

## Calcium and Ferrous ion crosstalk driving dopaminergic loss in patients with Parkinsons disease - a systematic review .

Dr. Arijit Mazumdar

Assistant Professor

Department of Physiology

P A Sangma International Medical College, USTM Campus

Baridua, Ri Bhoi, Meghalaya.

Dr. Arijit Mazumdar

Email: [mazumdararijit17@gmail.com](mailto:mazumdararijit17@gmail.com)

Cell: 9401449750

### Abstract:

Calcium and iron are vital trace elements that orchestrate critical eukaryotic cellular processes, from signaling and enzyme activity to energy metabolism. Disruptions in their homeostasis trigger cell dysfunction, oxidative damage, and eventual death. Parkinson's disease (PD), the second most prevalent neurodegenerative disorder worldwide, lacks curative therapies or interventions to halt its inexorable progression. Its pathological signatures—selective death of dopaminergic neurons in the substantia nigra and Lewy body formation via  $\alpha$ -synuclein aggregation—closely intertwine with dysregulated calcium ( $\text{Ca}^{2+}$ ) and iron homeostasis. This review elucidates how aberrant  $\text{Ca}^{2+}$  signaling exacerbates PD pathogenesis. Excessive cytosolic  $\text{Ca}^{2+}$  influx through voltage-gated channels (e.g., L-type VGCCs, NMDA receptors) or release from intracellular stores (mitochondrial MCU, ER RyRs/IP3Rs) overloads mitochondria, impairs ATP production, and activates catabolic pathways like calpains and caspases. This fosters  $\alpha$ -synuclein misfolding, synaptic dysfunction, and dopaminergic neurotoxicity, amplified by elevated dopamine auto-oxidation in high- $\text{Ca}^{2+}$  milieus. Concomitantly, iron dysregulation drives ferroptosis, an iron-dependent lipid peroxidation form of regulated cell death central to PD. Excess labile ferrous iron ( $\text{Fe}^{2+}$ ) in the basal ganglia—evident via MRI hypointensities—catalyzes Fenton reactions, generating hydroxyl radicals that peroxidise polyunsaturated fatty acids in neuronal membranes. Glutathione peroxidase 4 (GPX4) depletion, often via  $\alpha$ -synuclein-mediated ferroportin suppression or xCT inhibition, fails to neutralize these lipids, culminating in ferroptotic collapse. The  $\text{Ca}^{2+}$ - $\text{Fe}^{2+}$  nexus worsens this: calcium promotes iron release from ferritin, while both ions synergize in mitochondrial permeability transition pores, accelerating ROS bursts. Deciphering these intertwined imbalances is paramount for devising disease-modifying strategies, such as MCU inhibitors (e.g., Ru360), iron chelators (deferiprone), or ferroptosis blockers (ferrostatin-1), potentially synergizing to preserve dopaminergic integrity and mitigate PD progression. Parkinson's disease (PD) ranks as the second most common neurodegenerative disorder after Alzheimer's, affecting millions worldwide as populations age and lifespans extend. Its progressive nature draws intense research focus due to the growing burden on healthcare systems

**Keywords:** Parkinson's disease, Calcium homeostasis, Iron dysregulation, Ferroptosis,  $\alpha$ -Synuclein aggregation.

**Introduction:**

Parkinson's disease (PD), the second most common neurodegenerative disorder after Alzheimer's, commands intense research scrutiny amid a global aging crisis that drives its escalating prevalence.<sup>1</sup> As lifespans lengthen, PD's relentless progression—from motor deficits to cognitive decline<sup>2</sup>—strains healthcare systems worldwide.<sup>3</sup> Aging emerges as the dominant risk factor, heightening vulnerability in substantia nigra dopaminergic neurons, while genetic variants (e.g., SNCA, LRRK2) and environmental insults like pesticides or trauma contribute modestly.<sup>4</sup> These explain only a fraction of cases; most PD likely stems from their intricate interplay, spotlighting downstream cascades like metal dyshomeostasis.<sup>5</sup> Though trace elements, iron and calcium prove indispensable for oxygen handling, enzymatic catalysis, and signaling.<sup>6</sup> Their central nervous system overload sparks oxidative stress through reactive oxygen species, mitochondrial collapse, and protein/receptor wreckage.<sup>7</sup> Metal imbalance fuels neurodegeneration, disability, and neuroinflammation across disorders like PD and Alzheimer's.<sup>8</sup> Compelling data pinpoint calcium overload and iron deposition as PD linchpins. Basal ganglia iron buildup—evident on MRI—ignites Fenton chemistry, yielding hydroxyl radicals<sup>9</sup> that peroxidise lipids and trigger ferroptosis.<sup>10</sup> Concurrently, cytosolic Ca<sup>2+</sup> surges via L-type channels or MCU/IP3R<sup>11</sup> stores overwhelm mitochondria, exacerbate  $\alpha$ -synuclein aggregation,<sup>12</sup> and amplify dopaminergic toxicity via dopamine oxidation.<sup>13</sup> This review dissects the Ca<sup>2+</sup>-Fe<sup>2+</sup> axis in PD pathogenesis, probing therapeutic horizons.<sup>14</sup> Iron chelators (deferiprone), MCU blockers (Ru360), and ferroptosis inhibitors (ferrostatin-1) show preclinical promise, potentially synergizing with antioxidants to restore homeostasis,<sup>15</sup> forestall neuron loss, and pioneer disease-modifying interventions beyond symptomatic palliation.<sup>16,17</sup>

**Calcium Dysregulation in Parkinson's Disease**

Calcium ions (Ca<sup>2+</sup>) function as a universal second messenger across living organisms, enabling cells to sense and respond to environmental shifts.<sup>18</sup> In resting cells, cytoplasmic Ca<sup>2+</sup> remains tightly clamped at ~100 nmol/L, contrasting sharply with endoplasmic reticulum stores (0.5–1 mmol/L) and extracellular levels (1–2 mmol/L)—a 20,000-fold gradient that powers rapid signaling transients.<sup>19</sup> This steep disparity lets cells harness Ca<sup>2+</sup> pulses to toggle physiological states via downstream cascades regulating contraction, secretion, and transcription.<sup>20</sup> Neurons exploit this through plasma membrane fluxes: depolarization opens voltage-gated calcium channels (VGCCs), while agonists like the dihydropyridine BAY K8644 amplify L-type VGCC influx.<sup>21</sup> Since Ca<sup>2+</sup> sculpts every facet of neuronal biology—from excitability to survival—homeostasis demands exquisite precision, encompassing cytosolic levels, microdomain dynamics, buffering capacity, and entry kinetics.<sup>22</sup>

Substantia nigra dopaminergic (SN DA) neurons, prime targets in Parkinson's disease (PD), rely heavily on Ca<sup>2+</sup> for excitability,<sup>23</sup> dopamine release, mitochondrial ATP synthesis, enzymatic control, gene expression, and apoptosis thresholds.<sup>24,25</sup> Calcium mishandling accelerates aging-related neurodegeneration, disrupting neurogenesis, synaptic plasticity, and neurotransmission.<sup>26,27</sup> PD patients display elevated brain Ca<sup>2+</sup> versus controls, fuelling excessive dopamine synthesis and autotoxicity that precipitates dopaminergic demise.<sup>28,29,30</sup> Alpha-synuclein aggregates, PD's pathological signature, further derail calcium clearance and signaling.<sup>31</sup>

Regulation spans plasma membrane routes—VGCCs, NMDA/AMPA receptors<sup>32</sup>—and intracellular depots: mitochondrial calcium uniporter (MCU), endoplasmic reticulum RyRs/IP3Rs, plus store-operated systems like STIM/Orai and TRP channels.<sup>33,34</sup> In PD, these falter, unleashing cytosolic overloads that trigger mitochondrial collapse, ROS bursts, and proteinopathy<sup>35,36</sup>—cascades ripe for therapeutic interception via channel modulators or buffers.<sup>37</sup>

### Calcium Signaling and Homeostasis in Parkinson's Disease Neurons

Calcium ions ( $\text{Ca}^{2+}$ ) serve as the universal second messenger across all living organisms, enabling cells to detect and respond to environmental changes with remarkable precision.<sup>38</sup> In resting cells, cytoplasmic  $\text{Ca}^{2+}$  concentration remains clamped at approximately 100 nmol/L, while endoplasmic reticulum (ER) stores maintain 0.5–1 mmol/L and extracellular fluid holds 1–2 mmol/L.<sup>39,40</sup> This creates a staggering 20,000-fold gradient across the plasma membrane, allowing cells to generate rapid, localized  $\text{Ca}^{2+}$  transients that function as potent signaling cues.<sup>41</sup> These transients activate or inhibit  $\text{Ca}^{2+}$ -dependent pathways, orchestrating diverse physiological responses from contraction and secretion to gene transcription.<sup>42</sup>

In neurons,  $\text{Ca}^{2+}$  entry occurs primarily through plasma membrane channels triggered by electrical depolarization or chemical agonists.<sup>43</sup> The dihydropyridine derivative BAY K8644 exemplifies this by enhancing influx through L-type voltage-gated calcium channels (VGCCs).<sup>44</sup> Given  $\text{Ca}^{2+}$ 's pervasive influence on neuronal biology—spanning excitability, synaptic transmission, survival,<sup>45</sup> and death—maintaining precise homeostasis proves essential.<sup>46</sup> This involves tight regulation of cytosolic levels, microdomain formation, buffering capacity, and spatiotemporal entry patterns.<sup>47</sup>

Substantia nigra dopaminergic (SN DA) neurons, selectively vulnerable in Parkinson's disease (PD), depend critically on  $\text{Ca}^{2+}$  for multiple functions: membrane excitability, dopamine release, mitochondrial ATP production, enzymatic regulation, gene expression, and apoptosis execution.<sup>48,49</sup> Calcium dysregulation accelerates aging-related neurodegeneration by disrupting neurogenesis,<sup>50</sup> synaptic plasticity, and neurotransmission.<sup>51</sup> PD patients consistently show elevated brain  $\text{Ca}^{2+}$  levels compared to healthy controls, linking this overload to excessive dopamine biosynthesis and subsequent dopaminergic neurotoxicity.<sup>52,53,54</sup>

Alpha-synuclein aggregation, a pathological hallmark of PD, further compromises calcium homeostasis by impairing clearance mechanisms.<sup>55,56</sup> Multiple pathways govern  $\text{Ca}^{2+}$  fluxes in SN DA neurons.<sup>57</sup> Plasma membrane entry occurs via VGCCs and glutamate receptors (NMDA-R, AMPA-R). Intracellularly, ER releases  $\text{Ca}^{2+}$  through ryanodine receptors (RyR) and IP<sub>3</sub> receptors,<sup>58,59,60</sup> while mitochondria contribute via the mitochondrial calcium uniporter (MCU).<sup>61</sup> Store-operated calcium entry (SOCE) systems—STIM/Orai complexes and transient receptor potential (TRP) channels—replenish cytosolic stores after depletion.<sup>62</sup>

