

Relation of magnesium with insulin resistance and inflammatory markers in subjects with known Coronary artery disease

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ABSTRACT

Background: Magnesium plays an important role in regulating insulin action and secretion. Dietary magnesium deficiency and hypomagnesemia have been associated with inflammation. There is paucity of data regarding this in Indian subjects. Hence, we studied the relation of serum and dietary magnesium with insulin resistance (IR) and inflammatory markers. **Methods & Results:** We studied the relation of serum high sensitive C-Reactive Protein (hsCRP), interleukin-6 (IL-6) and tumor Necrosis Factor (TNF- α) with cardiovascular risk factors in three hundred coronary artery disease patients. Nutrition assessment was done at the time of recruitment. Patients were evaluated for anthropometry, cardiovascular risk factors, and blood samples were collected. Patients were divided into three groups according to serum magnesium levels; ≤ 1.6 (Group 1), $>1.6-2.6$ (Group 2) and >2.6 mg/dl (Group 3) and into two groups according to recommended dietary allowance (RDA) for magnesium (≤ 350 mg/day and >350 mg/day). More than half of patients (58.6%) were having hypomagnesemia and 62% were consuming magnesium below RDA. Insulin and IR was significantly higher and insulin sensitivity was significantly lower in group 1 compared to group 2 and 3. Insulin and IR was correlated negatively; and insulin sensitivity was correlated positively with serum magnesium. Similarly inflammatory markers were significantly higher in group 1 when compared with group 2 and 3 and they were correlated negatively with serum magnesium. Dietary magnesium was positively correlated with serum magnesium. Insulin and IR were significantly higher and insulin sensitivity was significantly lower in subjects with low RDA for magnesium. Insulin and IR were correlated negatively and sensitivity was correlated positively with dietary magnesium. Inflammatory markers; IL-6, TNF- α , and hs CRP were significantly high in subjects with low RDA for magnesium and were correlated negatively with dietary magnesium. **Conclusion:** Hypomagnesemia and low dietary magnesium is associated with IR and inflammatory markers in patients with coronary artery disease.

Keywords: Inflammatory markers; Hypomagnesemia; C-reactive protein; Interleukin-6; Tumor necrosis factor- α ; Coronary artery disease

INTRODUCTION

Magnesium is the second most abundant intracellular cation and is an essential element that has numerous biological func-

tions. Magnesium is plentiful in green leafy, cereal, grain, nuts and legumes, however, refining or processing of food may deplete magnesium content by nearly 85%¹. About 30% of ingested magnesium is absorbed by distal small intestine. Magnesium acts as a cofactor in many of enzymatic reactions in the human body. Magnesium is essential for the synthesis of nucleic acids and proteins, for intermediary metabolism particularly related to the neuromuscular and cardiovascular systems. Over 300 enzymes are dependent on magnesium. Magnesium influences the enzymatic activity by binding to legends such as ATP in ATP-requiring enzymes, enolase, pyruvate kinase, and pyrophosphatase, leading to conformational change during

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the catalytic process, eg. Na⁺,K⁺-ATPase, or aggregation of multi-enzyme complexes eg. Aldehyde dehydrogenase². Hence, magnesium plays an important role in regulating insulin action, insulin-mediated glucose uptake, and insulin secretion. Magnesium depletion may result in a defective tyrosine kinase activity at the insulin receptor level causing impairment in insulin action, and aggravation of insulin resistance³. This becomes more relevant in Indians as there is high prevalence of magnesium deficiency⁴; and Indians have higher prevalence of insulin resistance and type-2 diabetes mellitus (T2DM)⁵. Dietary magnesium deficiency and hypomagnesemia have also been associated with inflammatory response leading to endothelial dysfunction and insulin resistance⁶⁻⁷. High levels of inflammatory markers have been reported in Indian adolescents⁸.

