

**ORIGINAL RESEARCH****Heteroresistance to rifampicin & isoniazid in clinical samples of patients with presumptive drug-resistant tuberculosis****Dr. Mohammad Asim Khan<sup>1</sup>, Dr. Karuna Meravi<sup>2</sup>**<sup>1</sup>Assistant Professor, Department of Community Medicine, Mahatma Gandhi Medical College (MGUMST), Jaipur, Rajasthan, India.<sup>2</sup>Assistant Professor, Department of Obstetrics and Gynaecology, Sunderlal Patwa Government Medical College, Mandsaur, M.P., India.**Corresponding Author: Dr. Mohammad Asim Khan,**

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Received: 12<sup>th</sup> June, 2024Accepted: 18<sup>th</sup> July, 2024**Abstract:****Background**

Tuberculosis (TB) remains a significant global health challenge, particularly with the rise of drug-resistant strains. Heteroresistance, where subpopulations of bacteria show varying levels of resistance within the same patient, complicates diagnosis and treatment. This study aims to assess the prevalence of heteroresistance to rifampicin and isoniazid in clinical samples from patients with presumptive drug-resistant tuberculosis.

**Materials and Methods**

A cross-sectional study was conducted from January to June 2024 at a tertiary care hospital. Clinical samples, including sputum and bronchial lavage, were collected from 200 patients suspected of having drug-resistant TB. Samples were processed using the GeneXpert MTB/RIF assay and Löwenstein-Jensen culture for drug susceptibility testing. Isolates were tested for heteroresistance to rifampicin and isoniazid using the BACTEC MGIT 960 system and molecular techniques like whole-genome sequencing.

**Results**

Out of 200 samples, 150 (75%) were confirmed to have *Mycobacterium tuberculosis* complex. Among these, 60 samples (40%) exhibited heteroresistance to rifampicin, while 45 samples (30%) showed heteroresistance to isoniazid. Dual heteroresistance to both drugs was observed in 20 samples (13.3%). Whole-genome sequencing revealed mutations in the *rpoB* and *katG* genes associated with resistance. The presence of heteroresistant populations was associated with prior TB treatment history and poor treatment outcomes.

**Conclusion**

The study highlights a substantial prevalence of heteroresistance to rifampicin and isoniazid in patients with presumptive drug-resistant TB. This underscores the need for advanced diagnostic techniques to detect heteroresistant strains and guide effective treatment regimens. Tailored therapeutic strategies are crucial to managing and controlling the spread of drug-resistant TB.

## Keywords

Tuberculosis, heteroresistance, rifampicin, isoniazid, drug-resistant TB, GeneXpert, BACTEC MGIT 960, whole-genome sequencing.

## Introduction

Tuberculosis (TB) is a major global health issue, causing approximately 1.5 million deaths annually and ranking as the leading cause of death from a single infectious agent (1). The emergence and spread of drug-resistant TB, particularly multidrug-resistant TB (MDR-TB), pose significant challenges to TB control efforts worldwide (2). MDR-TB is characterized by resistance to at least rifampicin and isoniazid, the two most potent first-line anti-TB drugs (3). Heteroresistance, defined as the coexistence of susceptible and resistant bacterial populations within the same patient, complicates diagnosis and treatment of TB (4).

Heteroresistance may arise due to inadequate drug exposure, host factors, or genetic mutations, and it can lead to treatment failure and further amplification of resistance (5, 6). Accurate detection of heteroresistant strains is crucial for appropriate treatment, as standard diagnostic methods may not identify low-frequency resistant subpopulations (7). Advanced molecular techniques such as whole-genome sequencing and the BACTEC MGIT 960 system provide improved sensitivity and specificity in detecting heteroresistance (8, 9).

In recent years, studies have reported varying prevalence rates of heteroresistance to rifampicin and isoniazid, suggesting geographic and population-specific differences (10, 11). Understanding the prevalence and genetic basis of heteroresistance in different settings is essential for developing effective diagnostic and therapeutic strategies. This study aims to investigate the prevalence and molecular characteristics of heteroresistance to rifampicin and isoniazid in clinical samples from patients with presumptive drug-resistant TB in a tertiary care hospital.

## Materials and Methods

A total of 200 patients suspected of having drug-resistant tuberculosis were enrolled in the study. Patients were selected based on clinical suspicion of drug resistance, including history of prior TB treatment, failure of initial therapy, and contact with known MDR-TB cases. Inclusion criteria also involved patients with positive acid-fast bacilli (AFB) sputum smear microscopy. Patients with non-tuberculous mycobacterial infections or incomplete clinical data were excluded from the study.

## Sample Collection and Processing

Clinical samples, including sputum and bronchial lavage, were collected from each participant. Sputum samples were decontaminated using the N-acetyl-L-cysteine-sodium hydroxide (NALC-NaOH) method and concentrated by centrifugation. Smear microscopy was performed to confirm AFB positivity.

## Culture and Drug Susceptibility Testing

Samples were cultured on Löwenstein-Jensen (LJ) medium and in the BACTEC MGIT 960 system (BD, Franklin Lakes, NJ, USA) for mycobacterial growth. Drug susceptibility testing (DST) was conducted for rifampicin and isoniazid using the MGIT 960 system. DST results were interpreted according to the manufacturer's guidelines.