In PD, these regulatory systems fail, unleashing pathological  $\text{Ca}^{2+}$  surges that trigger mitochondrial dysfunction,<sup>63</sup> oxidative stress cascades, and protein misfolding<sup>64</sup>—driving inexorable dopaminergic degeneration characteristic of the disease.<sup>65,66</sup>

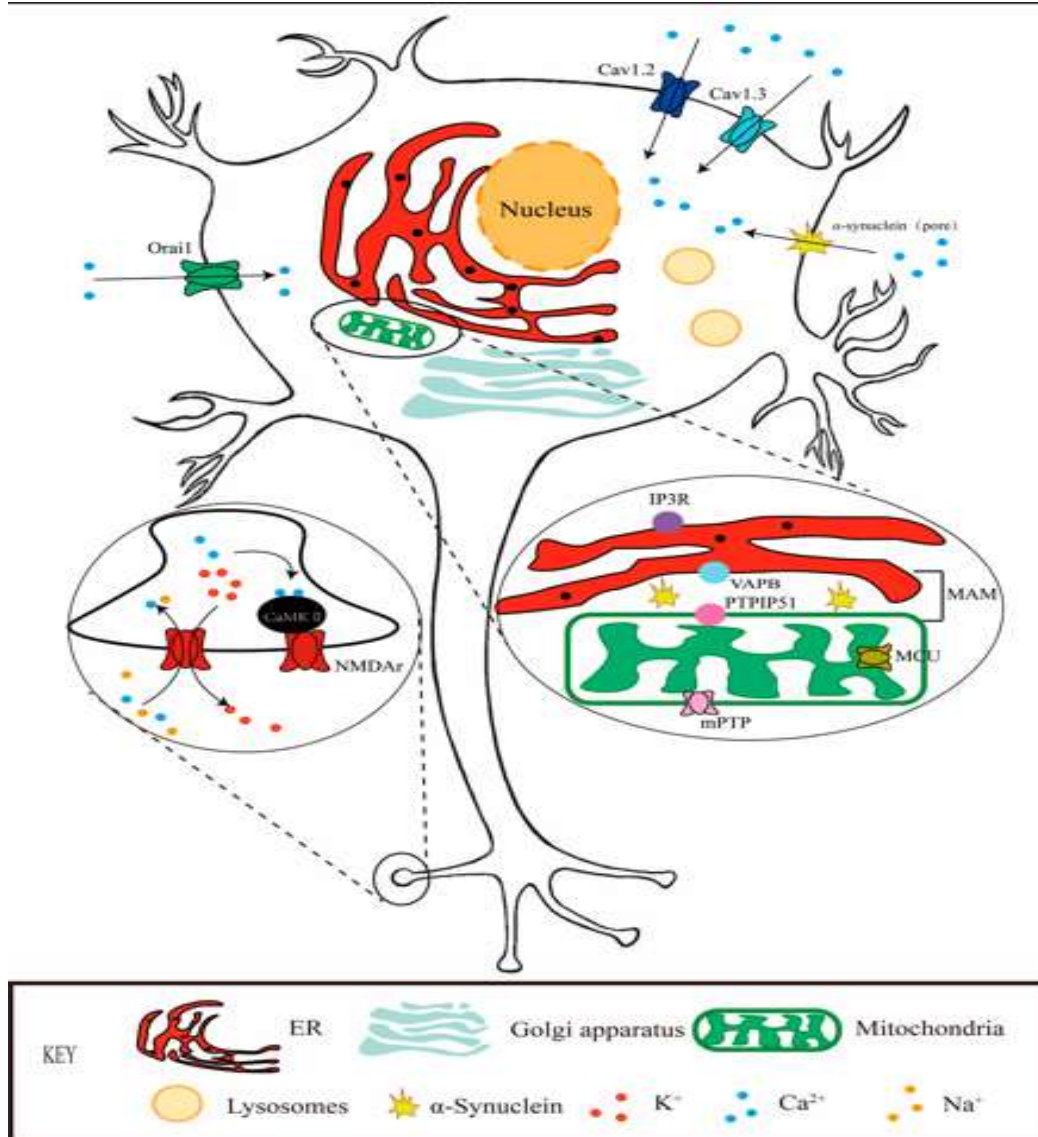


Fig 1: Neuronal  $\text{Ca}^{2+}$  Chaos: Entry Routes, ER-Mito Crosstalk and Overload Triggering Apoptosis"

Voltage-gated calcium channels (VGCCs) comprise ten distinct subtypes,<sup>67</sup> classified by their unique pharmacological profiles and pore-forming  $\alpha 1$  subunits, with function fine-tuned through alternative splicing and accessory subunits ( $\beta 1$ – $\beta 4$ ,  $\alpha 2\delta 1$ – $\alpha 2\delta 4$ ).<sup>68</sup> These channels cluster into three families based on sequence homology and properties: Cav1 (L-type), Cav2 (P/Q-, N-, R-type), and Cav3 (T-type).<sup>69,70</sup> The Cav1 family includes four L-type channels (Cav1.1–Cav1.4), highly sensitive to dihydropyridine (DHP) blockers at nanomolar concentrations.<sup>71</sup> In Parkinson's disease (PD), Cav1.2 and Cav1.3 emerge as key players, particularly in substantia nigra pars compacta (SNc) dopaminergic neurons.<sup>72</sup>

Juvenile SNc neurons predominantly express Cav1.2, but aging shifts reliance to Cav1.3, which supports oscillatory  $\text{Ca}^{2+}$  influx critical for autonomous pacing—the intrinsic,<sup>73,74</sup> slow depolarization sustaining basal dopamine release to the striatum.<sup>75</sup> Unlike Cav1.2, Cav1.3 channels resist full closure during pacing cycles, maintaining elevated cytosolic  $\text{Ca}^{2+}$  essential for physiologic dopamine secretion.<sup>77,78</sup> However, chronic  $\text{Ca}^{2+}$  excess synergizes with aging,

mitochondrial toxins, or genetic mutations, precipitating metabolic stress and organelle damage.<sup>79</sup> Notably, upregulated Cav1.3 expressions in early PD cerebral cortex precedes pathology, positioning calcium dysregulation as an initiator rather than mere consequence. DHP antihypertensives offer neuroprotection; epidemiological data show 20–30% reduced PD risk with chronic use.<sup>80</sup>

Cav2 family channels (Cav2.1–2.3) localize presynaptically, driving rapid neurotransmitter release via P/Q-, N-, and R-type currents.<sup>81</sup> T-type Cav3 channels (Cav3.1–3.3) activate at hyperpolarized potentials, shaping neuronal firing patterns through subthreshold oscillations.<sup>82,83</sup>

Dopaminergic neuron activity hinges on Ca<sup>2+</sup> carriers and homeostasis, with ~80% of cellular Ca<sup>2+</sup> sequestered in organelles.<sup>84,85,86</sup> While endoplasmic reticulum (ER) dominates as the primary reservoir, mitochondria, lysosomes, and Golgi also contribute significantly.<sup>87,88</sup> ER-mediated signaling proves pivotal, particularly store-operated Ca<sup>2+</sup> entry (SOCE).<sup>89</sup> ER Ca<sup>2+</sup> depletion activates STIM1 on the ER membrane, which couples to plasma membrane Orai1 channels, driving extracellular Ca<sup>2+</sup> influx that SERCA pumps refill into ER.<sup>90,91</sup> STIM1 governs classical SOCE, while lower-affinity STIM2 fine-tunes subtle fluctuations. Orai1/STIM1 mutations disrupt SOCE, linking to diverse pathologies.<sup>92,93</sup>

In PD, STIM1-TRPC1 complexes inhibit Cav1.3, deranging homeostasis, while reduced STIM1 occurs in Alzheimer's disease (AD).<sup>94</sup> Hippocampal STIM2 supports dendritogenesis via Ca<sup>2+</sup>/calmodulin-dependent kinase pathways.<sup>95</sup> Synaptotagmin-7 (Syt7), a presynaptic Ca<sup>2+</sup> sensor, amplifies spontaneous release through STIM2-mediated SOCE.<sup>96,97</sup> Chronic hyperactivity risks synaptic exhaustion and apoptosis, implicating presynaptic SOCE in neurodegeneration across PD, AD, and beyond.<sup>99,100</sup>

Mitochondrial Ca<sup>2+</sup> handling centers on ER-mitochondria associated membranes (MAMs), where ~20% of mitochondria tether ~10–25 nm from ER.<sup>101</sup> IP3R on ER releases Ca<sup>2+</sup> into cytosol, funnelled through VDAC1 on mitochondrial outer membrane into the matrix via MCU.<sup>102,103</sup> Pathologic VDAC1 upregulation triggers oligomerization, MCU hyperactivation, and MAM coupling to IP3R, culminating in Ca<sup>2+</sup> overload.<sup>104</sup> This opens mitochondrial permeability transition pore (mPTP) or promotes Bax/Bak pore formation, releasing cytochrome c to trigger caspase-9/3-mediated apoptosis.<sup>105,106</sup>

Paradoxically,  $\alpha$ -synuclein at MAMs disrupts VAPB-PTPIP51 tethers—whether wild-type or mutant—impairing ER-mito Ca<sup>2+</sup> transfer and ATP synthesis via tricarboxylic acid cycle dysregulation.<sup>107,108</sup> Both Ca<sup>2+</sup> excess (mPTP opening) and deficiency (energy failure) threaten neuronal survival.<sup>109</sup>