There are a few studies which evaluated the relation of dietary and serum magnesium with cardiovascular risk factors in Indian patients⁸⁻⁹, however, no study has evaluated the relation of dietary and serum magnesium with insulin resistance and inflammatory markers in Indian subjects. The study among post-menopausal women from United States which also included Asians, found no association of magnesium intake with inflammatory markers; though it was inversely related among whites and blacks¹⁰.

In view of this, we have measured markers of insulin resistance and inflammatory markers in a angiographically proven coronary artery disease. We hypothesized that serum and dietary magnesium will be inversely related with inflammatory markers and insulin resistance.

Methods

This was an extension of study conducted previously; where we have studied association of magnesium and traditional cardiovascular risk factors⁹. Three hundred patients with known coronary disease above the age of 25 years were included in this study. Patients, who were admitted in cardiology department for evaluation of chest pain and found angiography positive, were selected in the study consecutively. Exclusion criteria were presence of chronic kidney disease, hepatic dysfunction, known endocrinal or rheumatologic diseases, chronic infections or alcoholism. All cases were interviewed using a questionnaire, which included data on smoking, physical activity. Height, weight, waist, hip circumference were measured. Body mass index (BMI) and waist hip ratio (WHR) was calculated.

Nutrition assessment was done once at the time of recruitment; based on previous two days 24 hour dietary recall. Mean of recall of two days was taken. Diet was assessed using a computer based comprehensive diet assessment known as Diet soft software, version: 1.1.7 [developed by Invincible IDeAS (www.

invincibleideas.com) based on book Nutritive value of Indian Foods by C. Gopalan, B.V.Rama Sastri and S.C.Balasubramanian, National Institute of Nutrition, Indian council of Medical Research, Hyderabad, India]¹¹. Questionnaires and diet assessments were administered via interview by trained staff. Portion size estimation was undertaken using volume measures, circular measures, numbers and linear measures. To help participants estimate portion sizes, interviewers provided commonly used serving plates, bowls, utensils, cups and spoons. If measurements could not be given we recorded in three sizes: small, medium or large and any unusual intake was noted on the recall. For each nutrient, we used a standardized unit of measurement and reported values per 100 grams of edible portion of food product. There was no significant change in the dietary pattern of these participants and no change in lifestyle also.

Fasting blood samples were collected after 14 hour fasting. Magnesium was measured by Xylidyl blue method, which has no interference due to calcium. Patients were divided into three groups according to serum magnesium levels; group-1: ≤ 1.6 mg/dl (n=176, 58.6%), group 2: $> 1.6-2.6$ mg/dl (n=102, 34%) and group 3: > 2.6 mg/dl (n=26, 8.6%) and into two groups according to Recommended Dietary Allowance of magnesium¹¹; group-1 with dietary intake of magnesium ≤ 350 mg/day (n = 186, 62%) and group 2 (n = 114, 38%) with dietary intake of magnesium levels > 350 mg/day. Tumor necrosis factor-alpha (TNF- α), interleukin-6 (IL-6) and highly sensitive C-reactive protein (hsCRP), were measured with kits manufactured by Gen-probe Diaclone, France and Biocheck, CA, USA, respectively. Insulin was done by Microparticle enzyme immunoassay (MEIA). Insulin resistance and sensitivity was calculated by using homeostatic model analysis (HOMA) model [HOMA-IR=fasting insulin (μ IU/ml)*fasting glucose (mmol/l)/22.5 and quantitative insulin sensitivity check index (QUICKI) [QUICKI=1/(log(fasting insulin μ U/mL)+log(fasting glucose mg/dL))] respectively. Intra assay and inter assay precision was $<5\%$ and $<10\%$ respectively for above parameters. The study was approved by Institutional ethics committee of Deenanath Mangeshkar Hospital. Informed consent was obtained from all subjects.

