Original Research Article COMPARISON OF VARIOUS PHENOTYPIC METHODS OF CARBAPENEMASE DETECTION AMONG KLEBSIELLA ISOLATED FROM CLINICAL SPECIMENS

¹Dr Rishi Dhurve, ²Dr Puja Rawat, ³Dr Suneel Kumar Ahirwar, ⁴Dr Manish Purohit, ⁵Dr Satakshi Manwani, ⁶Dr. Yogesh Dodiyar, ⁷Dr. Shashi Gandhi

¹Ex PG student, Department of Microbiology, MGM Medical College, Indore, Madhya Pradesh, India
²Assistant Professor, Department of Pediatrics, Chirayu Medical College and Hospital, Bhopal, Madhya Pradesh, India
³Associate Professor, Department of Microbiology, MGM Medical College, Indore, Madhya Pradesh, India
⁴Associate Professor Department of Microbiology, MGM Medical College, Indore, Madhya Pradesh, India
⁵Assistant Professor, Department of Microbiology, Chirayu Medical College and Hospital, Bhopal, Madhya Pradesh, India
⁶Senior Resident, Department of Anesthesiology, Government Medical College, Ratlam, Madhya Pradesh, India
⁷Professor and Head, Department of Microbiology, MGM Medical College, Indore, Madhya Pradesh, India

drshashigandhi@gmail.com

ABSTRACT

Background: Carbapenems are frequently used as a last resort antibiotic in the treatment of infections due to multidrug-resistant Enterobacteriaceae. Their alarming increase worldwide is worrisome and is left with very restricted therapeutic alternatives.

Objectives: This study compares the various phenotypic methods for detection of carbapenemase producers in Klebsiella species isolated from different clinical samples.

Methods: All specimens received for culture and sensitivity were processed as per standard guidelines. Identification of Klebsiella spp. was done using biochemical tests as per standard guidelines. Carbapenemase production was detected phenotypic screening methods as per CLSI guidelines. Confirmation done by using Modified Hodge Test, Modified carbapenem inactivation method (mCIM) and EDTA-modified carbapenem inactivation method (eCIM)

Results: Out of total 160 Klebsiella isolates test for carbapenemase production 101 (63.1%) was positive. MHT was positive in 76.2% isolates, whereas MCIM and ECIM were positive in 91.1% isolates. The sensitivity, specificity and PPV of MHT with MCIM were 82.6%, 88.9%, and 98.7% respectively, whereas sensitivity, specificity and PPV of ECIM with MCIM were 100%, 100%, and 100% respectively.

Conclusion: MCIM in conjunction with ECIM test could be considered as more reliable phenotypic diagnostic methods for carbapenemase detection than the MHT in conjuction with the MCIM.

Keywords: MHT, MCIM, ECIM, Carbapenemase Producing Organisms

1. INTRODUCTION

Carbapenems are frequently used in the treatment of infections due to multidrug resistant Enterobacteriace. The emergence and spread of carbapenem-resistant gram-negative bacteria is a worldwide public health threat [1]. The mechanisms underlying carbapenem resistance in Enterobacteriace are complex and include both the production of carbapenem-hydrolyzing lactamases (carbapenemase-producing CRE) and resistance due to the presence of a combination of other factors, such as hyperproduction of AmpC-lactamases or extendedspectrum -lactamases (ESBLs) combined with altered membrane permeability [2-3]. The major mechanism of beta- lactamase production is excessive use of beta- lactams in treating infections caused by the Enterobacteriace in recent years, [4]. Among the carbapenemases, Klebsiella pneumoniae carbapenemase (KPC, Class A), imipenemase (IMP, Class B), Verona integrin-encoded MBL (VIM, Class B), New Delhi metallo beta-lactamase (NDM, Class B), and oxacillinases (OXA, Class D) are the major types. These enzymes confer carbapenem resistance through hydrolysis [5-6]. MBL and KPC Carbapenemases are mostly encoded by mobile transposon and/or integron determinants resulting in faster dissemination to the other members of Enterobacteriaceae with limited therapeutic choices [7]. Bacterial isolates, which are capable of producing carbapenemase enzymes, have the ability to inactivate a wide range of β -lactams, including penicillins, cephalosporins, carbapenems, and monobactams [8]. Both phenotypic and molecular-based assays are available for the detection of carbapenemase producers from cultured isolates. Phenotypic assays currently used in clinical practice consist of the following: (i) modified Hodge test [MHT]), modified carbapenem inactivation method [mCIM]); Carba NP, matrix-assisted laser desorption-ionization time of light mass spectrometry [MALDI-TOF MS] methods); and lateral flow immunoassays [9-10]. The modified Carbapenem Inactivation Method (mCIM) is a simple phenotypic test that detects carbapenemases-producing gram- negative bacteria that have only been evaluated for use on bacterial colonies [11]. The Clinical and Laboratory Standards Institute (CLSI) lowered the carbapenem breakpoints for the Enterobacteriaceae in 2010 [12]. The revised breakpoints recommend meropenem or imipenem susceptibility with MICs of $\leq 1 \mu g/ml$ and ertapenem susceptibility with MICs of $\leq 0.5 \ \mu g / ml$.