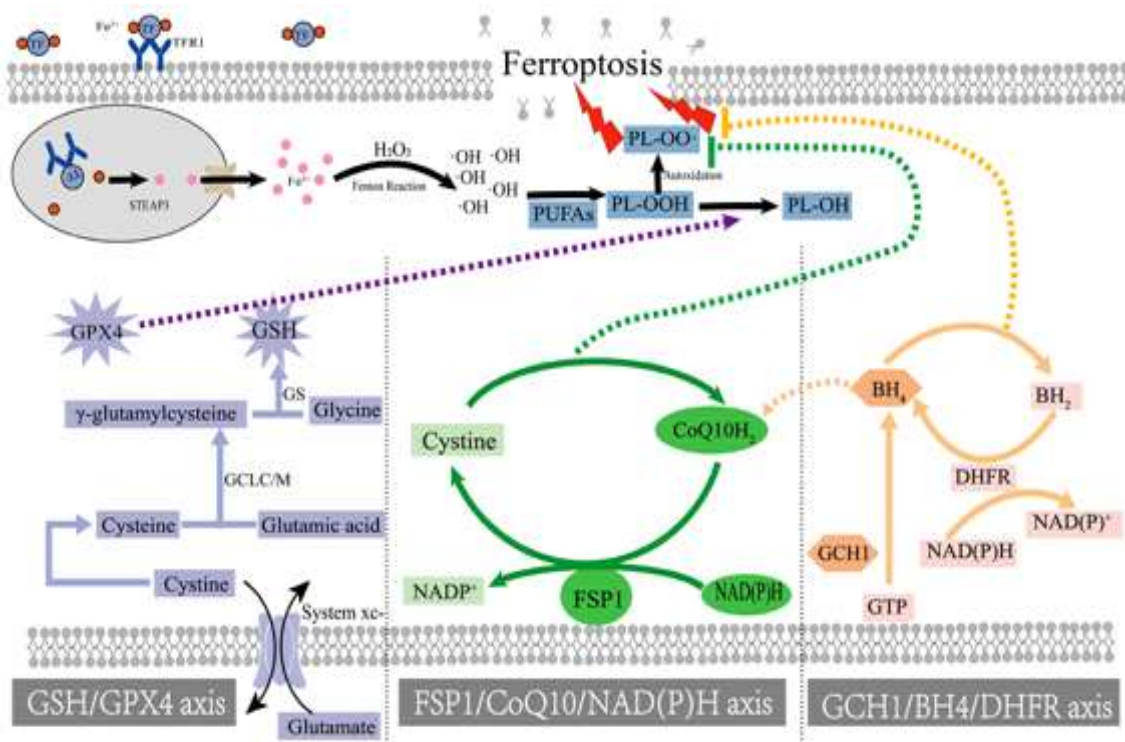
Ca<sup>2+</sup> signalling's ubiquity positions dyshomeostasis as a convergent aging mechanism across neurodegeneration.<sup>110</sup> Plasma membrane influx, intracellular buffering, and inter-organelle trafficking orchestrate metabolism, signaling, and survival.<sup>111</sup> Targeting VGCCs (DHPs), SOCE (STIM/Orai modulators), or MAM integrity holds transformative therapeutic promise for PD.<sup>112</sup>

### **Role of Iron in Parkinson's disease**

Iron serves as an essential trace element fuelling cellular metabolism, with outsized importance in the central nervous system where it supports synapse formation, myelination, and neurotransmitter synthesis and release.<sup>113,114</sup> These roles extend to childhood brain development, influencing IQ, cognition, motor skills, and social behavior.<sup>115,116</sup> However, brain ferritin levels rise with age, driving iron overload that contributes to neurodegeneration in older adults.<sup>117,120</sup>

Excess iron sparks oxidative stress, inflammation, and cell death, with elevated levels consistently detected in Parkinson's disease (PD)<sup>119</sup> and Alzheimer's disease (AD) brains,<sup>120</sup> particularly in basal ganglia and substantia nigra. As a redox-active metal, iron powers ATP production in neurons, but overload renders neural tissue vulnerable to oxidative damage and proteolysis.<sup>121</sup> Beyond aging, hereditary hemochromatosis (HH)—an autosomal recessive disorder from HFE gene mutations (C282Y, H63D) disrupting hepcidin regulation—accelerates brain iron deposition.<sup>122,123</sup> Though the blood-brain barrier normally shields the CNS during systemic imbalance, overload compromises this defence, potentially elevating PD risk in HH patients through dysregulated storage and export.<sup>124</sup> Iron dysregulation culminates in ferroptosis, a caspase-independent regulated cell death (RCD) form distinct from apoptosis, necrosis, or autophagy.<sup>129,126</sup> First described by Brent Stockwell, ferroptosis features shrunken mitochondria with reduced cristae density, outer membrane rupture, and no plasma membrane lysis or bioenergetic catastrophe. In neurodegeneration, it unleashes inflammation, neurotransmitter oxidation, synaptic failure, myelin breakdown, astrocyte dysfunction, and neuronal demise.<sup>125</sup>

Ferroptosis triggers via glutamate/iron/polyunsaturated fatty acid (PUFA) buildup or glutathione (GSH)/NAD(P)H/glutathione peroxidase 4 (GPX4) depletion. Iron uptake begins with transferrin receptor 1 (TFR1) binding, endosomal endocytosis, ferric ( $\text{Fe}^{3+}$ ) reduction by STEAP3, and ferrous ( $\text{Fe}^{2+}$ ) export via DMT1 into the labile iron pool (LIP) or ferritin storage.<sup>127</sup> Excess LIP  $\text{Fe}^{2+}$  catalyzes Fenton reactions, generating hydroxyl radicals that initiate PUFA peroxidation in plasma membranes.<sup>128</sup> Lipid hydroperoxides (PL-OOH) decompose into toxic aldehydes, epoxides, and oxo products, forming nanomembrane pores that rupture cells.<sup>130</sup> This peroxidation propagates cell-to-cell in waves, perpetuating iron-lipid vicious cycles.<sup>131</sup> Three anti-ferroptotic pathways counter this: the canonical GSH/GPX4 axis neutralizing peroxides; the FSP1/coenzyme Q10/NAD(P)H system; and the GCH1/bh4/DHFR route.<sup>132</sup> In PD, basal ganglia iron excess—visible as MRI hypointensities—amplifies dopaminergic ferroptosis, positioning iron chelators like deferiprone and ferroptosis inhibitors (ferrostatin-1) as promising disease-modifying strategies.<sup>133</sup>



**Figure 2. Anti-Ferroptotic Pathways:** TFR1-STEAP3 iron uptake → Fenton-driven PUFA peroxidation countered by GSH/GPX4, FSP1/CoQ10, and GCH1/BH4/DHFR radical scavenging.

### Ferroptosis Defence Pathways: GPX4, FSP1/CoQ10, and GCH1/BH4 Systems

Glutathione peroxidase 4 (GPX4) stands as the cornerstone selenoenzyme suppressing ferroptosis, the iron-dependent lipid peroxidation cell death. Physiologically, GPX4 harnesses glutathione (GSH) to reduce cytotoxic lipid hydroperoxides (PL-OOH) to harmless alcohols (PL-OH), directly countering the Fenton reaction-driven membrane damage in Parkinson's disease (PD). GSH depletion or GPX4 inactivation unleashes ferroptotic cascades in dopaminergic neurons.<sup>134,135,136</sup>

GSH biosynthesis hinges on the cystine/glutamate antiporter system  $x_c^-$ , which imports cystine for reduction to cysteine by GSH itself or thioredoxin reductase 1 (TRR1).<sup>137</sup> ATP-dependent glutamate-cysteine ligase (GCL) then forms  $\gamma$ -glutamylcysteine, followed by glutathione synthetase (GS) addition of glycine to yield GSH.<sup>138</sup> Transcription factors orchestrate this: Nrf2 (nuclear factor erythroid 2-related factor 2) upregulates GCL, GS, GCLC (GCL catalytic subunit), GSTs (glutathione S-transferases), HO-1 (heme oxygenase-1), and NQO1 (NAD(P)H quinone dehydrogenase 1), mounting a broad antioxidant defence.<sup>139</sup> Nrf2 modulation emerges as a promising neuroprotective strategy for PD, potentially amplifying GSH/GPX4 resilience against iron overload.<sup>140</sup>

Parallel to this canonical axis operates the FSP1-CoQ10-NAD(P)H pathway, a GSH-independent ferroptosis suppressor.<sup>140</sup> Ferroptosis suppressor protein 1 (FSP1) features N-myristoylation for membrane targeting and a flavoprotein oxidoreductase domain. FSP1 reduces coenzyme Q10 (CoQ10)—the lipophilic mitochondrial electron carrier—to ubiquinol

(CoQ10-H2) using NAD(P)H.<sup>141</sup> Both oxidized and reduced CoQ10 scavenge lipid peroxides at plasma membranes, safeguarding PUFA-rich phospholipids.<sup>142</sup> NQO1 synergizes with FSP1, further reducing ubiquinone to ubiquinol, with Nrf2 governing NQO1 expression.<sup>143</sup> This system's potency rivals GPX4/GSH, positioning FSP1/CoQ10 activators as viable PD therapeutics.<sup>144</sup>

The GCH1/BH4/DHFR pathway, though underexplored in neurodegeneration, offers a third defence line. Tetrahydrobiopterin (BH4)—essential for dopamine biosynthesis—traps lipid peroxidation radicals while boosting CoQ10 synthesis via phenylalanine hydroxylase.<sup>145</sup> GTP cyclohydrolase 1 (GCH1) rate-limits de novo BH4 production; dihydrofolate reductase (DHFR) recycles oxidized BH4.<sup>146</sup> DHFR inhibition synergizes with GPX4 blockers to induce ferroptosis therapeutically,<sup>147</sup> but BH4 supplementation may conversely protect neurons. GCH1/BH4's role in PD ferroptosis warrants deeper investigation given BH4's dopamine linkage.<sup>148</sup>

**Therapeutic Horizons**

Ferroptosis inhibition offers disease-modifying potential for PD, targeting iron dysregulation at multiple nodes. Iron chelators like deferiprone<sup>149</sup> reduce labile Fe<sup>2+</sup> pools, blunting Fenton chemistry.<sup>150</sup> Nrf2 agonists (e.g., dimethyl fumarate) amplify GSH/GPX4 and NQO1 defenses. GPX4 stabilizers or GSH precursors (N-acetylcysteine) restore canonical protection.<sup>151</sup> CoQ10 supplementation—already trialled in PD—leverages FSP1 pathway radical trapping.<sup>152</sup> Emerging GCH1/BH4 modulators could enhance dopamine synthesis alongside anti-oxidant effects.<sup>153</sup>

Recent (2021–2026) PD trials highlight ferrostatin-1 analogs, liproxstatin-1, and α-lipoic acid as ferroptosis inhibitors attenuating nigral lipid peroxidation in MPTP/6-OHDA models.<sup>154</sup> TFR1 antagonists curb iron import; xCT activators replenish GSH.<sup>155</sup> Combined chelation-antioxidant regimens show synergistic dopaminergic preservation.<sup>156</sup> These strategies shift PD therapy from L-DOPA palliation toward upstream homeostasis restoration, potentially halting progression in iron-vulnerable basal ganglia circuits.<sup>157,158</sup>

Medicine	Mechanism and Function
<b>Iron Chelators</b>	
Deferiprone	Iron chelator; inhibits pathological α-synuclein toxicity in sporadic PD mouse models
Desferrioxamine	Iron chelator; directly binds excess iron
<b>Nrf2 Pathway Activators</b>	