Statistical method

Statistical analysis was carried out using SPSS Version 20 (SPSS Inc. Chicago, USA) and EPI INFO 3.5.3 (CDC, Atlanta, GA, USA). Data were presented as mean \pm SD (95% confidence interval) unless specified. All non-parametric data involved were analyzed by chi-square test. All parametric data were analyzed by student's t-test. Pearson correlation was used to evaluate the correlation between dependent variables like serum and dietary magnesium and independent variables like cardiovascular risk factors. A p value of < 0.05 was considered statistically significant.

Results

Three hundred patients with known cardiovascular disease (M:216; F:84, age:25–92) were studied. A comparison of cardiovascular risk factors according to serum magnesium levels are given in table 1. Mean age of patients was 60.92 ± 12.48

years. There was no age difference between males and females (M: 60.95 ± 12.34 ; F: 61.03 ± 12.92 ; $p=0.10$). (Table 1) In present study more than half of patients (58.6%) were having low magnesium level and 62% were taking dietary magnesium below recommended dietary allowance (350 mg/day).

Table 1 Basic characteristics

Parameters	Male (N=216)	Female (N=84)	P value
Age (years)	60.95 ± 12.34 (62.00)	61.03 ± 12.92 (62.00)	0.102
Smoking n(%)	82 (38.0%)	29 (34.5%)	0.673
BMI (kg/m ²)	27.65 ± 3.69 (27.04)	28.52 ± 4.07 (27.94)	0.076
WHR	0.92 ± 0.05 (0.93)	0.91 ± 0.06 (0.93)	0.118
Less physical activity n(%)	87 (40.3%)	29 (34.5%)	0.431
Insulin (mU/L)	49.35 ± 41.58 (36.40)	52.27 ± 48.21 (42.40)	0.601
HOMA-IR	16.61 ± 17.16 (12.15)	22.28 ± 28.80 (12.01)	0.502
QUICKI	0.28 ± 0.04 (0.27)	0.27 ± 0.05 (0.21) ^a	0.464
hsCRP (mg/L)	11.87 ± 10.02 (11.78)	11.18 ± 8.72 (12.30)	0.578
IL-6 (pg/ml)	62.36 ± 73.85 (25.10)	70.15 ± 78.92 (25.00)	0.421
TNF- α (pg/ml)	23.82 ± 41.87 (9.75)	29.01 ± 38.15 (11.45)	0.324

Median is given in parentheses

BMI-Body mass index; WHR-Waist hip ratio; HOMA-IR: homeostatic model analysis – insulin resistance; QUICKI: quantitative insulin check index; hsCRP: highly sensitive C-reactive protein; IL-6: Interleukin-6; TNF- α : Tumor necrosis factor- α

Serum Magnesium

Insulin and insulin resistance were significantly higher and insulin sensitivity was significantly lower in group 1 when compared with group 2 and group 3. There was no significant difference between group-2 and group-3. (Table 2) Insulin and insulin resistance was correlated negatively; and insulin sensitivity was correlated positively with serum magnesium. (Table 4 & Figure 1). This significant association was maintained even after adjustment with age, sex, waist hip ratio and BMI. (Table 5)

Similarly IL-6, TNF- α and hsCRP was significantly higher in group1 when compared with group 2 and group 3. There was no significant difference between group-2 and group-3. Inflammatory markers were correlated negatively with serum magnesium (Table 4, Figure 1) This significant association was maintained even after adjustment with age, sex, waist hip ratio and BMI (Table 5).

Dietary Magnesium

Dietary magnesium was positively correlated with serum magnesium and this was maintained even after adjustment with age and sex in multiple regression analysis. Insulin and insulin resistance was significantly higher and insulin sensitivity was significantly lower in group-1 when compared with group-2 (Table 3). Insulin and insulin resistance were correlated negatively and sensitivity was correlated positively with dietary magnesium. (Figure 1) This significance remained for insulin resistance and sensitivity but not for insulin levels (Table 4). This significant association was maintained even after adjustment with age, sex, waist hip ratio and BMI (Table 5). Inflammatory markers; IL-6, TNF- α and hsCRP were significantly higher in group 1 compared to group 2 and were correlated negatively with dietary magnesium. Inflammatory markers were negatively correlated with dietary magnesium levels; which were maintained even after adjustment with age, sex, BMI and waist hip ratio in multiple regression analysis (Table 4–5 & Figure 1).

