

ANTIMICROBIAL EFFICACY OF DIFFERENT HERBAL EXTRACTS AS INTRACANAL MEDICAMENT AGAINST ENTEROCOCCUS FAECALIS

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ABSTRACT

Introduction: A successful endodontic treatment depends upon complete debridement of microflora from the root canal system. However, due to complex root canal configuration, complete debridement through mechanical instrumentation alone cannot remove entire bacterial load. So, the aim is *in vitro* evaluation of antimicrobial efficacy of biological extracts against *Enterococcus faecalis* MTCC-439 strain when used as intracanal medicaments. The medicaments used were Nisin, an antibiotic peptide; calcium hydroxide, curcumin and aloe vera

Materials and Methods: Eighty single rooted lower premolar teeth which were extracted for orthodontic purpose were collected. Tooth specimens were sectioned at cement-enamel junction with a diamond saw to obtain a standard root length.

Complete biomechanical preparation will be done, which will be followed by inoculation of *E. faecalis*

After inoculation, the samples will be kept in a closed eppendorf tube and incubated at 37°C for 21 days under aseptic conditions. The canals will be re-inoculated with fresh bacterial samples at every 3 days interval to ensure viability of bacteria.

The canal contents will be aspirated after 21 days of incubation, then rinsed with 5 mL saline and patted dry with sterile paper points. The specimens will be then randomly divided into following groups:

Group 1: Calcium hydroxide

Group 2: Aloe vera

Group 3: Curcumin

Group 4: Nisin Calcium hydroxide.

Results: In present *in vitro* study, Nisin showed no CFU while aloe vera and curcumin showed significantly less growth as compared to calcium hydroxide against *E. faecalis*.

Conclusions: Nisin outperforms curcumin and aloe vera in eliminating the *E. faecalis* when used as intracanal medicaments.

INTRODUCTION

A successful endodontic treatment depends upon complete debridement of microflora from the root canal system.[1] However, due to complex root canal configuration,

completed debridement through mechanical instrumentation alone cannot remove entire bacterial load.

Contemporarily use of chemical intracanal irrigators and medicaments are requisite to debride infected tissues and eradicate microorganisms from the root canal system.[2] Moreover, low oxygen tension, less nutrient availability, and enormous bacterial interactions lead to predominant colonization of facultative anaerobic species prevailing in the root canals.[3] In persistent peri radicular infections, *Enterococcus faecalis* had been isolated in about 24%–77% cases that perpetually resulted in failure of root canal therapy. This could be because of the ability of *E. faecalis* to survive at high alkaline environment and deeper tubular invasion. It grows through adhering on biofilm and colonizes on to the surface.[4]

Calcium hydroxide is a gold standard traditionally when used as intracanal medicaments. Although its antibacterial activity is on wide range of microflora of the root canal, but was found less effective against *E. faecalis*. [5-8] Moreover, increasing rates of cytotoxic reactions and inability to eliminate the microorganism from dentinal tubules by commercially available medicaments had laid a need of introduction of novel molecules used as intracanal medicaments. In last few decades, the use of alternative therapeutic agents had considerably increased and are derived from plants, insects, microorganisms, etc.[1,3,9]

A natural occurring peptide isolated from strains of *Lactococcus lactis* termed as Nisin had been recently introduced. This antibiotic peptide is a Class I bacteriocin. It is effective against Gram-positive bacteria and spores[7,3] including strains of *E. faecalis*. [10] Nisin intensively used as a food preservative in over 40 countries and is safe to humans. Recent documents are suggestive that nisin is effective in eradication of *E. faecalis* from root canals.[11]

Aloe vera (*Aloe barbadensis* miller) belongs to the Liliaceae family and is a cactus-like plant.[12] It is a natural medicament with a long history of usage in medicine and nutrition. The antimicrobial properties of *A. vera* against various species of microorganisms, including *E. faecalis*, have also been reported as *A. vera* has potent antibacterial, antiviral, and antifungal activities.[12,14] The possible reason for antimicrobial action of *A. vera* could be the presence of 75 potentially active constituents: vitamins, enzymes, minerals, sugars, lignin, saponins, salicylic acids, and amino acids.[15] The antimicrobial activity of *A. vera* might be attributed to the presence of carvacrol and thymol, and they are natural monoterpenes that act on the cell membrane of the organisms causing cellular death.[16]

Curcumin is a plant-derived agent (turmeric root), which also exhibits antioxidant and anticancer effects, thus having significant clinical relevance related to the prevention and treatment of numerous illnesses. Curcumin has already been used in the fabrication of electrospun fibres for biomedical applications (e.g., skin tissue regeneration), and more recently, it was employed as an intracanal irrigant during endodontic treatment, showing effective and promising disinfection results, probably explained by its permeabilization effects that cause damage of bacterial membranes.