Medicine	Mechanism and Function
Alpha lipoic acid	Antioxidant/iron chelator; activates SIRT1/Nrf2 signaling to regulate iron metabolism and suppress ferroptosis
Gastrodin	Antioxidant; upregulates Nrf2, GPX4, ferroportin-1 (FPN1), and HO-1 protein expression
Hinokitiol	Antioxidant/iron chelator; activates Nrf2 cytoprotective transcription
Icariside II	Antioxidant; activates Keap1/Nrf2/GPX4 signaling pathway
Morroniside	Antioxidant; activates Nrf2/ARE signaling to protect PD dopaminergic neurons from ferroptosis
Paeoniflorin	Antioxidant; activates Akt/Nrf2/GPx4 pathway
Quercetin	Antioxidant; inhibits ferroptosis via Nrf2 activation
<b>GPX4/GSH Axis Modulators</b>	
Lapatinib	Activates GPX4/GSH/NRF2 axis; inhibits oxidative markers (iron, TfR1, PTGS2, 4-HNE); suppresses p-EGFR/c-SRC/PKCβII/PLC-γ/ACSL-4 pathway
Idebenone	Inhibits NAD(P)H dehydrogenase downregulation, reduces lipid peroxidation, increases GPX4 expression
<b>Ferritinophagy &amp; Iron Regulators</b>	
Buddlejasaponin IVb	Suppresses IRP2-mediated iron overload

Medicine	Mechanism and Function
DI-3-n-butylphthalide	Regulates ferritin expression, promotes Nrf2 nuclear translocation, inhibits NCOA4-mediated ferritinophagy
<b>Antioxidants &amp; Multi-target</b>	
Doxycycline & Demeclocycline	Prevent intracellular oxidative stress and mitochondrial membrane depolarization
Probiotic Strain L. lactis MG1363-pMG36e-GLP-1	Activates Keap1/Nrf2/GPX4 pathway; downregulates ACSL4, upregulates FSP1 to suppress ferroptosis
<b>Novel Pathway Modulators</b>	
Pazopanib	Targets HSP90/CDC37 and multiple regulated cell death (RCD) mechanisms
Rapamycin	Autophagy inducer; inhibits ferroptosis via autophagy activation
$\beta$ -hydroxybutyrate	Alleviates oxidative stress/ferroptosis via ZFP36/ACSL4 axis modulation

Table 1: Depicting the different forms of Iron chelating agents and modulators

### Iron and Calcium Interplay

Iron generates reactive oxygen species (ROS) that serve vital signaling roles in neurons, sculpting synaptic plasticity through structural and functional remodelling. ROS fine-tunes key effectors like NMDA receptors, voltage-gated  $\text{Ca}^{2+}$  channels (VGCCs),  $\text{K}^{+}$  channels, and CaMKII<sup>159</sup>—central to activity-dependent plasticity. Notably, ROS triggers redox modifications of ryanodine receptors (RyR) in hippocampal neurons, unleashing ER  $\text{Ca}^{2+}$  release that phosphorylates plasticity-linked enzymes.<sup>160</sup> Iron-driven ROS similarly activates RyR-mediated  $\text{Ca}^{2+}$  signaling, boosting ERK1/2 phosphorylation even in  $\text{Ca}^{2+}$ -free conditions, underscoring physiologic synergy.<sup>161</sup> Yet excess ROS proves neurotoxic. Heightened iron sparks RyR-dependent  $\text{Ca}^{2+}$  liberation, promoting pathological mitochondrial fission and dysfunction.<sup>162</sup> Iron-catalyzed lipid peroxidation further devastates mitochondria, elevating matrix  $\text{Ca}^{2+}$  that hyperactivates calcineurin—a  $\text{Ca}^{2+}$ -dependent phosphatase—culminating in neuronal demise.<sup>163,164</sup>

This iron-Ca<sup>2+</sup> interplay runs bidirectional. Ca<sup>2+</sup> governs numerous antioxidant defenses and ROS-generating enzymes.<sup>164</sup> Lipid peroxidation by-products like 4-hydroxynonenal (4-HNE) directly gate plasma membrane Ca<sup>2+</sup> channels, including hippocampal VGCCs.<sup>165,166</sup> Elevated 4-HNE disrupts Na<sup>+</sup>/Ca<sup>2+</sup> pumps and alters channel permeability, provoking energy failure and cell death.<sup>167,68</sup> Conversely, Ca<sup>2+</sup> dysregulation amplifies ROS cascades.<sup>169</sup> Cytosolic Ca<sup>2+</sup> overload or mitochondrial influx unleashes ROS bursts, destabilizing the labile iron pool.<sup>170</sup> Excess cytoplasmic Ca<sup>2+</sup> stimulates neuronal nitric oxide synthase (nNOS) and NAD(P)H oxidase<sup>171,172</sup>—both ROS sources and plasticity modulators—forming vicious feed-forward loops.<sup>173</sup> This intimate crosstalk renders neurons exquisitely vulnerable: iron dysregulation provokes Ca<sup>2+</sup> storms, while Ca<sup>2+</sup> mishandling exacerbates iron toxicity.<sup>173,174</sup> In Parkinson's disease, substantia nigra iron deposits ignite Fenton chemistry, generating hydroxyl radicals that sensitize RyR/IP3R channels<sup>175</sup> and overwhelm mitochondrial Ca<sup>2+</sup> uniporters (MCU).<sup>176</sup> Resultant matrix Ca<sup>2+</sup> spikes open permeability transition pores (mPTP), while cytosolic surges activate calpains and calcineurin, shredding dopaminergic circuits.<sup>177,178</sup>

Therapeutically, dual-targeting holds promise. Iron chelators like deferiprone blunt ROS ignition,<sup>179</sup> while MCU inhibitors (Ru360) or RyR stabilizers (dantrolene) curb Ca<sup>2+</sup> amplification.<sup>180,181</sup> Nrf2 agonists simultaneously bolster GSH/GPX4 defenses against peroxidation while rebalancing ion homeostasis.<sup>182,183</sup> This convergence positions the Fe<sup>2+</sup>-Ca<sup>2+</sup> axis as a high-yield therapeutic nexus for neurodegeneration, where untangling one strand inevitably stabilizes the other.<sup>184</sup>

### Conclusion:

Parkinson's disease (PD) remains a prevalent neurodegenerative disorder with elusive etiology, where metal ion dyshomeostasis—particularly calcium (Ca<sup>2+</sup>) and iron—emerges as a pivotal driver of pathogenesis. This review underscores Ca<sup>2+</sup> as an indispensable second messenger, orchestrating neuronal excitability, synaptic plasticity, and survival through plasma membrane channels (VGCCs, NMDA-R) and intracellular stores (ER IP3R/RyR, mitochondrial MCU). Disruptions in Ca<sup>2+</sup> homeostasis precipitate dopaminergic demise in the substantia nigra, yet calcium channel blockers (e.g., dihydropyridines) reveal neuroprotective potential, illuminating microscopic Ca<sup>2+</sup> signaling as a therapeutic linchpin. Concurrently, nigral iron overload—evident via MRI hypointensities—fuels ferroptosis, an iron-dependent lipid peroxidation cascade distinct from apoptosis. While the canonical GPX4/GSH axis dominates research, with Nrf2 agonists amplifying antioxidant defenses (HO-1, NQO1, GCLC), parallel pathways warrant exploration. The FSP1/CoQ10/NAD(P)H system deploys ubiquinol radical trapping at membranes, though CoQ10 biosynthesis and membrane targeting pose key challenges. Similarly, the GCH1/BH4/DHFR pathway—critical for dopamine synthesis—offers untapped anti-peroxidant potential via BH4 radical scavenging, meriting PD-specific validation.

Critically, Ca<sup>2+</sup>-iron crosstalk amplifies vulnerability: iron-catalyzed ROS sensitizes RyR channels and VGCCs, unleashing cytosolic/mitochondrial Ca<sup>2+</sup> storms, while Ca<sup>2+</sup> overload destabilizes labile iron pools via nNOS/NADPH oxidase activation. This bidirectional synergy culminates in mPTP opening, calcineurin hyperactivation, and ferroptotic collapse.

### Future prospect:

Future therapies must transcend unidimensional targeting. Multifunctional agents—combining iron chelators (deferiprone), MCU inhibitors (Ru360), Nrf2 inducers, and CoQ10/FSP1 stabilizers—hold transformative promise. Precision nanotherapeutics or bifunctional small molecules addressing the  $\text{Ca}^{2+}$ - $\text{Fe}^{2+}$  nexus could pioneer disease-modifying interventions, shifting PD management from symptomatic palliation to homeostasis restoration.

**Author Contributions:** Writing & original draft preparation diagram and figures are done by the author.

**Funding:** Self-Funding

**Institutional Ethical Committee Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Conflicts of Interest:** The authors declare no conflicts of interest with any person or organization.

#### References:

1. Jefri, M.; Bell, S.; Peng, H.; Hettige, N.; Maussion, G.; Soubannier, V.; Wu, H.; Silveira, H.; Theroux, J.F.; Moquin, L.; et al. Stimulation of L-type calcium channels increases tyrosine hydroxylase and dopamine in ventral midbrain cells induced from somatic cells. *Stem Cells Transl. Med.* **2020**, *9*, 697–712. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
2. Liss, B.; Striessnig, J. The Potential of L-Type Calcium Channels as a Drug Target for Neuroprotective Therapy in Parkinson's Disease. *Annu. Rev. Pharmacol. Toxicol.* **2019**, *59*, 263–289. [[Google Scholar](#)] [[CrossRef](#)]
3. Berger, S.M.; Bartsch, D. The role of L-type voltage-gated calcium channels Cav1.2 and Cav1.3 in normal and pathological brain function. *Cell Tissue Res.* **2014**, *357*, 463–476. [[Google Scholar](#)] [[CrossRef](#)]
4. Samak, G.; Narayanan, D.; Jaggar, J.H.; Rao, R. CaV1.3 channels and intracellular calcium mediate osmotic stress-induced N-terminal c-Jun kinase activation and disruption of tight junctions in Caco-2 CELL MONOLAYERS. *J. Biol. Chem.* **2011**, *286*, 30232–30243. [[Google Scholar](#)] [[CrossRef](#)]
5. Kang, S.; Cooper, G.; Dunne, S.F.; Luan, C.H.; Surmeier, D.J.; Silverman, R.B. Antagonism of L-type  $\text{Ca}^{2+}$  channels CaV1.3 and CaV1.2 by 1,4-dihydropyrimidines and 4H-pyrans as dihydropyridine mimics. *Bioorg Med. Chem.* **2013**, *21*, 4365–4373. [[Google Scholar](#)] [[CrossRef](#)]
6. Hasreiter, J.; Goldnagl, L.; Bohm, S.; Kubista, H. Cav1.2 and Cav1.3 L-type calcium channels operate in a similar voltage range but show different coupling to  $\text{Ca}^{2+}$ -dependent conductances in hippocampal neurons. *Am. J. Physiol. Cell Physiol.* **2014**, *306*, C1200–C1213. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)] [[Green Version](#)]
7. Jung, J.H.; Na, H.K.; Jeong, S.H.; Chung, S.J.; Yoo, H.S.; Lee, Y.H.; Baik, K.; Kim, S.J.; Sohn, Y.H.; Lee, P.H. Effects of Dihydropyridines on the Motor and Cognitive