Table 2 Insulin resistance and inflammatory markers according to serum magnesium levels

Parameters	Group 1 (≤ 1.6 mg/dl) N=176	Group 2 (> 1.6 – 2.6 mg/dl) N=102	Group 3 (> 2.6 mg/dl) N=22	p-value#
Insulin (mU/L)	61.43 \pm 46.20 (54.55–68.31)	36.27 \pm 34.70 (29.94–43.61)	22.25 \pm 22.70 (12.15–32.36)	<0.0001
P-value		<0.0001*	<0.0001†, 0.136ffi	
HOMA-IR	24.60 \pm 24.70 (20.92–28.28)	9.61 \pm 9.14 (7.82–11.41)	6.80 \pm 7.58 (3.44–10.16)	<0.0001
P-value		<0.0001*	<0.0001†, 0.546ffi	
QUICKI	0.26 \pm 0.02 (0.25–0.26)	0.30 \pm 0.04 (0.29–0.31)	0.31 \pm 0.04 (0.27–0.28)	<0.0001
P-value		<0.0001*	<0.0001†, 0.144ffi	
IL-6(pg/ml)	98.00 \pm 77.18 (86.52–109.48)	18.62 \pm 41.20 (10.53–26.72)	9.81 \pm 7.99 (6.27–13.36)	<0.0001
P-value		<0.0001*	<0.0001†, 0.558ffi	
TNF- α (pg/ml)	33.84 \pm 49.98 (26.40–41.27)	13.57 \pm 16.95 (10.24–16.90)	11.10 \pm 9.35 (6.96–15.25)	<0.0001
P-value		<0.0001*	<0.012†, 0.792ffi	
hsCRP (mg/L)	17.11 \pm 8.34 (15.87–18.35)	3.94 \pm 5.21 (2.92–4.97)	4.06 \pm 4.66 (1.99–6.13)	<0.0001
P-value		<0.0001*	<0.0001†, 0.945ffi	

All values are given as mean \pm SD (95%CI)

*P value between Group-1 and Group-2, † P value between Group-1 and Group-3, ffi P value between Group-2 and Group-3, #P value among all groups

HOMA: homeostatic model analysis – insulin resistance; QUICKI: quantitative insulin check index; IL-6: Interleukin-6; TNF- α : Tumor necrosis factor- α ; hsCRP: highly sensitive C-reactive protein