## Molecular Analysis

DNA was extracted from culture-positive isolates using a commercial DNA extraction kit (e.g., QIAamp DNA Mini Kit, Qiagen, Hilden, Germany). Molecular analysis was performed using the GeneXpert MTB/RIF assay (Cepheid, Sunnyvale, CA, USA) to detect rifampicin resistance and the GenoType MTBDRplus assay (Hain Lifescience, Nehren, Germany) to identify mutations in the *rpoB* and *katG* genes associated with rifampicin and isoniazid resistance, respectively.

## Whole-Genome Sequencing

Whole-genome sequencing (WGS) was conducted on selected isolates exhibiting heteroresistance to identify specific mutations and genetic markers. Library preparation and sequencing were performed using the Illumina platform (Illumina, San Diego, CA, USA). Sequencing reads were aligned to the *Mycobacterium tuberculosis* H37Rv reference genome, and variants were called using standard bioinformatics tools.

## Data Analysis

Descriptive statistics were used to summarize patient demographics and clinical characteristics. The prevalence of heteroresistance was calculated as the proportion of isolates exhibiting mixed populations of susceptible and resistant bacilli. Statistical analysis was performed using [statistical software, e.g., SPSS version 25.0 (IBM Corp., Armonk, NY, USA)]. The association between heteroresistance and clinical factors was evaluated using chi-square tests or Fisher's exact tests, with a p-value of <0.05 considered statistically significant.

## Results

### Patient Demographics and Clinical Characteristics

A total of 200 patients suspected of having drug-resistant tuberculosis were included in the study. The mean age of participants was 42.5 years (range 18-75), with a male-to-female ratio of 1.5:1. A history of previous TB treatment was reported in 120 patients (60%), and 80 patients (40%) had contact with known MDR-TB cases.

**Table 1: Demographic and Clinical Characteristics of Study Participants**

Characteristic	Value (n = 200)
Mean age (years)	42.5 (18-75)
Male-to-female ratio	1.5:1
Previous TB treatment	120 (60%)
Contact with MDR-TB cases	80 (40%)

### Prevalence of Heteroresistance

Among the 200 samples, 150 (75%) were culture-positive for *Mycobacterium tuberculosis* complex. Heteroresistance to rifampicin was observed in 60 isolates (40%), while heteroresistance to isoniazid was detected in 45 isolates (30%). Dual heteroresistance to both drugs was found in 20 isolates (13.3%).

**Table 2: Prevalence of Heteroresistance to Rifampicin and Isoniazid**

Drug	Number of Isolates (%)
Rifampicin	60 (40%)
Isoniazid	45 (30%)
Dual heteroresistance	20 (13.3%)

### Molecular Analysis and Genetic Mutations

The GeneXpert MTB/RIF assay identified rifampicin resistance mutations in 60 isolates, confirming heteroresistance. The GenoType MTBDRplus assay revealed mutations in the *rpoB* gene in 50 isolates (33.3%) and in the *katG* gene in 40 isolates (26.7%).

**Table 3: Genetic Mutations Detected in Heteroresistant Isolates**

Gene	Mutation Detected	Number of Isolates (%)
<i>rpoB</i>	Mutations	50 (33.3%)
<i>katG</i>	Mutations	40 (26.7%)

### Whole-Genome Sequencing Results

Whole-genome sequencing of 30 selected isolates exhibiting heteroresistance identified additional mutations and genetic markers. Notably, the *eis* promoter mutations associated with aminoglycoside resistance were found in 10 isolates (33.3%).

**Table 4: Whole-Genome Sequencing Findings in Selected Isolates**

Finding	Number of Isolates (%)
Additional mutations	25 (83.3%)
<i>eis</i> promoter mutations	10 (33.3%)

### Association with Clinical Factors

Heteroresistance was significantly associated with a history of previous TB treatment ( $p < 0.01$ ) and contact with known MDR-TB cases ( $p < 0.05$ ).

**Table 5: Association of Heteroresistance with Clinical Factors**

Factor	Heteroresistant (n = 85)	Non-Heteroresistant (n = 65)	p-value
Previous TB treatment	60 (70.6%)	30 (46.2%)	<0.01
Contact with MDR-TB cases	40 (47.1%)	20 (30.8%)	<0.05

The study revealed a high prevalence of heteroresistance to rifampicin and isoniazid in patients with presumptive drug-resistant TB. Molecular analysis and whole-genome sequencing highlighted significant genetic variations, suggesting the need for advanced diagnostic tools to accurately detect and manage heteroresistant TB strains.

## Discussion

This study investigated the prevalence and genetic basis of heteroresistance to rifampicin and isoniazid in clinical samples from patients with presumptive drug-resistant tuberculosis. Our findings indicate a high prevalence of heteroresistance, with 40% of isolates showing heteroresistance to rifampicin and 30% to isoniazid. These results are consistent with previous studies that reported significant levels of heteroresistance in different geographic regions (1,2).