- Outcomes of Patients with Parkinson's Disease. *Mov. Disord.* **2023**, *38*, 843–853. [[Google Scholar](#)] [[CrossRef](#)]
8. Guzman, J.N.; Ilijic, E.; Yang, B.; Sanchez-Padilla, J.; Wokosin, D.; Galtieri, D.; Kondapalli, J.; Schumacker, P.T.; Surmeier, D.J. Systemic isradipine treatment diminishes calcium-dependent mitochondrial oxidant stress. *J. Clin. Invest.* **2018**, *128*, 2266–2280. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
  9. Ilijic, E.; Guzman, J.N.; Surmeier, D.J. The L-type channel antagonist isradipine is neuroprotective in a mouse model of Parkinson's disease. *Neurobiol. Dis.* **2011**, *43*, 364–371. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
  10. Zamponi, G.W.; Striessnig, J.; Koschak, A.; Dolphin, A.C. The Physiology, Pathology, and Pharmacology of Voltage-Gated Calcium Channels and Their Future Therapeutic Potential. *Pharmacol. Rev.* **2015**, *67*, 821–870. [[Google Scholar](#)] [[CrossRef](#)]
  11. Alexander, S.P.; Peters, J.A.; Kelly, E.; Marrion, N.V.; Faccenda, E.; Harding, S.D.; Pawson, A.J.; Sharman, J.L.; Southan, C.; Davies, J.A. The concise guide to pharmacology 2017/18: Ligand-gated ion channels. *Br. J. Pharmacol.* **2017**, *174* (Suppl. S1), S130–S159. [[Google Scholar](#)] [[CrossRef](#)]
  12. Surmeier, D.J. Determinants of dopaminergic neuron loss in Parkinson's disease. *FEBS J.* **2018**, *285*, 3657–3668. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
  13. Minakaki, G.; Krainc, D.; Burbulla, L.F. The Convergence of Alpha-Synuclein, Mitochondrial, and Lysosomal Pathways in Vulnerability of Midbrain Dopaminergic Neurons in Parkinson's Disease. *Front. Cell Dev. Biol.* **2020**, *8*, 580634. [[Google Scholar](#)] [[CrossRef](#)]
  14. Erhardt, B.; Marcora, M.S.; Frenkel, L.; Boichichio, P.A.; Bodin, D.H.; Silva, B.A.; Farias, M.I.; Allo, M.A.; Hocht, C.; Ferrari, C.C.; et al. Plasma membrane calcium ATPase downregulation in dopaminergic neurons alters cellular physiology and motor behaviour in *Drosophila melanogaster*. *Eur. J. Neurosci.* **2021**, *54*, 5915–5931. [[Google Scholar](#)] [[CrossRef](#)]
  15. Collins, H.E.; Zhang, D.; Chatham, J.C. STIM and Orai Mediated Regulation of Calcium Signaling in Age-Related Diseases. *Front. Aging* **2022**, *3*, 876785. [[Google Scholar](#)] [[CrossRef](#)]
  16. Nwokonko, R.M.; Cai, X.; Loktionova, N.A.; Wang, Y.; Zhou, Y.; Gill, D.L. The STIM-Orai Pathway: Conformational Coupling Between STIM and Orai in the Activation of Store-Operated Ca<sup>2+</sup> Entry. *Adv. Exp. Med. Biol.* **2017**, *993*, 83–98. [[Google Scholar](#)] [[PubMed](#)]
  17. Prakriya, M.; Lewis, R.S. Store-Operated Calcium Channels. *Physiol. Rev.* **2015**, *95*, 1383–1436. [[Google Scholar](#)] [[CrossRef](#)]
  18. Serwach, K.; Gruszczynska-Biegala, J. Target Molecules of STIM Proteins in the Central Nervous System. *Front. Mol. Neurosci.* **2020**, *13*, 617422. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]

19. Nguyen, N.T.; Han, W.; Cao, W.M.; Wang, Y.; Wen, S.; Huang, Y.; Li, M.; Du, L.; Zhou, Y. Store-Operated Calcium Entry Mediated by ORAI and STIM. *Compr. Physiol.* **2018**, *8*, 981–1002. [[Google Scholar](#)] [[PubMed](#)]
20. Meng, F.; Fleming, B.A.; Jia, X.; Rousek, A.A.; Mulvey, M.A.; Ward, D.M. Lysosomal iron recycling in mouse macrophages is dependent upon both LcytB and Steap3 reductases. *Blood Adv.* **2022**, *6*, 1692–1707. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
21. Gruszczynska-Biegala, J.; Kuznicki, J. Native STIM2 and ORAI1 proteins form a calcium-sensitive and thapsigargin-insensitive complex in cortical neurons. *J. Neurochem.* **2013**, *126*, 727–738. [[Google Scholar](#)] [[CrossRef](#)] [[PubMed](#)]
22. Agulhon C, Petravicz J, McMullen AB, Sweger EJ, Minton SK, Taves SR, Casper KB, Fiacco TA, McCarthy KD. What is the role of astrocyte calcium in neurophysiology? *Neuron* **59**: 932–946, 2008.
23. Alzheimer C, Schwandt PC, Crill WE. Modal gating of Na<sup>+</sup> channels as a mechanism of persistent Na<sup>+</sup> current in pyramidal neurons from rat and cat sensorimotor cortex. *J Neurosci* **13**: 660–673, 1993.
24. Amzica F, Massimini M, Manfridi A. Spatial buffering during slow and paroxysmal sleep oscillations in cortical networks of glial cells in vivo. *J Neurosci* **22**: 1042–1053, 2002.
25. Amzica F, Steriade M. Neuronal and glial membrane potentials during sleep and paroxysmal oscillations in the neocortex. *J Neurosci* **20**: 6648–6665, 2000.
26. Armstrong CM. Distinguishing surface effects of calcium ion from pore-occupancy effects in Na<sup>+</sup> channels. *Proc Natl Acad Sci USA* **96**: 4158–4163, 1999.
27. Armstrong CM, Matteson DR. Two distinct populations of calcium channels in a clonal line of pituitary cells. *Science* **227**: 65–67, 1985.
28. Astori S, Wimmer RD, Prosser HM, Corti C, Corsi M, Liaudet N, Volterra A, Franken P, Adelman JP, Luthi A. The Ca(V)3.3 calcium channel is the major sleep spindle pacemaker in thalamus. *Proc Natl Acad Sci USA* **108**: 13823–13828, 2011.
29. Avoli M, D'Antuono M, Louvel J, Kohling R, Biagini G, Pumain R, D'Arcangelo G, Tancredi V. Network and pharmacological mechanisms leading to epileptiform synchronization in the limbic system in vitro. *Prog Neurobiol* **68**: 167–207, 2002.
30. Avoli M, Drapeau C, Perreault P, Louvel J, Pumain R. Epileptiform activity induced by low chloride medium in the CA1 subfield of the hippocampal slice. *J Neurophysiol* **64**: 1747–1757, 1990.
31. Bal T, McCormick DA. Mechanisms of oscillatory activity in guinea-pig nucleus reticularis thalami in vitro: a mammalian pacemaker. *J Physiol* **468**: 669–691, 1993.
32. Barres BA, Chun LL, Corey DP. Ion channels in vertebrate glia. *Ann Rev Neurosci* **13**: 441–474, 1990.
33. Bean BP. Two kinds of calcium channels in canine atrial cells. Differences in kinetics, selectivity, and pharmacology. *J Gen Physiol* **86**: 1–30, 1985.
34. Beck A, Lohr C, Nett W, Deitmer JW. Bursting activity in leech Retzius neurons induced by low external chloride. *Pflügers Arch* **442**: 263–272, 2001.
35. Benesova J, Rusnakova V, Honsa P, Pivonkova H, Dzamba D, Kubista M, Anderova M. Distinct expression/function of potassium and chloride channels contributes to the

- diverse volume regulation in cortical astrocytes of GFAP/EGFP mice. *PLoS One* **7**: e29725, 2012.
36. Bennay M, Langer J, Meier SD, Kafitz KW, Rose CR. Sodium signals in cerebellar Purkinje neurons and Bergmann glial cells evoked by glutamatergic synaptic transmission. *Glia* **56**: 1138–1149, 2008.
  37. Bikson M, Ghai RS, Baraban SC, Durand DM. Modulation of burst frequency, duration, and amplitude in the zero-Ca<sup>2+</sup> model of epileptiform activity. *J Neurophysiol* **82**: 2262–2270, 1999.
  38. Bordey A, Sontheimer H. Properties of human glial cells associated with epileptic seizure foci. *Epilepsy Res* **32**: 286–303, 1998.
  39. Bracci E, Beato M, Nistri A. Extracellular K<sup>+</sup> induces locomotor-like patterns in the rat spinal cord in vitro: comparison with NMDA or 5-HT induced activity. *J Neurophysiol* **79**: 2643–2652, 1998.
  40. Brocard F, Shevtsova NA, Bouhadfane M, Tazerart S, Heinemann U, Rybak IA, Vinay L. Activity-dependent changes in extracellular Ca<sup>2+</sup> and K<sup>+</sup> reveal pacemakers in the spinal locomotor-related network. *Neuron* **77**: 1047–1054, 2013.
  41. Brocard F, Verdier D, Arsenault I, Lund JP, Kolta A. Emergence of intrinsic bursting in trigeminal sensory neurons parallels the acquisition of mastication in weanling rats. *J Neurophysiol* **96**: 2410–2424, 2006.
  42. Brown AM, Schwindt PC, Crill WE. Different voltage dependence of transient and persistent Na<sup>+</sup> currents is compatible with modal-gating hypothesis for sodium channels. *J Neurophysiol* **71**: 2562–2565, 1994.
  43. Brown P. Abnormal oscillatory synchronisation in the motor system leads to impaired movement. *Curr Opin Neurobiol* **17**: 656–664, 2007.
  44. Brumberg JC, Nowak LG, McCormick DA. Ionic mechanisms underlying repetitive high-frequency burst firing in supragranular cortical neurons. *J Neurosci* **20**: 4829–4843, 2000.
  45. Chattopadhyay N, Evliyaoglu C, Heese O, Carroll R, Sanders J, Black P, Brown EM. Regulation of secretion of PTHrP by Ca<sup>2+</sup>-sensing receptor in human astrocytes, astrocytomas, and meningiomas. *Am J Physiol Cell Physiol* **279**: C691–C699, 2000.
  46. Christensen RK, Petersen AV, Perrier JF. How do glial cells contribute to motor control? *Curr Pharmaceutical Design* **19**: 4385–4399, 2013.
  47. Ciccarelli R, Di Iorio P, Bruno V, Battaglia G, D'Alimonte I, D'Onofrio M, Nicoletti F, Caciagli F. Activation of A(1) adenosine or mGlu3 metabotropic glutamate receptors enhances the release of nerve growth factor and S-100beta protein from cultured astrocytes. *Glia* **27**: 275–281, 1999.
  48. Cobb SR, Larkman PM, Bulters DO, Oliver L, Gill CH, Davies CH. Activation of Ih is necessary for patterning of mGluR and mAChR induced network activity in the hippocampal CA3 region. *Neuropharmacology* **44**: 293–303, 2003.
  49. Colombo E, Franceschetti S, Avanzini G, Mantegazza M. Phenytoin inhibits the persistent sodium current in neocortical neurons by modifying its inactivation properties. *PLoS One* **8**: e55329, 2013.
  50. Coombes S, Bressloff PC. **Bursting: The Genesis of Rhythm in the Nervous System**. Hackensack, NJ: World Scientific, 2005, p. 401.