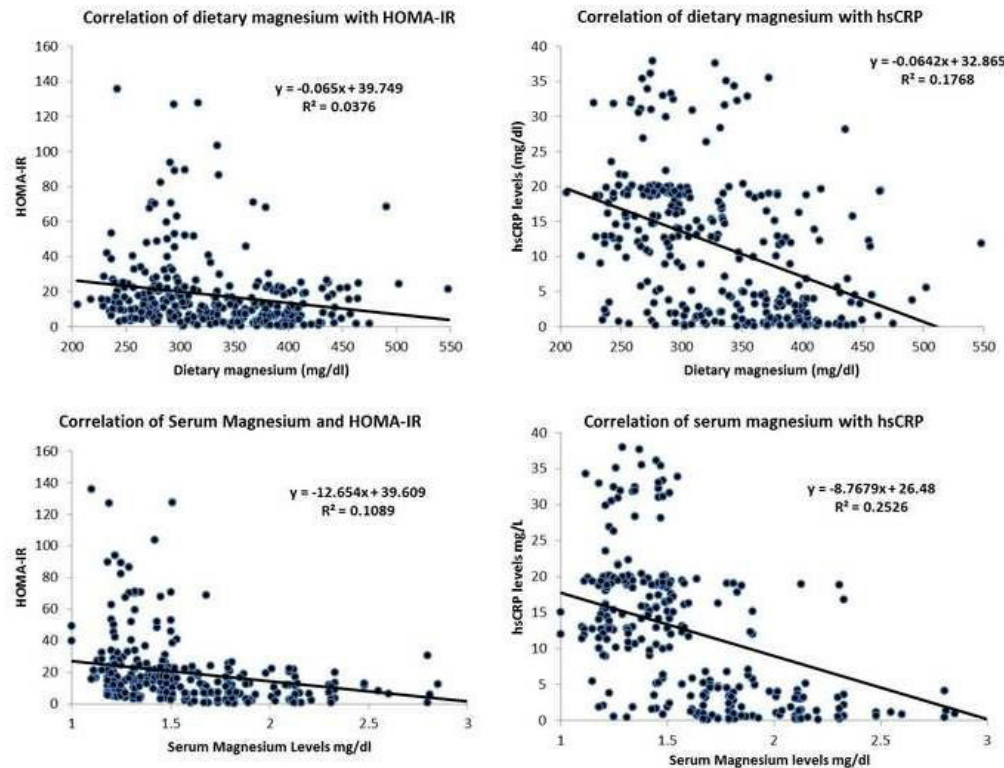


Figure 1: Relation of serum and dietary magnesium with insulin resistance (HOMA-IR) and inflammatory marker (hsCRP)

Table 3 Insulin resistance and inflammatory markers according to dietary magnesium levels

Parameters	Dietary magnesium intake		P Value
	≤ 350 mg/day N=186	>350 mg/day N=114	
Insulin (mU/L)	54.16 ± 42.30 (48.04–60.29)	43.65 ± 44.69 (35.37–51.95)	0.0420
HOMA-IR	22.07 ± 24.24 (18.56–25.58)	11.88 ± 12.66 (9.53–14.23)	<0.0001
QUICKI	0.27 ± 0.03 (0.26–0.27)	0.29 ± 0.04 (0.28–0.30)	<0.0001
IL-6 (pg/ml)	84.38 ± 78.82 (72.97–95.78)	32.19 ± 55.76 (21.84–42.53)	<0.0001
TNF-α (pg/ml)	32.12 ± 49.38 (24.98–39.27)	14.11 ± 14.93 (11.34–16.88)	<0.0001
hsCRP (mg/L)	14.90 ± 9.41 (13.59–16.32)	6.33 ± 7.45 (4.94–7.71)	<0.0001

All values are given as mean ± SD (95%CI)

HOMA: homeostatic model analysis – insulin resistance; QUICKI: quantitative insulin check index; IL-6: Interleukin-6; TNF-α: Tumor necrosis factor-α; hsCRP: highly sensitive C-reactive protein

Table 4 Correlation of serum and dietary magnesium with insulin resistance and inflammatory markers

Parameters	Serum magnesium		Dietary magnesium	
	r value	P-Value	r value	P-Value
Insulin	-0.301	<0.0001	-0.086	0.138
HOMA IR	-0.323	<0.0001	-0.180	0.002
QUICKI	0.416	<0.0001	0.252	<0.0001
IL-6	-0.420	<0.0001	-0.285	<0.0001
TNF-α	-0.212	<0.0001	-0.212	<0.0001
hsCRP	-0.495	<0.0001	-0.404	<0.0001

HOMA: homeostatic model analysis – insulin resistance; QUICKI: quantitative insulin check index; IL-6: Interleukin-6; TNF-α: Tumor necrosis factor-α; hsCRP: highly sensitive C-reactive protein

Table 5 Multiple regression analysis of Serum and dietary magnesium with insulin resistance and inflammatory factors after adjusting with age, sex, BMI and WHR