Aims & objectives: this study evaluates the various phenotypic methods of the carbapenemase producers detection in Klebsiella isolated from clinical specimens in our tertiary care center.

2. MATERIALS AND METHODS

This was a cross-sectional observational study carried out in the Department of Microbiology, MGM Medical College and associated M Y Hospital, Indore (M.P). Study was undertaken from June 2021 to June 2022 (01 years) after the approval from the ethics committee. A total of 160 Klebsiella species isolated from various clinical samples were enrolled in this study. **Inclusion Criteria:**

- All Klebsiella spp. isolated from different specimen received for culture and sensitivity in microbiology department
- Patients who provide written informed consent for the study **Exclusion Criteria**:
- Repeat isolation of Klebsiella spp. from same patient
- Patients who not provide consent for the study

All specimens received for culture and sensitivity were processed as per standard guidelines. Isolation and identification of Klebsiella spp. was done using biochemical tests as per

standard guidelines.

Antimicrobial sensitivity testing of isolated Klebsiella spp. for Imipenem and meropenem was done using Kirby Bauer disc diffusion and for Colistin using broth dilution method as per CLSI M100 2022 guidelines.

Screening criteria for carbapenemase production: Organism which were found resistant to imipenem and/or Meropenem were considered as probable Carbapenemase producers and considered as Screen positive.

Confirmation of carbapenemase production among screen positive Klebsiella was done using

- Modified Hodge Test
- Modified carbapenem inactivation method (mCIM)
- EDTA-modified carbapenem inactivation method (eCIM)

Carbapenemase positive: Zone diameter of 6–15 mm or presence of pinpoint colonies within a 16–18 mm zone

Carbapenemase negative: Zone diameter of \geq 19 mm (clear zone)

Carbapenemase indeterminate: Zone diameter of 16-18 mm

Statistical analysis: SPSS 20 was incorporated in the analysis of the collected data (SPSS inc). The categorical or dichotomous variables were expressed as absolute values and percentages, and were compared with Pearson test. The continuous variables with a normal distribution were described as the mean (+/-SD). A P value less than .05 was considered Statistically Significant.

3. RESULTS

A total of 160 Klebsiella species isolated from various clinical specimens, out of that the higher proportion 81.9% was Klebsiella pneumoniae, 14.4% Klebsiella oxytoca and least 3.8% was Klebsiella Ozaenae. The higher proportion (38.1%) was urine followed by pus (33.2%) and the lower proportion (3.1%) was CSF respectively

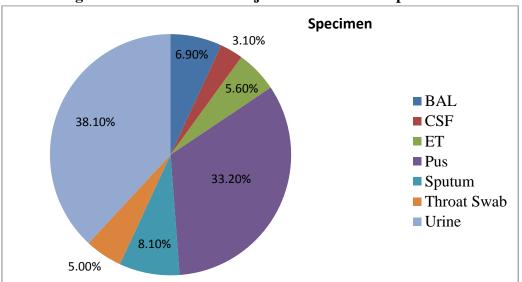


Figure 1: Distribution of subjects on the basis of specimens

All the Klebsiella isolated was screened for carbapenem resistance by production of carbapenemase. The KPC was used to confirm carbapenemase production among the carbapenem⁻ resistant Klebsiella isolates. Among the screening test 101 (63.1%) isolates was carbapenemase production test positive [figure: 2].