51. Crepel V, Panenka W, Kelly ME, MacVicar BA. Mitogen-activated protein and tyrosine kinases in the activation of astrocyte volume-activated chloride current. *J Neurosci* **18**: 1196–1206, 1998.
52. D'Ambrosio R, Gordon DS, Winn HR. Differential role of KIR channel and Na<sup>+</sup>/K<sup>+</sup>-pump in the regulation of extracellular K<sup>+</sup> in rat hippocampus. *J Neurophysiol* **87**: 87–102, 2002.
53. Darbon P, Tschertter A, Yvon C, Streit J. Role of the electrogenic Na/K pump in disinhibition-induced bursting in cultured spinal networks. *J Neurophysiol* **90**: 3119–3129, 2003.
54. Davey GE, Murmann P, Heizmann CW. Intracellular Ca<sup>2+</sup> and Zn<sup>2+</sup> levels regulate the alternative cell density-dependent secretion of S100B in human glioblastoma cells. *J Biol Chem* **276**: 30819–30826, 2001.
55. De Pina-Benabou MH, Srinivas M, Spray DC, Scemes E. Calmodulin kinase pathway mediates the K<sup>+</sup>-induced increase in Gap junctional communication between mouse spinal cord astrocytes. *J Neurosci* **21**: 6635–6643, 2001.
56. Deitmer JW, Verkhratsky AJ, Lohr C. Calcium signalling in glial cells. *Cell Calcium* **24**: 405–416, 1998.
57. Del Negro CA, Koshiya N, Butera RJ Jr, Smith JC. Persistent sodium current, membrane properties and bursting behavior of pre-Botzinger complex inspiratory neurons in vitro. *J Neurophysiol* **88**: 2242–2250, 2002.
58. Del Negro CA, Morgado-Valle C, Hayes JA, Mackay DD, Pace RW, Crowder EA, Feldman JL. Sodium and calcium current-mediated pacemaker neurons and respiratory rhythm generation. *J Neurosci* **25**: 446–453, 2005.
59. Destexhe A, Contreras D, Steriade M. Spatiotemporal analysis of local field potentials and unit discharges in cat cerebral cortex during natural wake and sleep states. *J Neurosci* **19**: 4595–4608, 1999.
60. Dibaj P, Kaiser M, Hirrlinger J, Kirchhoff F. Neusch Kir4 C.1. channels regulate swelling of astroglial processes in experimental spinal cord edema. *J Neurochem* **103**: 2620–2628, 2007.
61. Dietzel I, Heinemann U. Dynamic variations of the brain cell microenvironment in relation to neuronal hyperactivity. *Ann NY Acad Sci* **481**: 72–86, 1986.
62. Dietzel I, Heinemann U, Hofmeier G, Lux HD. Stimulus-induced changes in extracellular Na<sup>+</sup> and Cl<sup>-</sup> concentration in relation to changes in the size of the extracellular space. *Exp Brain Res* **46**: 73–84, 1982.
63. Djamshidian A, Grassl R, Seltenhammer M, Czech T, Baumgartner C, Schmidbauer M, Ulrich W, Zimprich F. Altered expression of voltage-dependent calcium channel alpha(1) subunits in temporal lobe epilepsy with Ammon's horn sclerosis. *Neuroscience* **111**: 57–69, 2002.
64. Duffy S, MacVicar BA. In vitro ischemia promotes calcium influx and intracellular calcium release in hippocampal astrocytes. *J Neurosci* **16**: 71–81, 1996.
65. el Manira A, Tegner J, Grillner S. Calcium-dependent potassium channels play a critical role for burst termination in the locomotor network in lamprey. *J Neurophysiol* **72**: 1852–1861, 1994.
66. Enkvist MO, McCarthy KD. Astroglial gap junction communication is increased by treatment with either glutamate or high K<sup>+</sup> concentration. *J Neurochem* **62**: 489–495, 1994.

67. Formenti A, De Simoni A, Arrigoni E, Martina M. Changes in extracellular  $\text{Ca}^{2+}$  can affect the pattern of discharge in rat thalamic neurons. *J Physiol* **535**: 33–45, 2001.
68. Forrest MD. Mathematical model of bursting in dissociated purkinje neurons. *PLoS One* **8**: e68765, 2013.
69. Franceschetti S, Lavazza T, Curia G, Aracri P, Panzica F, Sancini G, Avanzini G, Magistretti J.  $\text{Na}^+$ -activated  $\text{K}^+$  current contributes to postexcitatory hyperpolarization in neocortical intrinsically bursting neurons. *J Neurophysiol* **89**: 2101–2111, 2003.
70. Gamba G. Molecular physiology and pathophysiology of electroneutral cation-chloride cotransporters. *Physiol Rev* **85**: 423–493, 2005.
71. Ghamari-Langroudi M, Bourque CW. Flufenamic acid blocks depolarizing afterpotentials and phasic firing in rat supraoptic neurones. *J Physiol* **545**: 537–542, 2002.
72. Ghazanfar AA, Chandrasekaran C, Logothetis NK. Interactions between the superior temporal sulcus and auditory cortex mediate dynamic face/voice integration in rhesus monkeys. *J Neurosci* **28**: 4457–4469, 2008.
73. Goldman WF, Yarowsky PJ, Juhaszova M, Krueger BK, Blaustein MP. Sodium/calcium exchange in rat cortical astrocytes. *J Neurosci* **14**: 5834–5843, 1994.
74. Golomb D, Yue C, Yaari Y. Contribution of persistent  $\text{Na}^+$  current and M-type  $\text{K}^+$  current to somatic bursting in CA1 pyramidal cells: combined experimental and modeling study. *J Neurophysiol* **96**: 1912–1926, 2006.
75. Guatteo E, Franceschetti S, Bacci A, Avanzini G, Wanke E. A TTX-sensitive conductance underlying burst firing in isolated pyramidal neurons from rat neocortex. *Brain Res* **741**: 1–12, 1996.
76. Halliday GM, Stevens CH. Glia: initiators and progressors of pathology in Parkinson's disease. *Movement Disorders* **26**: 6–17, 2011.
77. Hammond C, Bergman H, Brown P. Pathological synchronization in Parkinson's disease: networks, models and treatments. *Trends Neurosci* **30**: 357–364, 2007.
78. Harris-Warrick RM. General principles of rhythmogenesis in central pattern generator networks. *Prog Brain Res* **187**: 213–222, 2010.
79. Heinemann U, Albrecht D, Kohr G, Rausche G, Stabel J, Wisskirchen T. Low- $\text{Ca}^{2+}$ -induced epileptiform activity in rat hippocampal slices. *Epilepsy Res Suppl* **8**: 147–155, 1992.
80. Heinemann U, Lux HD, Gutnick MJ. Extracellular free calcium and potassium during paroxysmal activity in the cerebral cortex of the cat. *Exp Brain Res* **27**: 237–243, 1977.
81. Hertz L, Chen Y, Spatz M. Involvement of non-neuronal brain cells in AVP-mediated regulation of water space at the cellular, organ, and whole-body level. *J Neurosci Res* **62**: 480–490, 2000.
82. Hess D, El Manira A. Characterization of a high-voltage-activated IA current with a role in spike timing and locomotor pattern generation. *Proc Natl Acad Sci USA* **98**: 5276–5281, 2001.
83. Hinterkeuser S, Schroder W, Hager G, Seifert G, Blumcke I, Elger CE, Schramm J, Steinhauser C. Astrocytes in the hippocampus of patients with temporal lobe epilepsy display changes in potassium conductances. *Eur J Neurosci* **12**: 2087–2096, 2000.
84. Hochman DW, D'Ambrosio R, Janigro D, Schwartzkroin PA. Extracellular chloride and the maintenance of spontaneous epileptiform activity in rat hippocampal slices. *J Neurophysiol* **81**: 49–59, 1999.