Thus, the aim of present study is to compare antimicrobial properties of various biological extract Nisin, curcumin, aloe vera as an intracanal medicament when compared with calcium hydroxide.

SUBJECTS AND METHODS

The medicaments used in the study were calcium hydroxide powder, Nisin (2.5%, Sigma Aldrich, USA), curcumin powder was dissolved in ethanol to obtain a stock solution (100 mg/mL). Materials used were absolute ethanol, brain heart infusion broth, trypticase soy blood agar plates, and *E. faecalis* (MTCC-439) strain.

Literature survey

1. Study of various articles related to bacterial leakage.
2. Study in detail about biomechanical preparation, intracanal medicaments and various herbal extracts and its uses.
3. The existing approaches to perform degree of infection in root canal wall.

Simulation

1. Effect of various intracanal medicaments in root canal treatment.
2. Composition of herbal extracts.
3. Analysis of the leakage values

Experimental setup

Single canal roots, morphological similarities with no previous root canal treatment taken.

Complete biomechanical preparation will be done, which will be followed by inoculation of *E. Faecalis*.

After inoculation, the samples will be kept in a closed Eppendorf tube and incubated at 37°C for 21 days under aseptic conditions. The canals will be re-inoculated with fresh bacterial samples at every 3 days interval to ensure viability of bacteria.

The canal contents will be aspirated after 21 days of incubation, then rinsed with 5 mL saline and patted dry with sterile paper points. The specimens will be then randomly divided into following groups:

Group 1: Calcium hydroxide

Group 2: Aloe vera

Group 3: Curcumin

Group 4: Nisin

Study and comparison

Eighty single-rooted lower premolar teeth which will be extracted for orthodontic purpose will be collected. Tooth specimens will be sectioned at cement-enamel junction with a diamond saw to obtain a standard root length. The specimens will be randomly divided into four groups (n = 20). The working length measurement will be done by measuring file length 1 mm less until tip of file was visible at the apical foramen. To standardize, the root will be cut to make working length as 10 mm. The root canals of the specimen will be instrumented with Protaper rotary files. After that copious intracanal irrigation will be done with 2.5% sodium hypochlorite and ethylenediamine tetra acetic acid (17% w/v) using 5 mL luer lock

syringe followed by irrigating with 0.9% normal saline solution (Marck Biosciences, India) to remove smear layer.

Root Canal infection-

- Each root canal will be inoculated with 24 h old cultured broths of the bacterial solution up to the canal orifice using a sterile endodontic needle in a microbiological safety cabinet.
- After inoculation, the samples will be kept in a cotton plugged test tube and incubated at 37°C for 21 days. Every 3rd day, the canals were reinoculated with fresh bacterial samples. The canal contents will then be aspirated after 21 days of incubation, then rinsed with 5 mL saline and patted dry with sterile paper points. The specimens will then be randomly divided into four groups (n = 20 each) for intracanal medicaments:
- The specimens will be divided into four groups which will be as follows-
 - Group 1: Calcium hydroxide
 - Group 2: Aloe vera
 - Group 3: Curcumin
 - Group 4: Nisin
- Preparation of calcium hydroxide mix-calcium hydroxide intracanal medicament is prepared by mixing powder with normal saline in the ratio of 1:1 to obtain the paste.
- Aloe vera gel (100% Aloe vera gel, Sillaneh Co., Iran).
- Preparation of aqueous Nisin - 200 mg/mL concentration of Nisin was prepared by dissolving it in distilled water.
- In all the samples, the prepared medicaments of 5 µL will be injected in the root canals and completely filled. The canals will then be sealed with sticky wax (Pyrax, India) and incubated at 37°C for 7 days. After 7 days of incubation, the wax will be removed from each of the canal entrances. The bacterial samples from each canal will be retrieved with sterile paper points and inoculated on brain-heart infusion broth (HiMedia Laboratories, India) and incubated at 37°C for 24 h. After that, each canal will be irrigated with 5 mL of saline and sterile paper points will be used to dry the canal.
- To evaluate the degree of infection of the canal wall and its radicular dentine, specimens of dentine chips will be retrieved. The dentin chips will be then transferred by placing files into 0.5 mL of brain-heart infusion broth through sterile Eppendorf tubes and incubated at 37°C for 24 h. After 24 h, 5 µL. of solution will be inoculated on TSY-blood agar plates from each tube and incubated at 37°C for 24 h to obtain bacterial colony forming unit (CFU) count. Statistical analysis will be done using Kruskal Wallis test