Parameters	Serum magnesium		Dietary magnesium	
	Beta coefficient	P-Value	Beta coefficient	P-Value
Insulin	-0.004	<0.0001	-0.125	0.1559
HOMA-IR	-0.008	<0.0001	-0.567	0.0017
QUICKI	5.545	<0.0001	393.9	<0.0001
IL-6	-0.003	<0.0001	-0.250	<0.0001
TNF-α	-0.003	0.0001	-0.365	0.0001
HsCRP	-0.029	<0.0001	-2.783	<0.0001

HOMA: homeostatic model analysis – insulin resistance; QUICKI: quantitative insulin check index; IL-6: Interleukin-6; TNF-α: Tumor necrosis factor-α; hsCRP: highly sensitive C-reactive protein

Discussion

Coronary artery disease is well-known for its association with various metabolic abnormalities such as hypertension, dyslipidemia, and a reduction in glucose tolerance, which are associated with underlying insulin resistance and inflammatory markers¹². Exact mechanism of underlying insulin resistance and chronic low grade inflammation is unknown. This has been explained by genetic and epigenetic mechanisms¹³. Genetic changes have long gestational period before the biological effect takes place. With recent epidemic of non-communicable disease, changes in environmental factors are more likely the cause. Environmental factors include environmental toxins¹⁴ and life style factors¹⁵. Among life style factors, physical activity is related to insulin resistance and inflammatory markers¹⁶. Another aspect of life style is nutritional factors, which has been changed considerably with change in economic progress and modernization¹⁷. Several studies have evaluated relation of nutritional factors with insulin resistance and inflammation^{18–20}.

Magnesium, being the second abundant intracellular cation and its involvement in more than 300 enzymatic reactions, has been studied for its relation with insulin resistance and inflammation¹. In the present study, insulin resistance, and insulin levels were higher and insulin sensitivity was lower in subject with hypomagnesaemia as compared to subjects with normal serum magnesium levels. Similar observation has been made by others, where serum magnesium was inversely related to HOMA IR³, and insulin levels².

Serum magnesium levels were positively correlated with insulin sensitivity (QUICKI) and negatively with insulin levels and insulin resistance (HOMA-IR), which persisted after adjustment for age, sex, BMI and WHR in this study. Studies in American adults³ and obese non diabetic children² have also reported positive correlation with quantitative insulin sensitivity check index with serum magnesium. Serum magnesium deficiency in this study may be secondary to decreased dietary magnesium intake as serum magnesium was positively related to dietary magnesium intake⁹. Studies among women with polycystic ovarian syndrome did not show any correlation between serum magnesium and insulin resistance^{21–22}. This may be due to hormonal alteration in women with polycystic ovarian syndrome. Sometimes, serum magnesium may not accurately reflect intracellular magnesium levels, which may be low, even when serum levels are within the normal range^{3,15}.

In present study, patients with low dietary magnesium intake had significantly higher insulin levels and insulin resistance (HOMA-IR) and lower insulin sensitivity (QUICKI). Univariate analysis confirmed this correlation, but in multiple regression analysis, correlation of dietary magnesium was significant for

insulin resistance but not for insulin after adjustment for age, sex, BMI, and smoking. Most of epidemiologic studies have shown an inverse association between magnesium intake and fasting insulin concentration^{23–24}.

In healthy postmenopausal women study red blood cells magnesium was directly correlated with insulin and HOMA-IR²⁵. Magnesium deficit may independently lead to a higher risk for insulin resistance and cardiovascular disease. Insulin resistance is implicated in progression of type-2 diabetes mellitus along with decreased insulin secretion. In follow up studies magnesium intake was inversely associated with incidence of diabetes in young American adults³, Chinese²⁶, and Australian adults²⁷. A large prospective cohort study of older Iowa women showed strong inverse associations with incidence of diabetes²⁸. A Meta-analysis of prospective cohort studies also demonstrated similar results²⁵. Contrary to this, in a few studies no association was detected between dietary magnesium intake and the risk for incident type 2 diabetes^{29–30}.