85. Hochman DW, Schwartzkroin PA. Chloride-cotransport blockade desynchronizes neuronal discharge in the “epileptic” hippocampal slice. *J Neurophysiol* **83**: 406–417, 2000.
86. Hounsgaard J, Nicholson C. Potassium accumulation around individual purkinje cells in cerebellar slices from the guinea-pig. *J Physiol* **340**: 359–388, 1983.
87. Huguenard JR. Low-threshold calcium currents in central nervous system neurons. *Ann Rev Physiol* **58**: 329–348, 1996.
88. Huh Y, Cho J. Discrete pattern of burst stimulation in the ventrobasal thalamus for anti-nociception. *PLoS One* **8**: e67655, 2013.
89. Jasinski PE, Molkov YI, Shevtsova NA, Smith JC, Rybak IA. Sodium and calcium mechanisms of rhythmic bursting in excitatory neural networks of the pre-Botzinger complex: a computational modelling study. *Eur J Neurosci* **37**: 212–230, 2013.
90. Jayakumar AR, Norenberg MD. The Na-K-Cl Co-transporter in astrocyte swelling. *Metab Brain Dis* **25**: 31–38, 2010.
91. Jensen MS, Yaari Y. Role of intrinsic burst firing, potassium accumulation, and electrical coupling in the elevated potassium model of hippocampal epilepsy. *J Neurophysiol* **77**: 1224–1233, 1997.
92. Ji H, Shepard PD. SK  $Ca^{2+}$ -activated  $K^+$  channel ligands alter the firing pattern of dopamine-containing neurons in vivo. *Neuroscience* **140**: 623–633, 2006.
93. Jinnō S, Ishizuka S, Kosaka T. Ionic currents underlying rhythmic bursting of ventral mossy cells in the developing mouse dentate gyrus. *Eur J Neurosci* **17**: 1338–1354, 2003.
94. Johnson SM, Smith JC, Funk GD, Feldman JL. Pacemaker behavior of respiratory neurons in medullary slices from neonatal rat. *J Neurophysiol* **72**: 2598–2608, 1994.
95. Karus C, Mondragao MA, Ziemens D, Rose CR. Astrocytes restrict discharge duration and neuronal sodium loads during recurrent network activity. *Glia* **63**: 936–957, 2015.
96. Kivi A, Lehmann TN, Kovacs R, Eilers A, Jauch R, Meencke HJ, von Deimling A, Heinemann U, Gabriel S. Effects of barium on stimulus-induced rises of  $[K^+]_o$  in human epileptic non-sclerotic and sclerotic hippocampal area CA1. *Eur J Neurosci* **12**: 2039–2048, 2000.
97. Kofuji P, Newman EA. Potassium buffering in the central nervous system. *Neuroscience* **129**: 1045–1056, 2004.
98. Kroeger D, Tamburri A, Amzica F, Sik A. Activity-dependent layer-specific changes in the extracellular chloride concentration and chloride driving force in the rat hippocampus. *J Neurophysiol* **103**: 1905–1914, 2010.
99. Kuhn AA, Trottenberg T, Kivi A, Kupsch A, Schneider GH, Brown P. The relationship between local field potential and neuronal discharge in the subthalamic nucleus of patients with Parkinson's disease. *Exp Neurol* **194**: 212–220, 2005.
100. Kuhn AA, Williams D, Kupsch A, Limousin P, Hariz M, Schneider GH, Yarrow K, Brown P. Event-related beta desynchronization in human subthalamic nucleus correlates with motor performance. *Brain* **127**: 735–746, 2004.
101. Langer J, Rose CR. Synaptically induced sodium signals in hippocampal astrocytes in situ. *J Physiol* **587**: 5859–5877, 2009.
102. Lascola CD, Kraig RP. Whole-cell chloride currents in rat astrocytes accompany changes in cell morphology. *J Neurosci* **16**: 2532–2545, 1996.

103. Latour I, Hamid J, Beedle AM, Zamponi GW, Macvicar BA. Expression of voltage-gated Ca<sup>2+</sup> channel subtypes in cultured astrocytes. *Glia* **41**: 347–353, 2003.
104. Launay P, Fleig A, Perraud AL, Scharenberg AM, Penner R, Kinet JP. TRPM4 is a Ca<sup>2+</sup>-activated nonselective cation channel mediating cell membrane depolarization. *Cell* **109**: 397–407, 2002.
105. Lee D. Coherent oscillations in neuronal activity of the supplementary motor area during a visuomotor task. *J Neurosci* **23**: 6798–6809, 2003.
106. Lee DJ, Hsu MS, Seldin MM, Arellano JL, Binder DK. Decreased expression of the glial water channel aquaporin-4 in the intrahippocampal kainic acid model of epileptogenesis. *Exp Neurol* **235**: 246–255, 2012.
107. Li Z, Hatton GI. Oscillatory bursting of phasically firing rat supraoptic neurones in low-Ca<sup>2+</sup> medium: Na<sup>+</sup> influx, cytosolic Ca<sup>2+</sup> and gap junctions. *J Physiol* **496**: 379–394, 1996.
108. Lian XY, Stringer JL. Astrocytes contribute to regulation of extracellular calcium and potassium in the rat cerebral cortex during spreading depression. *Brain Res* **1012**: 177–184, 2004.
109. Liao YF, Tsai ML, Chen CC, Yen CT. Involvement of the CaV3.2 T-type calcium channel in thalamic neuron discharge patterns. *Mol Pain* **7**: 43, 2011.
110. Liu Y, Harding M, Pittman A, Dore J, Striessnig J, Rajadhyaksha A, Chen X. CaV1.2 CaV1.3 L-type calcium channels regulate dopaminergic firing activity in the mouse ventral tegmental area. *J Neurophysiol* **112**: 1119–1130, 2014.
111. Luthi A, Bal T, McCormick DA. Periodicity of thalamic spindle waves is abolished by ZD7288, a blocker of Ih. *J Neurophysiol* **79**: 3284–3289, 1998.
112. Luu P, Tucker DM, Makeig S. Frontal midline theta and the error-related negativity: neurophysiological mechanisms of action regulation. *Clin Neurophysiol* **115**: 1821–1835, 2004.
113. MacGregor DJ, Leng G. Phasic firing in vasopressin cells: understanding its functional significance through computational models. *PLoS Comp Biol* **8**: e1002740, 2012.
114. Maier JX, Neuhoff JG, Logothetis NK, Ghazanfar AA. Multisensory integration of looming signals by rhesus monkeys. *Neuron* **43**: 177–181, 2004.
115. Mani RS, Boyes BE, Kay CM. Physicochemical and optical studies on calcium- and potassium-induced conformational changes in bovine brain S-100b protein. *Biochemistry* **21**: 2607–2612, 1982.
116. Mani RS, Shelling JG, Sykes BD, Kay CM. Spectral studies on the calcium binding properties of bovine brain S-100b protein. *Biochemistry* **22**: 1734–1740, 1983.
117. Marchetti C, Beato M, Nistri A. Evidence for increased extracellular K<sup>+</sup> as an important mechanism for dorsal root induced alternating rhythmic activity in the neonatal rat spinal cord in vitro. *Neurosci Lett* **304**: 77–80, 2001.
118. Medici V, Frassoni C, Tassi L, Spreafico R, Garbelli R. Aquaporin 4 expression in control and epileptic human cerebral cortex. *Brain Res* **1367**: 330–339, 2011.
119. Mirmiran M, Corner M. Neuronal discharge patterns in the occipital cortex of developing rats during active and quiet sleep. *Brain Res* **255**: 37–48, 1982.
120. Moody WJ, Futamachi KJ, Prince DA. Extracellular potassium activity during epileptogenesis. *Exp Neurol* **42**: 248–263, 1974.

121. Moraes DJ, da Silva MP, Bonagamba LG, Mecawi AS, Zoccal DB, Antunes-Rodrigues J, Varanda WA, Machado BH. Electrophysiological properties of rostral ventrolateral medulla presympathetic neurons modulated by the respiratory network in rats. *J Neurosci* **33**: 19223–19237, 2013.
122. Morquette P, Verdier D, Kadala A, Fethiere J, Philippe AG, Robitaille R, Kolta A. An astrocyte-dependent mechanism for neuronal rhythmogenesis. *Nat Neurosci* **18**: 844–854, 2015.
123. Mrejeru A, Wei A, Ramirez JM. Calcium-activated non-selective cation currents are involved in generation of tonic and bursting activity in dopamine neurons of the substantia nigra pars compacta. *J Physiol* **589**: 2497–2514, 2011.
124. Muller M, Schlue WR. Macroscopic and single-channel chloride currents in neuropile glial cells of the leech central nervous system. *Brain Res* **781**: 307–319, 1998.
125. Muramatsu Y, Kurosaki R, Watanabe H, Michimata M, Matsubara M, Imai Y, Araki T. Expression of S-100 protein is related to neuronal damage in MPTP-treated mice. *Glia* **42**: 307–313, 2003.
126. Nagelhus EA, Mathiesen TM, Ottersen OP. Aquaporin-4 in the central nervous system: cellular and subcellular distribution and coexpression with KIR4.1. *Neuroscience* **129**: 905–913, 2004.
127. Neckelmann D, Amzica F, Steriade M. Spike-wave complexes and fast components of cortically generated seizures. III. Synchronizing mechanisms. *J Neurophysiol* **80**: 1480–1494, 1998.
128. Neprasova H, Anderova M, Petrik D, Vargova L, Kubinova S, Chvatal A, Sykova E. High extracellular K<sup>+</sup> evokes changes in voltage-dependent K<sup>+</sup> and Na<sup>+</sup> currents and volume regulation in astrocytes. *Pflügers Arch* **453**: 839–849, 2007.
129. Newman EA, Zahs KR. Modulation of neuronal activity by glial cells in the retina. *J Neurosci* **18**: 4022–4028, 1998.
130. Nicholson C, Bruggencate GT, Steinberg R, Stockle H. Calcium modulation in brain extracellular microenvironment demonstrated with ion-selective micropipette. *Proc Natl Acad Sci USA* **74**: 1287–1290, 1977.
131. Nishimura Y, Asahi M, Saitoh K, Kitagawa H, Kumazawa Y, Itoh K, Lin M, Akamine T, Shibuya H, Asahara T, Yamamoto T. Ionic mechanisms underlying burst firing of layer III sensorimotor cortical neurons of the cat: an in vitro slice study. *J Neurophysiol* **86**: 771–781, 2001.
132. Noda H, Mikami A. Discharges of neurons in the dorsal paraflocculus of monkeys during eye movements and visual stimulation. *J Neurophysiol* **56**: 1129–1146, 1986.
133. O'Connor ER, Sontheimer H, Spencer DD, de Lanerolle NC. Astrocytes from human hippocampal epileptogenic foci exhibit action potential-like responses. *Epilepsia* **39**: 347–354, 1998.
134. Oke OO, Magony A, Anver H, Ward PD, Jiruska P, Jefferys JG, Vreugdenhil M. High-frequency gamma oscillations coexist with low-frequency gamma oscillations in the rat visual cortex in vitro. *Eur J Neurosci* **31**: 1435–1445, 2010.
135. Onimaru H, Arata A, Homma I. Intrinsic burst generation of preinspiratory neurons in the medulla of brainstem-spinal cord preparations isolated from newborn rats. *Exp Brain Res* **106**: 57–68, 1995.