RESULTS

The means CFUs count of the present study showed Nisin with no CFUs of *E. faecalis*. Curcumin, and Aloe vera can be used as an effective alternate intracanal medicament and had low CFU count as compared to calcium hydroxide alone. Statistically significant

difference was present in CFU count when different medicaments used against *E. faecalis* by Kruskal–Wallis test.

DISCUSSION

Formerly since a decade or more intracanal medicament had been used as an interim appointment dressing. The commonly used commercial synthetic medicaments are calcium hydroxide, phenolic compounds (eugenol and camphorated monochlorophenol), aldehydes (formocresol), halides (iodine potassium iodide), antibiotics, etc.[2] However, majority had been reported for its toxic effect, development of resistant strains and depletion of immune response. This had led the shift in paradigm from synthetic to naturally derived medicaments.

E. faecalis is a Gram-positive coccus, been a facultative anaerobic, it can penetrate deeper inside the dentinal tubules which could possibly result in root canal reinfection.[17] It resists chemico-mechanical instrumentation and can survive as a monoculture inside the root canals without any mutualism with other bacteria. *E. faecalis* can even withstand high alkaline pH during calcium hydroxide dressing.[18] Thus, rendering a need of an alternative method for eradicating this species and will be beneficial for the prognosis of endodontic treatment.

The present study showed Nisin as the most effective medicament against *E. faecalis* as there were none of the CFUs with Nisin. Nisin is an antimicrobial peptide, naturally occurring produced by *Streptococcus lactis*. It is commonly used as a food preservative in meat and dairy industry. Previously, studies had reported the antimicrobial activity of Nisin against *E. faecalis* both *in vitro* and *in vivo*. [12] Nisin exerts its bactericidal activity through pores formation by interacting with a specific molecule “Lipid II” and inhibit cell wall synthesis. The principal component of Gram-positive bacterial cell membrane is Lipid II. At a nanomolecular level, Nisin targets this Lipid II as a “docking molecule” to form pores on the cell membrane surface and effectively kills the bacteria.[19,20] Turner *et al.* showed significantly lower infected dentinal shaving of *E. faecalis* with Nisin when compared to calcium hydroxide.[11]

Calcium hydroxide was ineffective against *E. faecalis* in the present study. The results were in accordance with the studies done by Harrison *et al.* and Hemadri *et al.* [21,22] *E. faecalis* is resistant to highly alkaline environment due to the presence of proton pump as a primary resistance mechanism.[23] At pH of 11.5 *E. faecalis* cannot survive, however calcium hydroxide medicament results in alkalinity of pH 10.3 within radicular dentine as being reported *in vitro*. [24] This is due to buffering capacity of dentine which provides a decreasing pH from inner to peripheral root dentine.[23]