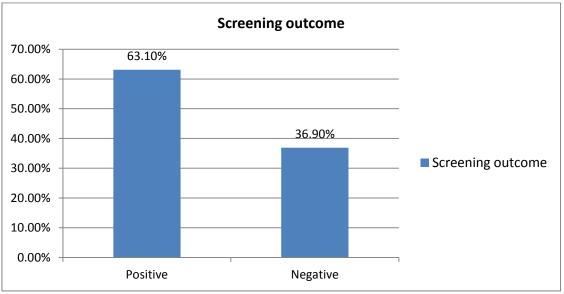


Figure 2 Distribution on Basis of Screening for carbapenamase production

The carbapenemase screening test positive isolated was confirmed by Modified Hodge test (MHT), Modified Carbapenem inactivation method (MCIM) and EDTA Carbapenem inactivation method (ECIM). All these confirmatory methods were compared.

MHT was positive in 76.2% isolates, whereas MCIM and ECIM were positive in 91.1% isolates. Details of comparison between them was shown in table: 1

Methods		Frequency (n=101)	Percent
Modified Hodge test (MHT)	Positive	77	76.2%
	Negative	24	23.8%
Modified Carbapenem inactivation	Positive	92	91.1%
method (MCIM)	Negative	9	8.9%
EDTA Carbapenem inactivation	Positive	92	91.1%
method (ECIM)	Negative	9	8.9%

Table 1 Comparisons between various methods of carbapenamase detection

The higher proportion 100% was for Positive outcome and the lower proportion 0.0% was for Negative outcome respectively. There was significant association between ECIM and MCIM outcomes (P<0.05).

ECIM	MCIM	MCIM		
	Positive	Negative		
Positive	92 (100%)	0 (0.0%)	92 (91.1%)	
Negative	0 (0.0%)	9 (100%)	9 (8.9%)	
Total	92 (100%)	9 (100%)	101 (100%)	
Chi-Square Value	df	P Value	Result	
101.000	1	0.00	Significant	

Table 2: Association between ECIM and MCIM

The higher proportion 98.7% was for Positive outcome and the lower proportion 1.3% was for Negative outcome respectively, association between MHT and MCIM outcome was significant (P<0.05).

MHT	MCIM		Total
	Positive	Negative	
Positive	76 (82.6%)	1 (11.1%)	77 (76.2%)
Negative	16 (17.4%)	8 (88.9%)	24 (23.8%)
Total	92 (100%)	9 (100%)	101 (100%)
Chi-Square Value	df	P Value	Result
23.133	1	0.00	Significant
23.133	1	0.00	Significant

Table 3: Distribution of MCIM Outcome for MHT Positive Outcome

4. **DISCUSSION**

Increased antibiotic resistance to carbapenems among Klebsiella isolates has become a major public health problem. The accurate and rapid detection of carbapenemase producing Klebsiella isolates is necessary for appropriate treatment, prevention of spreading, and control of infections. In the last decade, phenotypic methods were extensively used in clinical

laboratories for a first-line detection of the isolates producing carbapenemases [13].

In our study majority of Klebsiella species were isolated from urine cultures followed by pus culture, similar observation reported by Parveen RM et al [14] and S K Pawar et al [15], discordance to our study, Valarmathi et al [16] observed maximum number of Klebsiella were isolated from pus, whereas Manikandan et al [17] showed that most of the Klebsiella species were isolated from sputum sample.

The organism found resistant to imipenem and/or Meropenem were considered as probable Carbapenemase producers and considered as Screen positive.

Current study found 63.1% of Klebsiella isolates were found to be Carbapenemase producers by screen test, in agreement with S Pandurangan, et al [18] and Wattal C, et al [19] reported Carbapenemase producers by screen test were 52% and 51% respectively. The major differences were likely due to geographic region, the testing method used, and the organism.

For confirmation of carbapenemase production among screen positive, Modified Hodge test, MCIM and ECIM was done.

Modified Hodge test (MHT) showed 76.2% positivity among the screening positive isolates in the present study, consistence finding observed by M Beig, et al [20] and C.G.Carvalhaes, et al [21].