136. Orkand RK, Nicholls JG, Kuffler SW. Effect of nerve impulses on the membrane potential of glial cells in the central nervous system of amphibia. *J Neurophysiol* **29**: 788–806, 1966.
137. Pace RW, Mackay DD, Feldman JL, Del Negro CA. Inspiratory bursts in the preBotzinger complex depend on a calcium-activated non-specific cation current linked to glutamate receptors in neonatal mice. *J Physiol* **582**: 113–125, 2007.
138. Panaitescu B, Ruangkittisakul A, Ballanyi K. Silencing by raised extracellular  $\text{Ca}^{2+}$  of pre-Botzinger complex neurons in newborn rat brainstem slices without change of membrane potential or input resistance. *Neurosci Lett* **456**: 25–29, 2009.
139. Payne JA, Rivera C, Voipio J, Kaila K. Cation-chloride co-transporters in neuronal communication, development and trauma. *Trends Neurosci* **26**: 199–206, 2003.
140. Pena F, Parkis MA, Tryba AK, Ramirez JM. Differential contribution of pacemaker properties to the generation of respiratory rhythms during normoxia and hypoxia. *Neuron* **43**: 105–117, 2004.
141. Perez-Reyes E. Molecular physiology of low-voltage-activated t-type calcium channels. *Physiol Rev* **83**: 117–161, 2003.
142. Pinto DJ, Patrick SL, Huang WC, Connors BW. Initiation, propagation, and termination of epileptiform activity in rodent neocortex in vitro involve distinct mechanisms. *J Neurosci* **25**: 8131–8140, 2005.
143. Putzier I, Kullmann PH, Horn JP, Levitan ES.  $\text{CaV}1.3$  channel voltage dependence, not  $\text{Ca}^{2+}$  selectivity, drives pacemaker activity and amplifies bursts in nigral dopamine neurons. *J Neurosci* **29**: 15414–15419, 2009.
144. Ransdell JL, Temporal S, West NL, Leyrer ML, Schulz DJ. Characterization of inward currents and channels underlying burst activity in motoneurons of crab cardiac ganglion. *J Neurophysiol* **110**: 42–54, 2013.
145. Reyes RC, Verkhratsky A, Parpura V. Plasmalemmal  $\text{Na}^+/\text{Ca}^{2+}$  exchanger modulates  $\text{Ca}^{2+}$ -dependent exocytotic release of glutamate from rat cortical astrocytes. *ASN Neuro* **4**: 2012.
146. Richter DW, Camerer H, Sonnhof U. Changes in extracellular potassium during the spontaneous activity of medullary respiratory neurones. *Pflügers Arch* **376**: 139–149, 1978.
147. Robinson RB, Siegelbaum SA. Hyperpolarization-activated cation currents: from molecules to physiological function. *Ann Rev Physiol* **65**: 453–480, 2003.
148. Rojas H, Ramos M, Benaim G, Caputo C, DiPolo R. The activity of the  $\text{Na}^+/\text{Ca}^{2+}$  exchanger largely modulates the  $\text{Ca}^{2+}_i$  signal induced by hypo-osmotic stress in rat cerebellar astrocytes. The effect of osmolarity on exchange activity. *J Physiol Sci* **58**: 277–279, 2008.
149. Rose CR, Karus C. Two sides of the same coin: sodium homeostasis and signaling in astrocytes under physiological and pathophysiological conditions. *Glia* **61**: 1191–1205, 2013.
150. Rose CR, Ransom BR. Regulation of intracellular sodium in cultured rat hippocampal neurones. *J Physiol* **499**: 573–587, 1997.
151. Rubin JE, Hayes JA, Mendenhall JL, Del Negro CA. Calcium-activated nonspecific cation current and synaptic depression promote network-dependent burst oscillations. *Proc Natl Acad Sci USA* **106**: 2939–2944, 2009.

152. Rubino D, Robbins KA, Hatsopoulos NG. Propagating waves mediate information transfer in the motor cortex. *Nat Neurosci* **9**: 1549–1557, 2006.
153. Rybak IA, Shevtsova NA, St-John WM, Paton JF, Pierrefiche O. Endogenous rhythm generation in the pre-Botzinger complex and ionic currents: modelling and in vitro studies. *Eur J Neurosci* **18**: 239–257, 2003.
154. Sakatani S, Seto-Ohshima A, Itohara S, Hirase H. Impact of S100B on local field potential patterns in anesthetized and kainic acid-induced seizure conditions in vivo. *Eur J Neurosci* **25**: 1144–1154, 2007.
155. Sakatani S, Seto-Ohshima A, Shinohara Y, Yamamoto Y, Yamamoto H, Itohara S, Hirase H. Neural-activity-dependent release of S100B from astrocytes enhances kainate-induced gamma oscillations in vivo. *J Neurosci* **28**: 10928–10936, 2008.
156. Sathe K, Maetzler W, Lang JD, Mounsey RB, Fleckenstein C, Martin HL, Schulte C, Mustafa S, Synofzik M, Vukovic Z, Itohara S, Berg D, Teismann P. S100B is increased in Parkinson's disease and ablation protects against MPTP-induced toxicity through the RAGE and TNF-alpha pathway. *Brain* **135**: 3336–3347, 2012.
157. Scemes E, Spray DC. Extracellular K<sup>+</sup> and astrocyte signaling via connexin and pannexin channels. *Neurochem Res* **37**: 2310–2316, 2012.
158. Schnell C, Fresemann J, Hulsmann S. Determinants of functional coupling between astrocytes and respiratory neurons in the pre-Botzinger complex. *PLoS One* **6**: e26309, 2011.
159. Sederberg PB, Schulze-Bonhage A, Madsen JR, Bromfield EB, McCarthy DC, Brandt A, Tully MS, Kahana MJ. Hippocampal and neocortical gamma oscillations predict memory formation in humans. *Cereb Cortex* **17**: 1190–1196, 2007.
160. Sharott A, Doig NM, Mallet N, Magill PJ. Relationships between the firing of identified striatal interneurons and spontaneous and driven cortical activities in vivo. *J Neurosci* **32**: 13221–13236, 2012.
161. Somjen GG. Extracellular potassium in the mammalian central nervous system. *Ann Rev Physiol* **41**: 159–177, 1979.
162. Somjen GG, Muller M. Potassium-induced enhancement of persistent inward current in hippocampal neurons in isolation and in tissue slices. *Brain Res* **885**: 102–110, 2000.
163. Sontheimer H. Voltage-dependent ion channels in glial cells. *Glia* **11**: 156–172, 1994.
164. Spain WJ, Schwindt PC, Crill WE. Anomalous rectification in neurons from cat sensorimotor cortex in vitro. *J Neurophysiol* **57**: 1555–1576, 1987.
165. Steriade M, Nunez A, Amzica F. Intracellular analysis of relations between the slow (<1 Hz) neocortical oscillation and other sleep rhythms of the electroencephalogram. *J Neurosci* **13**: 3266–3283, 1993.
166. Stockle H, Ten Bruggencate G. Fluctuation of extracellular potassium and calcium in the cerebellar cortex related to climbing fiber activity. *Neuroscience* **5**: 893–901, 1980.
167. Su H, Alroy G, Kirson ED, Yaari Y. Extracellular calcium modulates persistent sodium current-dependent burst-firing in hippocampal pyramidal neurons. *J Neurosci* **21**: 4173–4182, 2001.
168. Subbarao KV, Stolzenburg JU, Hertz L. Pharmacological characteristics of potassium-induced, glycogenolysis in astrocytes. *Neurosci Lett* **196**: 45–48, 1995.

169. Tas PW, Massa PT, Kress HG, Koschel K. Characterization of an  $\text{Na}^+/\text{K}^+/\text{Cl}^-$  co-transport in primary cultures of rat astrocytes. *Biochim Biophys Acta* **903**: 411–416, 1987.
170. Tazerart S, Vinay L, Brocard F. The persistent sodium current generates pacemaker activities in the central pattern generator for locomotion and regulates the locomotor rhythm. *J Neurosci* **28**: 8577–8589, 2008.
171. Thimm J, Mechler A, Lin H, Rhee S, Lal R. Calcium-dependent open/closed conformations and interfacial energy maps of reconstituted hemichannels. *J Biol Chem* **280**: 10646–10654, 2005.
172. Trevelyan AJ, Sussillo D, Watson BO, Yuste R. Modular propagation of epileptiform activity: evidence for an inhibitory veto in neocortex. *J Neurosci* **26**: 12447–12455, 2006.
173. Tsuruyama K, Hsiao CF, Chandler SH. Participation of a persistent sodium current and calcium-activated nonspecific cationic current to burst generation in trigeminal principal sensory neurons. *J Neurophysiol* **110**: 1903–1914, 2013.
174. Uhlhaas PJ, Singer W. Neural synchrony in brain disorders: relevance for cognitive dysfunctions and pathophysiology. *Neuron* **52**: 155–168, 2006.
175. Ullrich ND, Voets T, Prenen J, Vennekens R, Talavera K, Droogmans G, Nilius B. Comparison of functional properties of the  $\text{Ca}^{2+}$ -activated cation channels TRPM4 and TRPM5 from mice. *Cell Calcium* **37**: 267–278, 2005.
176. Van Eldik LJ, Zimmer DB. Secretion of S-100 from rat C6 glioma cells. *Brain Res* **436**: 367–370, 1987.
177. Vega-Saenz de Miera EC, Rudy B, Sugimori M, Llinas R. Molecular characterization of the sodium channel subunits expressed in mammalian cerebellar Purkinje cells. *Proc Natl Acad Sci USA* **94**: 7059–7064, 1997.
178. Verkhratsky A, Steinhauser C. Ion channels in glial cells. *Brain Res Brain Res Rev* **32**: 380–412, 2000.
179. Wallen P, Grafe P, Grillner S. Phasic variations of extracellular potassium during fictive swimming in the lamprey spinal cord in vitro. *Acta Physiol Scand* **120**: 457–463, 1984.
180. Wang XJ. Neurophysiological and computational principles of cortical rhythms in cognition. *Physiol Rev* **90**: 1195–1268, 2010.
181. Westenbroek RE, Bausch SB, Lin RC, Franck JE, Noebels JL, Catterall WA. Upregulation of L-type  $\text{Ca}^{2+}$  channels in reactive astrocytes after brain injury, hypomyelination, and ischemia. *J Neurosci* **18**: 2321–2334, 1998.
182. Xu L, Zeng LH, Wong M. Impaired astrocytic gap junction coupling and potassium buffering in a mouse model of tuberous sclerosis complex. *Neurobiol Disease* **34**: 291–299, 2009.
183. Xu W, Lipscombe D. Neuronal  $\text{Ca}(\text{V})1.3\alpha(1)$  L-type channels activate at relatively hyperpolarized membrane potentials and are incompletely inhibited by dihydropyridines. *J Neurosci* **21**: 5944–5951, 2001.
184. Zanotti S, Charles A. Extracellular calcium sensing by glial cells: low extracellular calcium induces intracellular calcium release and intercellular signaling. *J Neurochem* **69**: 594–602, 1997.