REFERENCES

1. Akpata ES, Blechman H. Bacterial invasion of pulpal dentin wall *in vitro*. J Dent Res 1982;61:435-8.

2. Gomes BP, Souza SF, Ferraz CC, Teixeira FB, Zaia AA, Valdrighi L, *et al.* Effectiveness of 2% chlorhexidine gel and calcium hydroxide against *Enterococcus faecalis* in bovine root dentine *in vitro*. Int Endod J 2003;36:267-75.
3. Murray PE, Farber RM, Namerow KN, Kuttler S, Garcia-Godoy F. Evaluation of *Morindacitrifolia* as an endodontic irrigant. J Endod 2008;34:66-70.
4. Wang Z, Shen Y, Haapasalo M. Effectiveness of endodontic disinfecting solutions against young and old *Enterococcus faecalis* biofilms in dentin canals. J Endod 2012;38:1376-9.
5. Siqueira JF Jr. Aetiology of root canal treatment failure: Why well-treated teeth can fail. Int Endod J 2001;34:1-10.
6. Haapasalo M, Orstavik D. *In vitro* infection and disinfection of dentinal tubules. J Dent Res 1987;66:1375-9.
7. Evans MD, Baumgartner JC, Khemaleelakul SU, Xia T. Efficacy of calcium hydroxide: Chlorhexidine paste as an intracanal medication in bovine dentin. J Endod 2003;29:338-9.
8. Siqueira JF Jr., Lopes HP. Mechanisms of antimicrobial activity of calcium hydroxide: A critical review. Int Endod J 1999;32:361-9.
9. Sadr Lahijani MS, Raoof Kateb HR, Heady R, Yazdani D. The effect of German chamomile (*Matricaria recutita* L.) extract and tea tree (*Melaleuca alternifolia* L.) oil used as irrigants on removal of smear layer: A scanning electron microscopy study. Int Endod J 2006;39:190-5.
10. Severina E, Severin A, Tomasz A. Antibacterial efficacy of nisin against multidrug-resistant Gram-positive pathogens. J Antimicrob Chemother 1998;41:341-7.
11. Turner SR, Love RM, Lyons KM. An *in-vitro* investigation of the antibacterial effect of nisin in root canals and canal wall radicular dentine. Int Endod J 2004;37:664-71.
12. Newall CA, Anderson LA, Phillipson JD. Herbal Medicines. A Guide for Health-Care Professionals. London: The Pharmaceutical Press; 1996.
13. Arunkumar S, Muthuselvam M. Analysis of phytochemical constituents and antimicrobial activities of *Aloe vera* L. against clinical pathogens. World J Agric Sci 2009;5:572-6.
14. Thiruppathi S, Ramasubramanian V, Sivakumar T, Thirumalaivasu V. Antimicrobial activity of *Aloe vera* (L.) Burm. f. against pathogenic microorganisms. J Biosci Res 2010;1:251-8.
15. Bhardwaj A, Ballal S, Velmurugan N. Comparative evaluation of the antimicrobial activity of natural extracts of *Morindacitrifolia*, papain and *Aloe vera* (all in gel formulation), 2% chlorhexidine gel and calcium hydroxide, against *Enterococcus faecalis*: An *in vitro* study. J Conserv Dent 2012;15:293-7.
16. Abbaszadegan A, Sahebi S, Gholami A, Delroba A, Kiani A, Iraj A, *et al.* Time-dependent antibacterial effects of *Aloe vera* and *Zataria multiflora* plant essential oils compared to calcium hydroxide in teeth infected with *Enterococcus faecalis*. J Investig Clin Dent 2016;7:93-101.
17. de Lucena JM, Decker EM, Walter C, Boeira LS, Löst C, Weiger R. Antimicrobial effectiveness of intracanal medicaments on *Enterococcus faecalis*: Chlorhexidine versus octenidine. Int Endod J 2013;46:53-61.

18. Evans M, Davies JK, Sundqvist G, Figdor D. Mechanisms involved in the resistance of *Enterococcus faecalis* to calcium hydroxide. Int Endod J 2002;35:221-8.
19. Driessen AJ, van den Hooven HW, Kuiper W, van de Kamp M, Sahl HG, Konings RN, *et al.* Mechanistic studies of antibiotic-induced permeabilization of phospholipid vesicles. Biochemistry 1995;34:1606-14.
20. Ohara P, Torabinejad M, Kettering JD. Antibacterial effects of various endodontic medicaments on selected anaerobic bacteria. J Endod 1993;19:498-500.
21. Hemadri M, Sophia T, Sajjan G. Nisin vs calcium hydroxide – Antibacterial efficacy on *Enterococcus faecalis* – An *in-vitro* study. IJCD 2011;2:55-61
22. Harrison JW, Bellizzi R, Osetek EM. The clinical toxicity of endodontic medicaments. J Endod 1979;5:42-7.
23. Love RM. *Enterococcus faecalis* – A mechanism for its role in endodontic failure. Int Endod J 2001;34:399-405.
24. Nerwich A, Figdor D, Messer HH. pH changes in root dentin over a 4-week period following root canal dressing with calcium hydroxide. J Endod 1993;19:302-6