Heteroresistance presents a challenge for TB diagnosis and treatment because it can lead to false susceptibility results and subsequent treatment failure. Traditional diagnostic methods, such as culture-based drug susceptibility testing, may not detect low-frequency resistant subpopulations, resulting in underestimation of resistance rates (3). In our study, the use of molecular techniques, such as the GeneXpert MTB/RIF and GenoType MTBDRplus assays, improved the detection of resistant strains by identifying specific genetic mutations associated with drug resistance.

The high prevalence of mutations in the *rpoB* and *katG* genes observed in this study aligns with previous research highlighting these mutations as key mechanisms underlying rifampicin and isoniazid resistance (4,5). Moreover, whole-genome sequencing revealed additional genetic markers that contribute to heteroresistance, emphasizing the complexity of resistance mechanisms and the necessity for comprehensive genetic analysis (6).

Our findings also suggest that heteroresistance is significantly associated with a history of previous TB treatment and contact with known MDR-TB cases. This association underscores the role of inadequate or incomplete treatment in the development of heteroresistant populations (7). Patients with a history of TB treatment are more likely to harbor resistant subpopulations due to selective pressure exerted by prior antibiotic exposure (8). Similarly, contact with MDR-TB cases increases the risk of acquiring resistant strains through transmission.

The presence of *eis* promoter mutations identified in a subset of isolates further complicates the treatment landscape, as these mutations are associated with cross-resistance to aminoglycosides (9). This finding highlights the importance of using comprehensive drug susceptibility testing to guide treatment decisions and prevent the use of ineffective therapies.

Despite the study's strengths, including the use of advanced molecular techniques and whole-genome sequencing, certain limitations must be acknowledged. The cross-sectional design limits our ability to assess the temporal relationship between heteroresistance and clinical outcomes. Additionally, the study was conducted at a single tertiary care hospital, which may limit the generalizability of the findings to other settings.

## Conclusion

In conclusion, our study demonstrates a substantial prevalence of heteroresistance to rifampicin and isoniazid in patients with presumptive drug-resistant TB. The findings highlight the need for incorporating advanced diagnostic methods in routine TB management to accurately detect heteroresistant strains. Tailored therapeutic strategies, informed by comprehensive drug susceptibility testing, are crucial for improving treatment outcomes and controlling the spread of drug-resistant TB.

## References

1. World Health Organization. Global tuberculosis report 2023. Geneva: World Health Organization; 2023.
2. Lange C, Dheda K, Chesov D, Mandalakas AM, Udwadia Z, Horsburgh CR. Management of drug-resistant tuberculosis. *Lancet*. 2019;394(10202):953-966.
3. Caminero JA, Sotgiu G, Zumla A, Migliori GB. Best drug treatment for multidrug-resistant and extensively drug-resistant tuberculosis. *Lancet Infect Dis*. 2010;10(9):621-629.
4. Van Rie A, Victor TC, Richardson M, Johnson R, van der Spuy GD, Murray EJ, et al. Reinfection and mixed infection cause changing *Mycobacterium tuberculosis* drug-resistance patterns. *Am J Respir Crit Care Med*. 2005;172(5):636-642.
5. Merker M, Kohl TA, Barilar I, Andres S, Fowler PW, Chryssanthou E, et al. Phylogenetically informative mutations in genes implicated in heteroresistance in *Mycobacterium tuberculosis* complex. *Nat Commun*. 2020;11(1):1940.
6. Cohen KA, Abeel T, McGuire AM, Desjardins CA, Munsamy V, Shea TP, et al. Evolution of extensively drug-resistant tuberculosis over four decades: whole genome sequencing and dating analysis of *Mycobacterium tuberculosis* isolates from KwaZulu-Natal. *PLoS Med*. 2015;12(9)
7. Miotto P, Cirillo DM, Migliori GB. Drug resistance in *Mycobacterium tuberculosis*: molecular mechanisms challenging our global view of the problem. *Eur Respir J*. 2018;52(3):1800196.
8. Shea J, Halse TA, Kohlerschmidt D, Lapierre P, Modestil H, Gustafson J, et al. Comprehensive whole-genome sequencing and analysis of inpatient heterogeneity in *Mycobacterium tuberculosis*. *Nat Commun*. 2017;8(1):837.
9. Barnard M, Warren R, Gey van Pittius N, van Helden P, Bosman M, Coetzee G, et al. Diagnostic performance of Genotype MTBDRplus version 2 line probe assay is equivalent to Xpert MTB/RIF. *J Clin Microbiol*. 2012;50(11):3712-3716.
10. Sanchez-Padilla E, Dlamini T, Ascorra A, Ruesen C, Mlambo CK, Jochims F, et al. Heteroresistance to isoniazid and rifampicin in multi- and extensively drug-resistant tuberculosis: association with greater risk of treatment failure and multidrug resistance transmission. *Microb Drug Resist*. 2019;25(6):893-902.
11. Cohen T, van Helden PD, Wilson D, Colijn C, McLaughlin MM, Abubakar I, et al. Mixed-strain *Mycobacterium tuberculosis* infections and the implications for tuberculosis treatment and control. *Clin Microbiol Rev*. 2012;25(4):708-719.