Our study found Modified Hodge test was found to be 100% sensitive and specific for detection of carbapenem resistance in K. pneumoniae. Bashir H et al [22], reported modified Hodge test as a reliable and sensitive laboratory method for detection of carbapenem.

Current study reported MHT and MCIM are equally sensitive and effective methods for the detection of carbapenemase producer in Klebsiella species, our results comparable with the Tamma PD, et al [23].

ECIM and MCIM test showed similar sensitivity with (91.1%) positives and (8.9%) negatives, accordance to the Khare et al [24] and Bartolini A, et al [25].

Among various phenotypic methods for detection of carbapenem resistance, modified Hodge test (the Clover leaf test) is relatively simple and easy. It can be easily incorporated as a routine technique in laboratories with heavy workload. Since this test is recommended by CLSI, it is extensively used as a phenotypic method for detection of carbapenem resistance.

5. CONCLUSION

We have concluded that MCIM and ECIM test showed similar sensitivity, However, Modified Hodge test (MHT) showed lower sensitivity for detection of carbapenemase producers. As MCIM with ECIM can differentiate between both, it could be utilized as simple, reliable, cost-effective phenotypic method for carbapenemase detection which will contribute in the formulation of better treatment plan to curtail therapeutic failure.

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6. REFERENCES

- 1. Bush K and Fisher JF. Epidemiological expansion, structural studies, and clinical challenges of new β -lactamases from gram-negative bacteria. Annu Rev Microbiol 2011; 65:455-78.
- 2. Vasoo S, Barreto JN, Tosh PK. 2015. Emerging issues in gram-negative bacterial resistance: an update for the practicing clinician. Mayo Clin Proc 90:395–403. https://doi.org/10.1016/j.mayocp.2014.12.002.
- Lutgring JD, Limbago BM. 2016. The problem of carbapenemase producingcarbapenem-resistant-Enterobacteriaceae detection. J Clin Microbiol 54:529–534. https://doi.org/10.1128/JCM.02771-15.
- 4. Livermore DM, Woodford N. The beta-lactamase threat in Enterobacteriaceae, Pseudomonas and Acinetobacter. Trends Microbiol 2006;14:413-20.
- 5. Singh M, Kakati B, Agarwal RK, Kotwal A. Detection of Klebsiella pneumoniae carbapenemases (KPCs) among ESBL/MBL producing clinical isolates of Klebsiella pneumoniae. Int J Curr Microbiol App Sci. 2015; 4(4):726-731.
- 6. Shields RK, Potoski BA, Haidar G, et al. Clinical outcomes, drug toxicity and emergence of ceftazidime avibactam resistance among patients treated for carbapenem-resistant Enterobacteriaceae infections. Clin Infect Dis. 2016 2; 63(12):1615-1618. doi: 10.1093/cid/ciw636.
- 7. Athanassios Tsakris, AggelikiPoulou Spyros Pournaras, EvangeliaVoulgari, Georgia Vrioni, Katerina Themeli-Digalaki, Dimitra Petropoulou and DanaiSofianou. 2010. A simple phenotypic method for the differentiation of metallo- lactamases and class A KPC carbapenemases in Enterobacteriaceae clinical isolates. J. Antimicrob. Chemother, 65: 1664 1671.
- 8. E. A. Idelevich, K. Sparbier, M. Kostrzewa, and K. Becker, "Rapid detection of antibiotic resistance by MALDI-TOF mass spectrometry using a novel direct-on-target microdroplet growth assay," Clinical Microbiology and Infection, vol. 24, no. 7, pp. 738–743, 2018.
- Lupo A, Papp-Wallace KM, Sendi P, Bonomo RA, Endimiani A. 2013. Non-phenotypic tests to detect and characterize antibiotic resistance mechanisms in Enterobacteriaceae. Diagn Microbiol Infect Dis 77:179–194. <u>https://doi.org/10.1016/j</u>. diag microbio. 2013.06.001.
- 10. Rood IGH, Li Q. 2017. Review: molecular detection of extended spectrum-betalactamase- and carbapenemase-producing Enterobacteriaceae in a clinical setting. Diagn Microbiol Infect Dis 89:245–250. https://doi.org/10.1016/j.diagmicrobio.2017.07.013.
- Pierce VM, Simner PJ, Lonsway DR, et al. Modified carbapenem inactivation method for phenotypic detection of carbapenemase production among Enterobacteriaceae. J Clin Microbiol. 2017; 55 (8):2321–2333. doi:10.1128/JCM.00193-17.
- 12. ClinicalandLaboratoryStandardsInstitute.2018.Performancestandards for antimicrobial susceptibility testing, 28th ed. CLSI supplement M100. Clinical and Laboratory Standards Institute, Wayne, PA.

