinity between miRNA and cells to achieve the purpose of treatment. For LNA gapmers technology, the direct target is lncRNA. After the target is combined with lncRNA, RNA endonuclease can be activated rapidly to degrade lncRNA. Unlike miRNA and lncRNA, circRNA can resist ribonuclease endonuclease, so it cannot be degraded by ribonuclease endonuclease, remaining stable in vivo. Therefore, for circRNA, it may become a new method of gene therapy.

CircRNA can combine with miRNA to inhibit the expression of the target gene. Foreign media reports have confirmed that circRNA molecules can target miR-122, which is closely related to the hepatitis C virus

, to inhibit the formation of the virus and achieve the purpose of disease treatment. CircRNA can also play a role by regulating gene expression, protein translation, production and other mechanisms. Therefore, the synthetic circRNA may also target genes or proteins and directly affect diseases. In recent years, it has been shown that circRNA can play an essential role in the regulation of cardiovascular diseases and achieve the goal of treating cardiovascular diseases<sup>[68]</sup>. However, the current understanding of the biological function of circRNA needs further study, and there is limited knowledge of its biosynthesis and degradation process.

In conclusion, circRNA is considered to be an abnormal splicing product. Currently, some studies suggest that circRNA can play an important role in gene therapy, particularly in a series of cardiovascular diseases such as atherosclerosis, myocardial infarction and myocardial fibrosis. However, the mechanism of circRNA in cardiovascular diseases is not much clear, and there are still many gaps of knowledge. It is necessary to strengthen the relevant research and explore the mechanisms of circRNA in cardiovascular diseases to find a new target of cardiovascular disease treatment and diagnosis.

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