This can possibly be explained by the other dietary components such as fibres in food that are high in magnesium³¹. Magnesium supplementation in subjects with insulin resistance and type 2 diabetes mellitus resulted in improvement of insulin sensitivity^{32–33} and beta-cell response to glucose³². These findings suggest that increased consumption of magnesium-rich foods such as whole grains, beans, nuts, and green leafy vegetables may reduce the risk of type 2 diabetes mellitus.

Intracellular magnesium plays a key role in regulating insulin action, insulin-mediated-glucose uptake and vascular tone³⁴. Reduced intracellular magnesium concentrations result in a defective tyrosine-kinase activity, post-receptor impairment in insulin action, and worsening of insulin resistance in diabetic patients¹. It was observed that autophosphorylation of the beta-subunit of the insulin receptor was significantly reduced by 50% in magnesium-deficient rats and the tyrosine kinase activity of insulin receptors by hypomagnesaemia³⁵.

Clinical and animal studies have described influence of magnesium on inflammation^{36–37}. In present study, hypomagnesaemia was associated with increased levels of inflammatory markers suggesting associated low grade inflammation. Other studies have also reported strong correlation of low serum magnesium with low grade systemic inflammation demonstrated by elevated CRP^{38–39} and TNF- α concentrations^{40–41}. In children elevated hsCRP levels were independently associated with hypomagnesaemia⁴². In Indian children and adolescents, high levels of inflammatory markers have been reported,¹² which can partially be explained by low magnesium levels.

Serum magnesium levels are reflection of dietary intake⁹. Several studies have assessed relation of dietary magnesium with inflammatory markers³⁹⁻⁴⁴. In this study, higher inflammatory markers were observed among patients with magnesium intake lower than RDA. Inflammatory markers were negatively correlated with dietary magnesium intake. One study reported lower CRP concentration (12%), in the subjects in the highest quintiles when compared with lowest quintiles³⁹. In our study, this difference was more marked with hs CRP levels, 42.8% lower in patients that high magnesium intake compared to lower intake. Similar observation was also reported by other studies among different population: Italian population⁴⁴, Nurses' Health Study⁴⁵, and postmenopausal women¹³. However, magnesium supplementation did not significantly attenuate inflammatory markers compared to placebo in intervention studies^{31,46}. This further suggests probable role of other dietary constituents associated with magnesium intake significantly modulate observed relation between magnesium and inflammation⁴⁷. In National Health and Nutrition Examination Survey (NHANES) survey, individual with intake of magnesium below the RDA were 1.48–1.75 times more likely to have elevated CRP³⁹.

Dietary magnesium deficiency increases proinflammatory neuropeptides such as substance P which acts through neurokinin-1 receptors of inflammatory and endothelial cells, and may stimulate release of inflammatory mediators⁴⁸. Magnesium acts as a natural calcium antagonist, the molecular basis for the inflammatory response is probably the result of a modulation of the intracellular calcium concentration. Potential mechanisms includes the priming of phagocytic cells, the opening of calcium channels, activation of N-methyl-D-aspartate (NMDA) receptors, the activation of nuclear factor-kappa B (NFkB)^{37,38}.

Limitation of this study is that intracellular magnesium, a more sensitive indicator of magnesium balance, was not measured. Magnesium is primarily an intracellular cation; roughly 1% of whole-body magnesium is found extracellular space, and the free intracellular fraction is the portion regulating enzyme pathways⁴⁹. However, serum magnesium exhibits a good correlation with intracellular free magnesium measured by nuclear magnetic resonance spectroscopy⁵⁰.

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Conflict of Interest: None of the authors have any conflicts of interests

Conclusion

Lower serum and dietary magnesium is associated with insulin resistance and inflammatory markers in Indian patients with coronary artery disease. Higher insulin resistance has been observed in Indian children⁵¹. Increased dietary intake since childhood may positively modify insulin resistance and chronic low grade inflammation, and may decrease the impact of non-communicable diseases associated with these pathogenetic mechanisms.

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