- 13. S.R. Evans, A.M. Hujer, H. Jiangetal., "Informing antibiotic treatment decisions: evaluating rapid molecular diagnostics to identify susceptibility and resistance to carbapenems against acinetobacter spp. in PRIMERS III," Journal of Clinical Microbiology, vol. 55, no. 1, pp. 134–144, 2017.
- Parveen RM, Harish BN, Parija SC. Emerging carbapenem resistance among nosocomial isolates of Klebsiella pneumoniae in South India. Int J Pharma Bio Sci. 2010; 1:1-11.
- K Pawar S, T. Mohite S, V. Shinde R, R. Patil S, S. Karande G. Carbapenem resistant Enterobacteriaceae: Prevalence and bacteriological profile in a tertiary teaching hospital from rural western India. Indian J Microbiol Res. 2018;5(3):342- 347. doi:10.18231/2394-5478.2018.0072.
- 16. Valarmathi, S., M. Rajasekara Pandian and B. Senthilkumar. 2013. Incidence and screening of wound infection causing microorganisms. J. Acad. Indus. Res. 1(8): 508.
- 17. Manikandan, C., and A.Amsath. 2013. Antibiotic susceptibility of bacterial strains isolated from patients with respiratory tract infections. International Journal of Pure and Applied Zoology. 1(1): 61-69.
- Sathya Pandurangan, Shamsadh BegumEsak* and Anbumani Narayanasamy, Phenotypic Detection Methods of Carbapenemase Production in Enterobacteriaceae, Int.J. Curr. Microbiol.App.Sci (2015) 4(6): 547-552.
- 19. Wattal C, Goel N, Oberoi JK, Raveendran R, Datta S, Prasad KJ. 2010. Surveillance of multidrug resistant organisms in a tertiary care hospital in Delhi, India. J Assoc Physicians India, 58: 32 6.
- 20. Masoumeh Beig , Mohammad Taheri and Mohammad Reza Arabestani, Comparison of Different Phenotypic Tests versus PCR in the Detection of Carbapenemase-Producing Pseudomonas aeruginosa Isolates in Hamadan, Iran, International Journal of Microbiology Volume 2021, Article ID 5582615, 8 pages.
- 21. C.G.Carvalhaes, R.C.Picao, A.G.Nicoletti, D.E.Xavier, and A. C. Gales, "Cloverleaf test (modified Hodge test) for detecting carbapenemase production in Klebsiella pneumoniae: be aware of false positive results," Journal of Antimicrobial Chemotherapy, vol. 65, no. 2, pp. 249–251, 2010.
- 22. Bashir H, Kakru D, Qadri S, Bali N, Bashir S, Lone S. Carbapenem resistant Klebsiella pneumoniae in a tertiary care Hospital in Northern India. International Journal. 2014; 2:92-102.
- Tamma PD, Simner PJ.2018. Phenotypic detection of carbapenemase producing organisms from clinical isolates. J Clin Microbiol 56:e01140-18.https://doi.org/10 .1128/JCM.01140-18.
- Khare AP, Gopinathan A, Leela KV, Naik S. Comparison of Three Phenotypic Methods of Carbapenemase Enzyme Detection to Identify Carbapenem-resistant Enterobacterales. J Pure Appl Microbiol. 2022;16(4):2679-2687. doi: 10.22207/JPAM.16.4.37.
- 25. Bartolini A, Frasson I, Cavallaro A, Richter SN, Palu G. Comparison of phenotypic methods for the detection of carbapenem non-susceptible Enterobacteriaceae. Gut Pathog. 2014; 6:13. doi: 10.1186/1757-4749-6-13.